1 2 3	English has problems throughout the manuscript. Major or minor errors in English throughout the document have not been highlighted. JPPRes accepts writing in American English or British English but not a mixture of them. Please attend to this.						
4 5	Please send figures with higher resolution in PNG or JPG format with the required size to be placed in the exact location. No super heavy images that make the size of the document huge.						
6 7 8	Hepatoprotective Study of Indonesian Water Kefir Against CCl4-Induced Liver Injury in Rats						
9	Running title: Hepatoprotective study of water kefir						
10	Widhya Aligita ^{1,3*} , Marlia Singgih ² , Entris Sutrisno ³ , I Ketut Adnyana ^{1*}						
11	Please check that ALL the authors' names are written as First Name followed by Last Name.						
12 13 14 15 16 17 18 19 20 21	¹ Department of Pharmacology & Clinical Pharmacy, School of Pharmacy, Bandung Institute of Technology, Bandung, Indonesia. ² Department of Pharmacochemistry, School of Pharmacy, Bandung Institute of Technology, Bandung, Indonesia. ³ Department of Pharmacology & Clinical Pharmacy, Faculty of Pharmacy, Bhakti Kencana University, Bandung, Indonesia. *E-mail address: widhya.aligita@bku.ac.id / ketut@itb.ac.id *Corresponding author address: Department of Pharmacology & Clinical Pharmacy, School of Pharmacy, Bandung Institute of Technology, Ganeca, 10, 40132, Bandung, Indonesia. Tel: +62-22-2504852						
22 23	The institutional e-mail of the ALL authors must be declared (and add rows according to the number of authors):						
	Author	Institutional e-mail	Other e-mail	ORCID			
		(manuatory)	(gmail, yahoo, etc.)	(0000-1111-2222-3333)			
	Widhya Aligita	widhya.aligita@bku.ac.id	w.aligita@gmail.com	0000-0001-8338-4115			
Marlia Singgih marlia@fa.itb.ac.id 0000-0002-535				0000-0002-5351-1731			

0000-0003-3830-6411 0000-0001-5217-2312

entris.sutrisno@bku.ac.id

ketut@itb.ac.id

24

Entris Sutrisno

I Ketut Adnyana

25 26 Contribution Details (to be ticked marked (X) as applicable and add columns according to the number of

authors):

27	Please check that the authors' names are written as Last Name followed by First Name initial(s). i.e., "Smith J"
28	instead "John Smith". Authors must have the same order as on the first page.

Contribution	Widhya Aligita	Marlia Singgih	Entris Sutrisno	I Ketut Adnyana
Concepts or Ideas	x	х	х	х
Design	х	х	х	х
Definition of intellectual content	х	х	х	х
Literature search	Х			х
Clinical trial				
Experimental studies	х			х
Data acquisition	х			х
Data analysis	х			х
Statistical analysis	х			х
Manuscript preparation	х	х	х	х
Manuscript editing	x	x	х	X
Manuscript review	x	x	X	X

29

30 ABSTRACT

- 31 32 33 Context: Water kefir is a fermented beverage that is typically made in the home by inoculating a sugar-rich solution with a microbial community (water kefir grains). Several studies on the metabolite content and hepatoprotective effects of water kefir have been published, but CCl4-induced acute liver injury has not been studied.
- 34 35 Objectives: The purpose of this study was to evaluate the efficacy of water kefir in vivo against hepatoprotective CCl4-induced acute liver injury and to in silico investigate metabolites that play an important role in hepatoprotective mechanisms.
- 36 37 38 Methods: Water kefir significantly and dose-dependently alleviated acute liver injury caused by carbon tetrachloride (CCl4). Water kefir administration at all doses produced results comparable to the positive control (curcuma extract). Furthermore, using
- molecular docking, the metabolites found in water kefir were evaluated for their role in the NF-KB and Nrf2 signaling pathways.
- 39 Results: Molecular docking simulations showed that the 25 metabolites tended to interact with the NF-KB receptor compared to
- 40 41 Nrf2. Fumaric acid was the strong metabolite that interacts with the NF-xB receptor with a free energy of binding and inhibition constant of -6.66 kcal/mol and 13.22 µM, respectively.
- 42 43 Conclusions: This study concludes that water kefir has hepatoprotective properties by decreasing inflammation and fibrosis levels.
- 44

45 Keywords: Hepatoprotective; Molecular docking; Metabolites; NF-KB; Nrf2; Water kefir

- 46 Please check if these keywords have been accepted by MeSH Browser 2022 or 2023 (https://meshb-
- 47 prev.nlm.nih.gov/). If not, type keywords accepted by this MeSH Browser.

48 **INTRODUCTION**

49 In most cases, water kefir is created by blending water kefir grains, sugar, and dried fruit 50 in a container. Water kefir's exact origins are unknown; however, two hypotheses have been 51 proposed regarding its history: the first suggests that water kefir grains were brought to 52 Europe from the Caucasus by soldiers returning from the Crimean War in the late nineteenth century (1); the second theory proposes that water kefir grains originated in Mexico from the 53 54 *Opuntia cactus* through natural processes (2). Sugary kefir grains, Balm of Gilead, African bees, 55 Japanese beer seeds, Ale nuts, and California bees are some other names for water kefir. Tibi 56 grains and ginger beer plant are other names for water kefir (2-4). Water kefir is appealing to 57 both consumers and researchers due to the variety of microbiota it contains, the fact that it is 58 an alternative to dairy products, the versatility with which it can be flavored, the fact that it is 59 low in calories and sugar, the ease with which it can be produced, and the health benefits it 60 offers.

61 Water kefir has been used medicinally for a very long time, and recent research has 62 indicated that it may have a variety of positive effects on people's health. It has been 63 demonstrated that water kefir contains non-pathogenic bacteria, in conjunction with the 64 production of organic acids, can inhibit the growth of pathogenic microbes, such as Shigella 65 sp., Salmonella sp., Staphylococcus aureus, and E. coli; and also, filamentous fungi such as Aspergillus ochraceus, A. niger, A. flavus, Penicillium sp., and Rhizopus sp. (add bibliography(ies)) 66 67 In addition to its antibacterial properties, water kefir possesses a broad spectrum of 68 pharmacological effects. Some of these therapeutic effects are anti-inflammatory (5,6), antioxidant (6-8), hepatoprotective (9,10), antihyperglycemic and antihyperlipidemic (11,12), 69 70 anti-edematous (13), antitumor (14), antihypertensive (15), immunomodulant (16), and anti-71 ulcerogenic (17). However, no studies have been reported on the hepatoprotective effects of 72 water kefir against carbon tetrachloride (CCl₄)-induced liver injury.

73 Studies have shown that acute liver injury is frequently accompanied by high levels of 74 oxidative stress and inflammatory responses (18). These findings have been found in several 75 studies. The most important signaling pathways that are involved in the regulation of 76 inflammation and antioxidation are the nuclear factor (NF-KB) and nuclear factor erythroid 2-77 related factor 2 (Nrf2) pathways, respectively. It has been shown that activating Nrf2 and 78 inhibiting NF-κB can reduce the amount of damage done to the liver. For instance, curcumin 79 protects against aflatoxin B1-induced liver injury by increasing the expression of Nrf2 and 80 related downstream antioxidant molecules (such as superoxide dismutase (SOD), catalase 81 (CAT), heme oxygenase 1 (HO-1), and NAD(P)H quinone dehydrogenase 1 (NQO-1)) (19). In 82 addition, methoxy eugenol, a molecule that is derived from nutmeg and Brazilian red propolis,

has been shown to exhibit hepatoprotective activity both *in vitro* and *in vivo*. This may be
attributed to the fact that it targets the NF-κB signaling pathway, which has been shown to
have anti-inflammatory effects (20).

86 Ethanol and lactic acid are the primary metabolites found in fermented water kefir, while 87 lower quantities of glycerol, mannitol, and acetic acid can also be found in the beverage. 88 Additionally, a variety of aromatic and volatile compounds are produced, including ethyl 89 acetate, isoamyl acetate, ethyl octanoate, ethyl hexanoate, and ethyl decanoate, among others 90 (compared to their threshold levels) (21). The chemical constituents of both phytochemicals 91 and secondary metabolites in natural products, including water kefir, are certainly capable of 92 providing various pharmacological effects for the body (22,23). However, an *in silico* study to 93 evaluate the metabolite content in water kefir has not been reported yet. Because of its capacity 94 to speed up the process of identifying and optimizing lead compounds, the in silico method 95 has become the front-runner in the race to improve the speed and accuracy of the process of 96 discovering new drugs. This is because the *in silico* method can identify and optimize lead 97 compounds more quickly. Techniques such as molecular docking and molecular dynamics 98 (MD) were able to directly indicate a small number of compounds that have high affinity and 99 selectivity by analyzing how the ligand and target interact with one another (24). Please, the 100 style for indicating bibliographic citations does not comply with JPPRes requirements, please 101 address this and improve it. 102 There are numerous studies demonstrating the hepatoprotective activity of kefir. Therefore,

the authors should state here the novel effect they intend to find, which has not been reported
 in the scientific literature. Please better state the research problem and how you intend to solve
 it.

106 Therefore, the purpose of this study was to evaluate the hepatoprotective effects of water 107 kefir in CCl₄-induced rats while also investigating the stability interactions of its metabolites 108 within the NF- κ B and Nrf2 receptors using molecular docking study. Our findings suggest 109 that water kefir and its the metabolites it produces could be a promising therapeutic agent for 110 the management of liver injury that is both safe and effective. Please move this last sentence 111 to the Conclusions, it is not valid for an Introduction.

112

113 MATERIAL AND METHODS

114 Materials and Reagents

Carbon tetrachloride, rats, diagnostic kits for alanine aminotransferase (ALT) (Proline, IFCC
 mod.), aspartate aminotransferase (AST) (Proline, IFCC mod.), Elisa Kit TNF-α (Bioassay
 Technology Laboratory), Elisa Kit TGF-β (Bioassay Technology Laboratory). Other chemicals

118 used in this study were of analytical reagent grade.

119 Experimental sample and reference extract

- 120 It is necessary to indicate here where these materials were obtained from. Indicate who
- 121 identified the material. The manuscript should include references to voucher specimens of the

122 plants (deposited in a major regional herbarium) or to the material examined, including their

123 registration number(s). It should be mentioned which parts of the plant have been used. The

124 GPS coordinates of the collection site of the species should also be indicated. It is mandatory

- 125 that the authors indicate how both the kefir and the turmeric extract were prepared, the quality
- 126 of these products, etc.

127 Animals and Experimental Design

128 Rats (Wistar strain, male, 200-250 g) were maintained on normal pellet food and tap water (ad 129 *libitum*?). Four mice in each group were used. All procedures relating to animals and their care conformed to the international guidelines Principles of Laboratory Animals Care (NIH 130 131 publication no. 85-23, revised 1985). In order to develop an animal model with liver injury, the 132 rats received CCl₄ (20% in olive oil) 1.25 mL/kg BW every 2 days via a gavage tube (25). The 133 rats were randomized into five groups after the development of animals with liver injury, 134 which is characterized by a significant increase in serum ALT level, as follows: (1) positive 135 control group, (2) Curcuma extract group 100 mg/kg BW, (3) water kefir 15 mL/kg BW group, 136 (4) water kefir 35 mL/kg BW group, and (5) water kefir 50 mL/kg BW group; with addition 137 (6) negative control group. Each group received group-specific treatment for two weeks, along with administration of 1.25 mL/kg BW of CCl₄ (20% in olive oil) every three days. Serum ALT 138 139 level, as the main parameter, was measured prior to induction, following induction, and 140 following treatment. Meanwhile, following therapy, serum AST, TNF- α , TGF- β levels, and 141 liver histopathological were evaluated. Please indicate how the serum was extracted in the 142 rats? from which part of the animal and how the blood was extracted. How were AST, TNF-143 α, TGF-a levels measured? give details of this. How was the liver extracted from the animals? 144 how was the liver histopathological analysis performed.

145 Only manuscripts of experiments conducted in accordance with the appropriate guidelines 146 will be eligible for publication. When working with experimental animals, reference must be 147 made to principles of laboratory animal care or similar regulations **and to approval by the** 148 **local ethical committee**. **The approval number and the corresponding date must be** 149 **provided**. It must clearly indicate that appropriate measures were taken to minimize pain or 150 discomfort, and details of animal care should be provided.

151 If the Curcuma (what curcuma species, scientific name, please?) extract was used as a 152 reference, the authors should justify here with bibliographic citation where this extract was 153 tested, at this declared dose, as hepatoprotective.

154

155 Molecular docking simulation

156 The PyRx software (26) was used to perform molecular docking experiments to anticipate 157 the manner of binding that happens between metabolites as small-molecule ligands and 158 biological macromolecules. The NCBI PubChem database (https://pubchem.nlm.nih.gov/, 159 accessed on 03 May 2023) was used to derive the three-dimensional structure of water kefir 160 metabolites (27). Target proteins like NF-κB (PDB code: 1A3Q) and Keap1 (PDB code: 4L7B) 161 were obtained from the RCSB Protein Data Bank (http://www.rcsb.org/, accessed on 03 May 162 2023). PyRx program was used to reduce protein, ligand converted to PDBQT (28), then 163 maximized GRID parameter (29) performed docking study (30). The BIOVIA Discovery Studio 164 2017 R2 program was used to view the protein and ligand complex and distance (31). The 165 BIOVIA Discovery Studio 2017 R2 tool was also utilized to find protein active sites.

166 Statistical Analysis

All of the information is displayed in the form of individual data points as well as the mean along with the standard error of the mean. The statistical analysis was carried out with the help of Minitab software (version 19.0), and to make comparisons between several different groups, a one-way analysis of variance with Dunnett's post hoc test was utilized. All statistical 171 graphs were created with Microsoft Excel 2019 in their respective versions. The levels of 172 significance that were considered to have been reached were * p < 0.05.

173 **RESULTS**

174 In vivo evaluation of hepatoprotective activity

175 The serum ALT levels, as the main parameter for the liver damage, were measured prior to 176 and following induction, as well as following treatment (Fig. 1). Meanwhile, after treatment AST, TNF-α, and TGF-β levels were also evaluated. These findings were analyzed using a one-177 178 way ANOVA with a 95% confidence level and a two-sided confidence interval. A significant 179 rise in blood ALT levels demonstrated the establishment of an animal model with liver injury, 180 according to statistical analysis, following the administration of CCl₄. When compared to the 181 positive control group, ALT serum levels reduced significantly after two weeks of therapy with curcuma extract or water kefir. The three doses of water kefir groups demonstrated 182 183 equivalent activity when curcuma extract was used as the standard treatment and there was 184 no significant difference between the three doses of water kefir. When compared to the 185 positive control group, AST levels were also reduced dramatically following treatment with 186 curcuma extract or water kefir. TNF-a levels in the water kefir group were significantly lower 187 than in the positive control group at dosages of 35 and 50 mL/kg body weight. Even though there was no statistically significant difference in TGF-β levels, the group that received the 188 189 treatment demonstrated a decrease in TGF-B levels.



Fig. 1. (a) Before induction, after induction, and after treatment serum ALT level; (b) After treatment serum AST level; (c) After treatment serum TNF- α level; (d) After treatment serum TGF- β level. * indicates a significantly different result when compared to the positive control group; # indicates a significantly different result when compared to the curcuma extract group; p < 0.05; n = 4 mice in each group.

190

A histological examination of a normal liver group revealed a typical central vein bordered with endothelial cells and normal hepatocytes, resulting in hepatic cords of cells with distinct cell borders and sinusoidal gaps (Fig 2A & 2B). The CCl4-induced group developed centrilobular hepatic necrosis level 4, which was characterized by massive vacuolization and necrosis of epithelial cells in the liver's centrilobular region (blue arrow) (Fig 2C & 2D). The

- 196 group that received either curcuma extract or water kefir treatment improved in varied
- 197 necrotic conditions ranging from level 1 (water kefir 50 mL/ kg BW) (Fig 2K & 2L) to level 2
- 198 (curcuma extract (Fig 2E & 2F), water kefir 15 mL/kg BW (Fig 2G & 2H), water kefir 30 mL/kg
- 199 BW (Fig 2I & 2J)).



Fig. 2. Liver histology after CCl4 intoxication and treatment with HE staining. Negative control group (A & B), positive control group (C & D), curcuma extract group (E & F), water kefir 15 mL/kg BW (G & H), water kefir 30 mL/kg BW (I & J), and water kefir 50 mL/kg BW (K & L). 1: central vein; 2: normal liver epithelial cell; 3: portal track; blue arrow: necrosis and liver epithelial cell vacuolization at centrolobular region.

201 Molecular docking

Molecular docking studies are considered a powerful tool for predicting the potential targets of bioactive molecules. In order to carry out molecular docking simulations, one of the most critical steps is to identify the target active site. If the target protein is crystallized with a 205 native ligand, in many instances, the location of the active site can be established without any 206 difficulty (32). However, the NF-kB (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) proteins 207 do not have a native ligand, so the active site was determined. Active site prediction in docking 208 is a computational method for predicting the location and orientation of a receptor protein's 209 binding site for a ligand molecule. The active site prediction is based on a protein structural 210 analysis and the identification of amino acid residues that are likely to interact with the ligand. 211 The projected binding site is then utilized as a starting point for molecular docking, a computer 212 method for predicting a ligand molecule's binding affinity and orientation to a receptor 213 protein. The active site prediction for target proteins (Keap1 and NF-kB) gives the grid box 214 coordinates (x y z) of 17.500880, 62.323000, and 0.973748; and -1.751769, -20.743853, and -215 29.010438, respectively (Fig. 3).



Fig. 3. Binding pocket (red color) generated using the BIOVIA Discovery Studio 2017 tool, coupled with the sequence containing the highlighted residues that constitute the binding pocket. A) NF-kB (PDB ID 1A3Q) B) Keap1 (PDB ID 4L7B).

216

217 The docking results of the 25 metabolites could interact with target proteins (Keap1 and 218 NF-κB) (Table 1). In general, all metabolites were able to form interactions with both NF-κB receptors (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) (Table 1). For volatile compounds, 2-219 220 phenyl ethanol and benzaldehyde interact most strongly with the NF-KB receptor (PDB ID 221 1A3Q) with almost the same binding energy of -4.19 kcal/mol. As for organic acids, succinic 222 acid, fumaric acid, and citric acid provide nearly the same strong interactions. Bond energy 223 values of succinic acid, fumaric acid, and citric acid of -6.24, -6.66, and -6.25 kcal/mol, 224 respectively. In sugars, glucose provides the strongest interaction with the NF-kB receptor 225 (PDB ID 1A3Q) where the binding energy was -3.71 kcal/mol. As for the nrf2 Keap1 receptor 226 (PDB ID 4L7B), 2-phenylacetaldehyde from volatile compounds was able to interact most 227 strongly with the Keap1 receptor (PDB ID 4L7B) with a binding energy of -3.46 kcal/mol. As 228 for organic acids, succinic acid provides nearly the same strong interaction with a bond energy 229 value of -3.02 kcal/mol, respectively. In sugars, glucose provides the strongest interaction with 230 the Keap1 receptor (PDB ID 4L7B) where the binding energy was -3.09 kcal/mol.

231Table 1. The free energy of binding and inhibition constant of water kefir metabolite active site interaction on NF-232κB (PDB ID 1A3Q) and Keap1 (PDB ID 4L7B)

No.	Metabolites	PDB: 1A3Q		PDB: 4L7B	
		Free Energy of	Inhibition	Free Energy	Inhibition
		Binding, ∆G	Constant, Ki	of Binding,	Constant, Ki
		(kcal/mol)	(µM)	ΔG	(µM)
				(kcal/mol)	

	Volatile compounds				
1	2-Methylbutanol	-3.31	3,770	-2.68	10,890
2	3-Methylbutanol	-3.22	4,360	-2.50	14,670
3	2-Phenylethanol	-4.19	851.52	-3.12	5,140
4	Ethyl acetate	-3.19	4,590	-2.43	16,670
5	2-Methylbutanal	-3.26	4,050	-2.35	19,050
6	1-Octanol	-2.65	11,490	-2.11	28,220
7	2-Phenylethyl acetate	-3.99	1,190	-3.33	3,590
8	Benzyl acetate	-4.16	887.66	-3.44	3,020
9	Benzaldehyde	-4.19	853.74	-3.19	4,550
10	Furfuryl acetate	-3.60	2,320	-3.32	3,660
11	Isobutyl acetate	-3.19	4,610	-2.94	7,020
12	Isopentyl acetate	-3.20	4,510	-3.05	5,820
13	2-Phenylacetaldehyde	-4.07	1,030	-3.46	2,930
14	3-Methylbutanal	-3.16	4,850	-2.73	10,020
15	Gluconic acid	-3.11	5,280	-1.63	64,290
	Organic acids				
16	Lactic acid	-4.92	249.28	-2.06	30,680
17	Succinic acid	-6.24	26.68	-3.02	6,140
18	Fumaric acid	-6.66	13.22	-2.45	16,100
19	Citric acid	-6.25	26.18	-1.87	42,440
20	Malic acid	-5.65	72.69	-2.53	13,860
21	Acetic acid	-4.99	219.33	-2.44	16,280
22	Ethanol	-2.39	17,800	-2.08	29,800
	Sugars				
23	Sucrose	-1.40	93,710	-1.33	105,320
24	Glucose	-3.71	1,900	-3.09	5,440
25	Fructose	-3.21	4,400	-2.65	11,440
	Please, it is mandatory to add a reference NF- kB inhibitor/activator.				

234 The theoretical binding modes of the top three metabolites with their target proteins (Keap1 235 and NF-KB) were shown in Fig. 3 and 4, respectively. The molecular docking results suggested 236 that these metabolites anchor in Keap1 and NF-KB to form a complex through hydrogen bonds 237 with various residues. The interaction of 2-phenyl ethanol with the active site of NF-KB was 238 formed by two hydrogen bonds and one Pi-Alkyl interaction with amino acid residues of 239 PRO223 and LYS252, and LYS252, respectively. The interaction of benzaldehyde with the 240 active site of NF-KB was formed by a hydrogen bond with the amino acid residue of LYS252. 241 The interaction of fumaric acid with the active site of NF-KB was formed by six hydrogen 242 bonds with the amino acid residues of LYS221, LYS252, and TYR285. The interaction of glucose 243 with the active site of NF-KB was formed by six hydrogen bonds with the amino acid residues 244 SER220, LYS221, SER226, and LYS252. Although glucose and fumaric acid were able to be 245 formed by six hydrogen bonds, different types of amino acid residues were involved in the 246 interaction so that fumaric acid interacted more strongly with the active site of NF-KB (Fig. 4).

247 Comment about reference compound and compare.



Fig. 4. 2D and 3D representation of the kind of interaction on the formation of the NF-κB active site (PDB ID 1A3Q). A) 2-Phenylethanol, B) Benzaldehyde, C) Fumaric acid, and D) Glucose. Add a reference compound

249 The interaction of 2-phenylacetaldehyde with the active site of Keap1 was formed by one 250 hydrogen bond and one Pi-Alkyl interaction with amino acid residues of GLN530 and 251 ALA556, respectively. The interaction of succinic acid with the active site of Keap1 was formed 252 by four hydrogen bonds with the amino acid residues of ARG336, HIS575, and PHE577. The 253 interaction of glucose with the active site of Keap1 was formed by six hydrogen bonds with the amino acid residues LYS551, ARG553, and ASP573. Although glucose can form more 254 255 hydrogen bonds than 2-phenylacetaldehyde, the different types of amino acid residues 256 involved have not been able to have a significant effect on the binding energy of its interaction 257 with the active site of Keap1 (Fig. 5).







260 Overall, the metabolites of water kefir prefer to interact with NF-κB and nrf2 Keap1 261 receptors. Where, fumaric acid and 2-phenylacetaldehyde are metabolites that have the 262 strongest interaction with NF-κB (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) receptors, 263 respectively.

264 **DISCUSSION**

265 Increased liver enzyme production is one of the abnormalities indicating liver damage. This increase in secretion increases the leakage of enzymes into the circulation. Transaminase is a 266 267 frequent enzyme used to evaluate liver disorders. The transaminases AST and ALT catalyze 268 the transfer of the α -amino group from alanine and aspartic acid to α -ketoglutaric acid. AST 269 is found in the liver, heart, and skeletal muscle, as well as the kidney, brain, pancreas, lung, 270 leukocytes, and erythrocytes. While ALT is dominantly found in the liver, although it is also 271 found in low concentrations in other tissues (33). Consequently, ALT was used as the principal 272 hepatotoxicity criterion in this study.

273 Carbon tetrachloride (CCl4) is a component of the hepatotoxin, which acts after metabolic 274 activation and is extensively employed as a liver-damaging agent. And in this study, the 275 administration of CCl4 to animals caused hepatotoxicity, which was characterized by a 276 significant increase in ALT after several administration. CCl4 is metabolized by the enzyme 277 cytochrome p450 (CYP2E1) in the endoplasmic reticulum, to a highly reactive trichloromethyl 278 radical. The trichloromethyl radical rapidly reacts with oxygen to form the trichloromethyl 279 peroxyl radical, which rapidly reacts with lipids to form lipid peroxidation products. Free 280 radicals will disrupt the endoplasmic reticulum (ER) and lead to lipid accumulation, decreased protein synthesis, and increased oxidase activity. CCl4 hepatotoxicity is characterized by hepatocellular necrosis with fat deposition (34). At the molecular level, administration of CCl4 can activate tumor necrosis factor (TNF)- α , nitric oxide (NO), and transforming growth factor (TGF)- α and - β in cells, processes that precipitate cell self-destruction or fibrosis. TNF- α leads to apoptosis, whereas TGF- β leads to fibrosis (35).

286 In terms of its pathophysiological underpinnings, liver illness is linked to a condition known 287 as dysbiosis, which refers to an imbalance in the make-up of the gut microbiota (36–38). Both 288 qualitative and quantitative changes in the gut microbiome have the potential to affect the 289 composition of products produced by the microbiota, such as short-chain fatty acids and bile 290 acids (36). Qualitative changes include an imbalance between harmful and helpful 291 microbiomes, whereas numeric changes involve changes to the overall microbiota. In addition 292 to these symptoms, intestinal inflammation, disruption of the intestinal barrier, and the 293 transfer of microbial products can all be caused by dysbiosis (39). For this reason, the condition 294 of the gut microbiome is an important factor in the initiation and development of chronic liver 295 disease (40). Based on the results of the study, treatment with water kefir for 2 weeks after the 296 occurrence of liver damage was able to improve the overall condition of the liver which was 297 marked by a significant decrease in the values of AST, ALT, TNF- α , TGF- β , and significant 298 improvement in liver histology.

299 Water kefir contains a number of microorganisms that have been linked to health benefits, such as the probiotics *L. paracasei* and *B. cereus* (41). In several animal models of liver injury, *L.* 300 301 paracasei was able to ameliorate liver abnormalities. Increased levels of antioxidants like 302 glutathione and catalase are linked to this activity, as is the downregulation of pro-303 inflammatory transcription factors such as nuclear B-factor, lipopolysaccharide, and Toll-like 304 receptor 4. Improvements in intestinal barrier function and histological integrity were also 305 observed. Increased expression of claudin-1 and occludin-1, two components of tight junctions, occurs simultaneously with the restoration of the p38 MAPK pathway (41-44). 306 307 Bacillus is a kind of endospore-forming bacterium that can endure extremely cold temperatures and lengthy periods of storage without dying; its spores can even tolerate the 308 309 acidic environment of the stomach and make it all the way to the small intestine (45). *B. cereus* 310 has been shown to reduce ALT levels, an indicator of liver healing, in various animal models 311 of liver injury. It protects the liver by reducing inflammation, enhancing the gut flora, and 312 strengthening the tight junctions in the intestines (46–48). In addition, hepatocyte necrosis and 313 serum levels of ALT, ZO-1, AST, and TAC were significantly reduced after pretreatment with 314 *Bacillus spores*. This effect is comparable to that of the popular hepatoprotective compound 315 silymarin (49).

316 Beer, vinegar, cider, and cocoa bean masses are just some of the places where Gram-317 negative acetic acid bacteria have been found. One of the features that sets acetic acid bacteria apart from others is their alkaline-stable lipid membrane (50). Their "oxidative" fermentation 318 319 metabolism is responsible for the principal metabolic process in these bacteria, the oxidation 320 of ethanol to acetic acid. Important in the food and beverage sector and beyond, fermentation 321 helps mediate the transition of diverse substrates into products. Although lactic acid bacteria 322 have been studied more extensively than acetic acid bacteria (51,52), various studies have 323 shown promising results concerning the pharmacological effects of acetic acid bacteria, 324 especially as a hepatoprotective agent. Acetic acid, which is the main metabolite of acetic acid bacteria, can lower inflammation and the severity of liver injury in rats with septic shock by 325 326 increasing the expression of TRIM40. TRIM40 has been shown to minimize liver damage, 327 decrease the synthesis and release of cytokines such as IL-6 and TNF-a, raise the expression of 328 IL-10, improve survival in septic mice, and block the activity of the TLR4 signaling pathway. Acetic acid injection decreased inflammation as well as the production of inflammatory cytokines (53). Acetic acid also stimulates the AMPK signaling pathway, which leads to enhanced lipid oxidation and reduced hepatic lipid and body fat deposition (54,55).

332 Apart from microorganisms that directly provide hepatoprotective effects, the metabolites 333 produced from these microorganisms also have the potential as hepatoprotective. Molecular 334 docking is a technique that is utilized in the context of NF-kB and Nrf2 to make predictions 335 regarding the binding affinity and orientation of small-molecule inhibitors to their active site. 336 The transcription factor known as NF-KB is an essential component in the management of both 337 the immune system and the inflammatory response (56). The expression of important 338 inflammatory genes can be inhibited by small-molecule inhibitors that impair the interaction 339 between NF-KB and DNA. These inhibitors have the potential to be used in therapeutic 340 applications. The Nrf2 binding site is Kelch-like ECH-associated protein 1 in the context of 341 nuclear factor erythroid 2-related factor 2, also known as Nrf2 (Keap1) (57,58). Small-molecule 342 inhibitors that disrupt the link between Keap1 and Nrf2 can activate the Nrf2-ARE signaling 343 pathway, which has been demonstrated to have cytoprotective effects. Keap1 is a negative 344 regulator of Nrf2, and this association can be disrupted by small-molecule inhibitors (59).

345 Binding energy and Ki are important parameters used in molecular docking to evaluate the 346 strength of the interaction between a ligand and a receptor protein. Binding energy is the 347 energy released when a ligand binds to a receptor protein, and it is calculated as the difference 348 between the energy of the bound complex and the energy of the unbound ligand and protein 349 (bibliography). Ki, on the other hand, is the dissociation constant of the ligand-receptor 350 complex, which represents the concentration of the ligand required to occupy 50% of the 351 receptor binding sites. Both binding energy and Ki are used to predict the binding affinity and 352 selectivity of a ligand to a receptor protein (bibliography). Molecular docking studies often 353 report the binding energy and Ki values of the docked ligand-receptor complex to evaluate 354 the strength of the interaction and to compare the binding affinity of different ligands 355 (bibliography).

356 Based on the results of an in silico study of water kefir metabolite compounds, it was known that fumaric acid and 2-phenylacetaldehyde have strongest interaction with NF-KB (PDB ID 357 358 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) receptors, respectively (bibliography). Fumaric acid has 359 been studied for its potential as a hepatoprotection. Fumaric acid protected rat livers against 360 cadmium toxicity by decreasing oxidative stress and improving hepatic serum. Fumaric acid 361 prevented cadmium-accrued hepatocellular damage and showed the potential to avert hepatic 362 injury against cadmium in rats (60). Fumaric acid esters were found to ameliorate inflammation and oxidative stress in rats, which was associated with less adiposity and ectopic 363 364 fat accumulation (61).

365 NF- κ B and Nrf2 are two transcription factors that play important roles in regulating 366 inflammation and cell survival. While NF-KB is involved in the inflammatory response, Nrf2 367 is involved in the antioxidant response (62,63). Both transcription factors have been 368 investigated as potential targets for the development of hepatoprotective agents (62,64–66). 369 There is evidence of crosstalk between the Nrf2 and NF-kB pathways (63,66). The Nrf2 370 pathway inhibits the activation of the NF-KB pathway by increasing antioxidant defenses and 371 HO-1 expression, which efficiently neutralizes ROS and detoxifies (63). The crosstalk between Nrf2 and NF- κ B could be a new therapeutic target against hepatotoxicity (66). Researchers 372 373 have tried to identify molecule activators of Nrf2 as chemoprevention ROS-dependent 374 carcinogenesis, while others have focused on identifying Nrf2 inhibitors to increase sensitivity 375 of cancer cells to chemotherapy (67). While NF- κ B and Nrf2 are involved in different cellular 376 processes, they have both been investigated as potential targets for the development of 377 hepatoprotective agents. Molecular docking studies have been used to investigate the 378 interaction of potential hepatoprotective agents with these transcription factors. There is also 379 evidence of crosstalk between the Nrf2 and NF-κB pathways, which could be a new 380 therapeutic target against hepatotoxicity.

381 In the pathogenesis of hepatocellular damage, liver fibrosis, and HCC, the NF-KB plays a 382 crucial role in regulating inflammation and cell death (bibliography). In response to many stimuli that may pose a threat to the host, NF-KB is activated, setting in motion processes such 383 384 as inflammation, immunity, wound healing, and pathogen clearance (bibliography). 385 Pathogen-derived chemicals that activate Toll-Like Receptors (TLRs) include 386 lipopolysaccharide (LPS), viral and bacterial DNA and RNA, and inflammatory cytokines 387 including tumor necrosis factor (TNF) and interleukins (IL)-1 (bibliography). Numerous genes 388 containing kB-binding sites are transcribed in response to NF-kB activation; these genes play 389 important roles in regulating inflammation, the immunological response, and cell survival. In 390 an NF-kB-dependent manner, lipopolysaccharide (LPS) activates TLR4 on dormant HSCs by 391 reducing BAMBI expression (an inhibitory TGF-β pseudoreceptor) and increasing Kupffer cell 392 chemotaxis. Due to low levels of BAMBI, recruited Kupffer cells secrete TGF-B, which 393 stimulates HSCs unrestrictively. When HSCs have been activated, NF-KB serves a second 394 crucial function by increasing their chances of survival. LPS, Kupffer cell-derived mediators 395 (such as IL-1 and TNF), and angiotensin II, which is produced and acts on HSCs in an autocrine 396 way, all play roles in activating NF-KB in activated hepatic stellate cells. More activated HSCs 397 and extracellular matrix are deposited in the liver as a result of greater HSC activation and 398 survival (68).

TNF-α and TGF-β can stimulate one another's production, and TNF-α has been shown to influence TGF-β expression in a variety of cells and tissues (69). TNF-α is an inflammatory cytokine that contributes to liver inflammation, and chronic liver inflammation results in liver fibrosis. TNF-α exerts its effects on liver fibrosis via multiple mechanisms, including TGF-β signaling activation (70). Targeting TNF-α and TGF-β signaling pathways may, therefore, have therapeutic potential for treating liver diseases. In regards to hepatoprotective effects, the relationship between TGF-β and TNF-α is complex and not completely understood.

406 CONCLUSION

407 We evaluated the hepatoprotective effect of Indonesian water kefir on CCl4-induced liver-408 damaged rats. Water kefir administration improved the condition of liver damage, 409 characterized by a decrease in serum levels of AST, ALT, TNF, TGF, and an improvement of 410 liver tissue profile. *In silico* evaluation showed that the metabolites in water kefir were able to 411 interact with target proteins in the NF-κB and Nrf2 pathways. It was concluded that water 412 kefir improves the condition of the liver by reducing the level of necrosis and fibrosis.

- 413 CONFLICT OF INTEREST
- 414 The authors declare no conflict of interest.

415 ACKNOWLEDGMENT

This research was funded by the Center of Research and Community Service, Bhakti Kencana University,
 Bandung, West Java, Republic of Indonesia. (please add grant number)

418	RE	FERENCES
419	Thi	is section is one of the most important sections of the document. Please pay close attention
420	to i	t:
421		 The style of the References does not meet the requirements of JPPRes.
422		 Please write the References according to the Instructions to the Author or the
423		examples:
424		 <u>https://jppres.com/jppres/archive/</u>
425		 <u>https://jppres.com/jppres/volume-11-issue-2/</u>
426		• Also, all authors and DOIs must be given.
427		• Please check that all references in this section have been cited in the text and vice
428		versa.
429		• Please check that the journals cited are prestigious in the discipline and are not
430		predatory journals (<u>https://www.openacessjournal.com/blog/predatory-journals-</u>
431		<u>list/#I_%E2%80%93_predatory_journals</u>). References must be written free of codes
432		from any bibliography processor, such as Endnotes, Mendeley, Zotero, etc
433 434	1.	Ward M. V. The ginger-beer plant, and the organisms composing it: a contribution to the study of fermentation-yeasts and bacteria. Phil Trans R Soc Lond B. 1892 Dec 31;183:125–97.
435 436	2.	Moinas M, Horisberger M, Bauer H. The structural organization of the Tibi grain as revealed by light, scanning and transmission microscopy. Arch Microbiol. 1980 Dec;128(2):157–61.
437 438 439	3.	Kebler LF. "California Bees."11The author stated that the work on this subject is the result of investigation for the Post a c e Department relative to improper use of the mails in connection with this product. The Journal of the American Pharmaceutical Association (1912). 1921 Dec;10(12):939–43.
440	4.	Pidoux M. Kefir Grain Flora. Mircen Journal. 1989;5:223-8.
441 442 443	5.	Diniz RO, Garla LK, Schneedorf JM, Carvalho JCT. Study of anti-inflammatory activity of Tibetan mushroom, a symbiotic culture of bacteria and fungi encapsulated into a polysaccharide matrix. Pharmacological Research. 2003 Jan;47(1):49–52.
444 445 446	6.	Aligita W, Tarigan PN, Susilawati E. ANTI INFLAMMATORY AND ANTIOXIDANT ACTIVITY OF KEFIR WATER. IJBPAS [Internet]. 2020 Jan 1 [cited 2022 Nov 10];9(1). Available from: https://ijbpas.com/pdf/2020/January/MS_IJBPAS_2019_4904.pdf
447 448	7.	Alsayadi M, Aljawfi Y, Belarbi M, Sabri FZ. ANTIOXIDANT POTENCY OF WATER KEFIR. Journal of Microbiology, Biotechnology and Food Sciences. 2013;2(6):2444–7.
449 450 451	8.	Darvishzadeh P, Orsat V, Martinez JL. Process Optimization for Development of a Novel Water Kefir Drink with High Antioxidant Activity and Potential Probiotic Properties from Russian Olive Fruit (Elaeagnus angustifolia). Food Bioprocess Technol. 2021 Feb;14(2):248–60.
452 453 454	9.	Aspiras BEE, Flores RFAC, Pareja MC. Hepatoprotective effect of Fermented Water Kefir on Sprague- Dawley rats (Rattus norvegicus) induced with sublethal dose of Acetaminophen. INT J CURR SCI. 2015;17(E):18–28.
455 456 457	10.	Aligita W, Alex V, Taaraungan S, Susilawati E. HEPATOPROTECTIVE ACTIVITY OF WATER KEFIR. IJBPAS [Internet]. 2021 Jun 1 [cited 2022 Nov 10];10(6). Available from: https://ijbpas.com/pdf/2021/June/MS_IJBPAS_2021_5493.pdf
458 459 460	11.	Alsayadi M, Jawfi YA, Belarbi M, Soualem-Mami Z, Merzouk H, Sari DC, et al. Evaluation of Anti- Hyperglycemic and Anti-Hyperlipidemic Activities of Water Kefir as Probiotic on Streptozotocin-Induced Diabetic Wistar Rats. JDM. 2014;04(02):85–95.
461 462	12.	Rocha-Gomes A, Escobar A, Soares JS, Silva AA da, Dessimoni-Pinto NAV, Riul TR. Chemical composition and hypocholesterolemic effect of milk kefir and water kefir in Wistar rats. Rev Nutr. 2018 Mar;31(2):137–45.

- 463
 464
 464
 465
 465
 13. Moreira MEC, Santos MHD, Zolini GPP, Wouters ATB, Carvalho JCT, Schneedorf JM. Anti-Inflammatory and Cicatrizing Activities of a Carbohydrate Fraction Isolated from Sugary Kefir. Journal of Medicinal Food. 2008 Jun;11(2):356-61.
- 466
 467
 468
 468
 14. Zamberi NR, Abu N, Mohamed NE, Nordin N, Keong YS, Beh BK, et al. The Antimetastatic and Antiangiogenesis Effects of Kefir Water on Murine Breast Cancer Cells. Integr Cancer Ther. 2016 Dec;15(4):NP53-66.
- 469
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
 470
- 471 16. Calatayud M, Börner RA, Ghyselinck J, Verstrepen L, Medts JD, Abbeele PV den, et al. Water Kefir and
 472 Derived Pasteurized Beverages Modulate Gut Microbiota, Intestinal Permeability and Cytokine Production
 473 In Vitro. Nutrients. 2021 Oct 29;13(11):3897.
- 474
 475
 476
 17. Rodrigues KL, Araújo TH, Schneedorf JM, Ferreira C de S, Moraes G de OI, Coimbra RS, et al. A novel beer fermented by kefir enhances anti-inflammatory and anti-ulcerogenic activities found isolated in its constituents. Journal of Functional Foods. 2016 Mar;21:58–69.
- 477 18. Dai C, Li H, Wang Y, Tang S, Velkov T, Shen J. Inhibition of Oxidative Stress and ALOX12 and NF-κB
 478 Pathways Contribute to the Protective Effect of Baicalein on Carbon Tetrachloride-Induced Acute Liver
 479 Injury. Antioxidants. 2021 Jun 18;10(6):976.
- 480
 481
 481
 482
 482
 483
 484
 484
 484
 484
 485
 485
 486
 486
 487
 487
 487
 488
 488
 488
 489
 480
 480
 480
 481
 481
 481
 481
 481
 482
 482
 482
 482
 483
 484
 484
 484
 484
 484
 485
 485
 486
 487
 487
 487
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
 488
- 483 20. De Souza Basso B, Haute GV, Ortega-Ribera M, Luft C, Antunes GL, Bastos MS, et al. Methoxyeugenol
 484 deactivates hepatic stellate cells and attenuates liver fibrosis and inflammation through a PPAR-γ and NF485 kB mechanism. Journal of Ethnopharmacology. 2021 Nov;280:114433.
- 486
 487
 21. Laureys D, De Vuyst L. Water kefir as a promising low-sugar probiotic fermented beverage. Arch Public Health. 2014 Jun;72(S1):P1, 2049-3258-72-S1-P1.
- 488
 489
 489
 490
 42. A. Asnawi, Aman LO, Nursamsiar, A. Yuliantini, E. Febrina. MOLECULAR DOCKING AND MOLECULAR
 490
 490
 490
 491
 491
 492
 493
 494
 494
 494
 494
 494
 494
 495
 495
 496
 496
 497
 498
 498
 498
 499
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
 490
- 491 23. Nursamsiar, Nur S, Febrina E, Asnawi A, Syafiie S. Synthesis and Inhibitory Activity of Curculigoside A
 492 Derivatives as Potential Anti-Diabetic Agents with β-Cell Apoptosis. Journal of Molecular Structure. 2022
 493 Oct;1265:133292.
- 494
 495
 496
 24. Febrina E, Alamhari RK, Abdulah R, Lestari K, Levita J, Supratman U. MOLECULAR DOCKING AND MOLECULAR DYNAMICS STUDIES OF ACALYPHA INDICA L. PHYTOCHEMICAL CONSTITUENTS WITH CASPASE-3. Int J App Pharm. 2021 Dec 11;210–5.
- 497 25. Yeh YH, Hsieh YL, Lee YT, Hsieh CH. Protective effects of cholestin against carbon tetrachloride-induced
 498 hepatotoxicity in rats. e-SPEN, the European e-Journal of Clinical Nutrition and Metabolism. 2011
 499 Dec;6(6):e264-71.
- 500 26. Dallakyan S, Olson AJ. Small-Molecule Library Screening by Docking with PyRx. In: Hempel JE, Williams
 501 CH, Hong CC, editors. Chemical Biology [Internet]. New York, NY: Springer New York; 2015 [cited 2023 Jun
 502 25]. p. 243-50. (Methods in Molecular Biology; vol. 1263). Available from: http://link.springer.com/10.1007/978-1-4939-2269-7_19
- Patel SH, Tan JP, Börner RA, Zhang SJ, Priour S, Lima A, et al. A temporal view of the water kefir microbiota and flavour attributes. Innovative Food Science & Emerging Technologies. 2022 Aug;80:103084.
- S06 28. Asnawi A, Nedja M, Febrina E, Purwaniati. Prediction of a Stable Complex of Compounds in the Ethanol
 S07 Extract of Celery Leaves (Apium graveolens L.) Function as a VKORC1 Antagonist. TJNPR. 2023 Feb
 S08 28;7(2):2362–70.

- Solation 29. Asnawi A, Febrina E, Aligita W, Kurnia D, Aman LO, Yuliantini A. SCREENING OF ASHITABA
 (ANGELICA KEISKEI K.) COMPOUNDS AS POTENTIAL MYCOBACTERIUM TUBERCULOSIS KASA
 INHIBITORS. Int J App Pharm. 2022 Dec 27;80–5.
- 512 30. Febrina E, Asnawi A, Abdulah R, Lestari K, Supratman U. IDENTIFICATION OF FLAVONOIDS FROM
 513 ACALYPHA INDICA L. (EUPHORBIACEAE) AS CASPASE-3 ACTIVATORS USING MOLECULAR
 514 DOCKING AND MOLECULAR DYNAMICS. Int J App Pharm. 2022 Dec 27;162–6.
- 515 31. Ischak NI, Aman LO, Hasan H, Kilo AL, Asnawi A. In silico screening of Andrographis paniculata
 516 secondary metabolites as anti-diabetes mellitus through PDE9 inhibition. Res Pharm Sci. 2023 Feb;18(1):100–
 517 11.
- 518 32. Li Z, Huang Y, Wu Y, Chen J, Wu D, Zhan CG, et al. Absolute Binding Free Energy Calculation and Design of a Subnanomolar Inhibitor of Phosphodiesterase-10. J Med Chem. 2019 Feb 28;62(4):2099-111.
- 520 33. Lee TH, Kim WR, Poterucha JJ. Evaluation of Elevated Liver Enzymes. Clinics in Liver Disease. 2012
 521 May;16(2):183–98.
- S22
 S4. Ritesh KR, Suganya A, Dileepkumar HV, Rajashekar Y, Shivanandappa T. A single acute hepatotoxic dose of CCl 4 causes oxidative stress in the rat brain. Toxicology Reports. 2015;2:891–5.
- Weber LWD, Boll M, Stampfl A. Hepatotoxicity and Mechanism of Action of Haloalkanes: Carbon
 Tetrachloride as a Toxicological Model. Critical Reviews in Toxicology. 2003 Jan;33(2):105–36.
- S26 36. Romero-Luna HE, Peredo-Lovillo A, Hernández-Mendoza A, Hernández-Sánchez H, Cauich-Sánchez PI,
 S27 Ribas-Aparicio RM, et al. Probiotic Potential of Lactobacillus paracasei CT12 Isolated from Water Kefir
 S28 Grains (Tibicos). Curr Microbiol. 2020 Oct;77(10):2584–92.
- 529 37. Zavala L, Golowczyc MA, van Hoorde K, Medrano M, Huys G, Vandamme P, et al. Selected *Lactobacillus*530 strains isolated from sugary and milk kefir reduce *Salmonella* infection of epithelial cells *in vitro*. Beneficial
 531 Microbes. 2016 Sep 1;7(4):585–95.
- 532 38. Laureys D, De Vuyst L. The water kefir grain inoculum determines the characteristics of the resulting water
 533 kefir fermentation process. J Appl Microbiol. 2017 Mar;122(3):719–32.
- 534 39. Laureys D, Aerts M, Vandamme P, De Vuyst L. Oxygen and diverse nutrients influence the water kefir
 535 fermentation process. Food Microbiology. 2018 Aug;73:351–61.
- 40. Lee NY, Shin MJ, Youn GS, Yoon SJ, Choi YR, Kim HS, et al. *Lactobacillus* attenuates progression of
 nonalcoholic fatty liver disease by lowering cholesterol and steatosis. Clin Mol Hepatol. 2021 Jan 1;27(1):110–
 24.
- 539 41. Fijan S. Microorganisms with Claimed Probiotic Properties: An Overview of Recent Literature. IJERPH. 2014 May 5;11(5):4745-67.
- 541
 42. Tsai YS, Lin SW, Chen YL, Chen CC. Effect of probiotics *Lactobacillus paracasei* GKS6, *L. plantarum* GKM3, and
 542
 543
 543
 2020;14(4):299.
- 43. Yao F, Jia R, Huang H, Yu Y, Mei L, Bai L, et al. Effect of Lactobacillus paracasei N1115 and fructooligosaccharides in nonalcoholic fatty liver disease. aoms. 2019;15(5):1336-44.
- 546
 547
 548
 44. Ji Y, Xie Q, Meng X, Wang W, Li S, Lang X, et al. Lactobacillus paracasei improves dietary fatty liver by reducing insulin resistance and inflammation in obese mice model. Journal of Functional Foods. 2022
 548
 549
 540
 540
 541
 541
 542
 543
 544
 544
 544
 544
 544
 544
 544
 544
 545
 545
 546
 547
 548
 547
 548
 548
 548
 548
 549
 549
 549
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 540
 <l
- 549
 45. Elshaghabee FMF, Rokana N, Gulhane RD, Sharma C, Panwar H. Bacillus As Potential Probiotics: Status, Concerns, and Future Perspectives. Front Microbiol. 2017 Aug 10;8:1490.
- 46. Li YT, Ye JZ, Lv LX, Xu H, Yang LY, Jiang XW, et al. Pretreatment With Bacillus cereus Preserves Against D Galactosamine-Induced Liver Injury in a Rat Model. Front Microbiol. 2019 Jul 31;10:1751.

- 47. Xue J, Shen K, Hu Y, Hu Y, Kumar V, Yang G, et al. Effects of dietary Bacillus cereus, B. subtilis, Paracoccus marcusii, and Lactobacillus plantarum supplementation on the growth, immune response, antioxidant capacity, and intestinal health of juvenile grass carp (Ctenopharyngodon idellus). Aquaculture Reports. 2020 Jul;17:100387.
- 48. Kim B, Kwon J, Kim MS, Park H, Ji Y, Holzapfel W, et al. Protective effects of Bacillus probiotics against high-fat diet-induced metabolic disorders in mice. Aguila MB, editor. PLoS ONE. 2018 Dec 31;13(12):e0210120.
- 49. Neag MA, Catinean A, Muntean DM, Pop MR, Bocsan CI, Botan EC, et al. Probiotic Bacillus Spores Protect
 Against Acetaminophen Induced Acute Liver Injury in Rats. Nutrients. 2020 Feb 27;12(3):632.
- 562 50. Lynch KM, Wilkinson S, Daenen L, Arendt EK. An update on water kefir: Microbiology, composition and production. International Journal of Food Microbiology. 2021 May;345:109128.
- 564 51. Semjonovs P, Denina I, Linde R. Evaluation of Physiological Effects of Acetic Acid Bacteria and Yeast
 565 Fermented Non-alchocolic Beverage Consumption in Rat Model. J of Medical Sciences. 2014 Apr
 566 15;14(3):147-52.
- 567 52. Hong Y, Sheng L, Zhong J, Tao X, Zhu W, Ma J, et al. Desulfovibrio vulgaris, a potent acetic acid-producing
 bacterium, attenuates nonalcoholic fatty liver disease in mice. Gut Microbes. 2021 Jan 1;13(1):1930874.
- 569 53. Yang H, Meng L, Ai D, Hou N, Li H, Shuai X, et al. Acetic acid alleviates the inflammatory response and liver injury in septic mice by increasing the expression of TRIM40. Exp Ther Med [Internet]. 2019 Feb 13
 571 [cited 2023 Jan 3]; Available from: http://www.spandidos-publications.com/10.3892/etm.2019.7274
- 572 54. Li L, He M, Xiao H, Liu X, Wang K, Zhang Y. Acetic Acid Influences BRL-3A Cell Lipid Metabolism via the
 573 AMPK Signalling Pathway. Cell Physiol Biochem. 2018;45(5):2021–30.
- 574 55. Kondo T, Kishi M, Fushimi T, Kaga T. Acetic Acid Upregulates the Expression of Genes for Fatty Acid
 575 Oxidation Enzymes in Liver To Suppress Body Fat Accumulation. J Agric Food Chem. 2009 Jul
 576 8;57(13):5982-6.
- 577 56. Dai C, Li H, Wang Y, Tang S, Velkov T, Shen J. Inhibition of Oxidative Stress and ALOX12 and NF-κB
 578 Pathways Contribute to the Protective Effect of Baicalein on Carbon Tetrachloride-Induced Acute Liver
 579 Injury. Antioxidants (Basel). 2021 Jun 18;10(6):976.
- 580 57. Jiang ZY, Lu MC, You QD. Nuclear Factor Erythroid 2-Related Factor 2 (Nrf2) Inhibition: An Emerging
 581 Strategy in Cancer Therapy. J Med Chem. 2019 Apr 25;62(8):3840–56.
- 582 58. Zhao H, Eguchi S, Alam A, Ma D. The role of nuclear factor-erythroid 2 related factor 2 (Nrf-2) in the
 protection against lung injury. Am J Physiol Lung Cell Mol Physiol. 2017 Feb 1;312(2):L155–62.
- 584
 59. Zhao H, Eguchi S, Alam A, Ma D. The role of nuclear factor-erythroid 2 related factor 2 (Nrf-2) in the protection against lung injury. Am J Physiol Lung Cell Mol Physiol. 2017 Feb 1;312(2):L155–62.
- 586 60. Kaur G, Shivanandappa TB, Kumar M, Kushwah AS. Fumaric acid protect the cadmium-induced
 587 hepatotoxicity in rats: owing to its antioxidant, anti-inflammatory action and aid in recast the liver function.
 588 Naunyn-Schmiedeberg's Arch Pharmacol. 2020 Oct;393(10):1911–20.
- 589 61. Šilhavý J, Zídek V, Mlejnek P, Landa V, Šimáková M, Strnad H, et al. Fumaric Acid Esters Can Block Pro 590 Inflammatory Actions of Human CRP and Ameliorate Metabolic Disturbances in Transgenic Spontaneously
 591 Hypertensive Rats. Sookoian SC, editor. PLoS ONE. 2014 Jul 10;9(7):e101906.
- 592 62. Wang B, Cui S, Mao B, Zhang Q, Tian F, Zhao J, et al. Cyanidin Alleviated CCl4-Induced Acute Liver Injury
 593 by Regulating the Nrf2 and NF-κB Signaling Pathways. Antioxidants. 2022 Dec 1;11(12):2383.
- 63. Ganesh Yerra V, Negi G, Sharma SS, Kumar A. Potential therapeutic effects of the simultaneous targeting of the Nrf2 and NF-κB pathways in diabetic neuropathy. Redox Biology. 2013;1(1):394–7.

- Li H, Weng Q, Gong S, Zhang W, Wang J, Huang Y, et al. Kaempferol prevents acetaminophen-induced liver injury by suppressing hepatocyte ferroptosis *via* Nrf2 pathway activation. Food Funct. 2023;14(4):1884–96.
- 65. Rahman ZU, Al Kury LT, Alattar A, Tan Z, Alshaman R, Malik I, et al. Carveol a Naturally-Derived Potent and Emerging Nrf2 Activator Protects Against Acetaminophen-Induced Hepatotoxicity. Front Pharmacol. 2021 Jan 28;11:621538.
- 601 66. Gao W, Guo L, Yang Y, Wang Y, Xia S, Gong H, et al. Dissecting the Crosstalk Between Nrf2 and NF-κB
 602 Response Pathways in Drug-Induced Toxicity. Front Cell Dev Biol. 2022 Feb 2;9:809952.
- 603 67. Sharifi-Rad J, Seidel V, Izabela M, Monserrat-Mequida M, Sureda A, Ormazabal V, et al. Phenolic
 604 compounds as Nrf2 inhibitors: potential applications in cancer therapy. Cell Commun Signal. 2023 May
 605 1;21(1):89.
- 606
 68. Luedde T, Schwabe RF. NF-κB in the liver linking injury, fibrosis and hepatocellular carcinoma. Nat Rev
 607
 68. Gastroenterol Hepatol. 2011 Feb;8(2):108–18.
- 608
 69. Liu Z wei, Zhang Y ming, Zhang L ying, Zhou T, Li Y yang, Zhou G cheng, et al. Duality of Interactions
 609 Between TGF-β and TNF-α During Tumor Formation. Front Immunol. 2022 Jan 5;12:810286.
- 610 70. Yang YM, Seki E. TNFα in Liver Fibrosis. Curr Pathobiol Rep. 2015 Dec;3(4):253-61.
- 611
- 612 Please, insert here the Supplementary Data, if any.
- 613 614

	Hepatoprotective Study of Indonesian Water Kefir Against					
	CCl ₄ -Induced Liver Injury in Rats					
•	Ru	nning title: Hepatoprotective	e study of water kefir			
	Aligita W ^{1,3*} , Singgih M ² , Sutrisno E ³ , Adnyana IK ^{1*}					
)						
	¹ Department of Pharmacology & Clinical Pharmacy, School of Pharmacy, Bandung Institute of Technology, Bandung, Indonesia. ² Department of Pharmacochemistry, School of Pharmacy, Bandung Institute of Technology, Bandung, Indonesia. ³ Department of Pharmacology & Clinical Pharmacy, Faculty of Pharmacy, Bhakti Kencana University, Bandung, Indonesia. *E-mail address: <u>widhya.aligita@bku.ac.id</u> / <u>ketut@itb.ac.id</u>					
	*Corresponding author address: Department of Pharmacology & Clinical Pharmacy, School of Pharmacy, Bandung Institute of Technology, Ganeca, 10, 40132, Bandung, Indonesia. Tel: +62-22-2504852					
Т. ат	The institutional e-mail of tuthors):	the ALL authors must be declared ((and add rows according to t	he number of		
А	Author	Institutional e-mail	Other e-mail	ORCID		

Author	Institutional e-mail (mandatory)	Other e-mail (gmail, yahoo, etc.)	ORCID (0000-1111-2222-3333)
Widhya Aligita	widhya.aligita@bku.ac.id	w.aligita@gmail.com	0000-0001-8338-4115
Marlia Singgih	marlia@fa.itb.ac.id		0000-0002-5351-1731
Entris Sutrisno	entris.sutrisno@bku.ac.id		0000-0003-3830-6411
I Ketut Adnyana	ketut@itb.ac.id		0000-0001-5217-2312

Contribution Details (to be ticked marked (X) as applicable and add columns according to the number of authors): 21

Contribution	Aligita W	Singgih M	Sutrisno E	Adnyana IK
Concepts or Ideas	Х	Х	Х	Х
Design	x	х	х	x
Definition of intellectual content	х	х	х	x
Literature search	х			x
Clinical trial				
Experimental studies	X			x
Data acquisition	x			X
Data analysis	х			x
Statistical analysis	х			x
Manuscript preparation	х	х	х	x
Manuscript editing	х	Х	х	x
Manuscript review	х	х	Х	X

24 ABSTRACT

- 25 26 27 Context: Water kefir is a fermented beverage that is typically made in the home by inoculating a sugar-rich solution with a microbial community (water kefir grains). Several studies on the metabolite content and hepatoprotective effects of water kefir have been published, but CCl4-induced acute liver injury has not been studied.
- 28 29 Objectives: To evaluate the efficacy of water kefir in vivo against hepatoprotective CCl4-induced acute liver injury and to in silico investigate metabolites that play an important role in hepatoprotective mechanisms.
- Methods: The present study aimed to investigate the hepatoprotective activity of water kefir in an animal model caused by carbon tetrachloride (CCl4). Furthermore, using molecular docking, the metabolites found in water kefir were evaluated for their role in the NF-KB and Nrf2 signaling pathways.
- 30 31 32 33 34 35 36 37 Results: Water kefir significantly and dose-dependently alleviated acute liver injury caused by carbon tetrachloride (CCl4). Water kefir administration at all doses produced results comparable to the positive control (Curcuma extract). Molecular docking simulations showed that, compared to Nrf2, the 25 metabolites were more likely to interact with the NF-B receptor. Fumaric acid is the strong metabolite that interacts with the NF-KB receptor with a free energy of binding and an inhibition constant of -6.66 kcal/mol and 13.22 µM, respectively.
- 38 39 Conclusions: Water kefir administration improved the condition of liver damage, characterized by a decrease in serum levels of AST, ALT, TNF-α, TGF-β, and an improvement in the liver tissue profile. In silico evaluation showed that the metabolites in water 40 kefir were able to interact with target proteins in the NF-κB and Nrf2 pathways. It was concluded that water kefir improves the 41 condition of the liver by reducing the level of necrosis and fibrosis.
- 42
- 43 Keywords: Free radicals; Liver diseases; Kefir; Molecular docking simulation; Probiotics
- 44

45 INTRODUCTION

46 In most cases, making water kefir involves combining dried fruit, sugar, and water kefir 47 grains in a container. Water kefir's exact origins are unknown; however, two hypotheses have 48 been proposed regarding its history: the first suggests that water kefir grains were brought to 49 Europe from the Caucasus by soldiers returning from the Crimean War in the late nineteenth 50 century (Ward 1892); the second theory proposes that water kefir grains originated in Mexico from the Opuntia cactus through natural processes (Moinas et al. 1980). Sugary kefir grains, 51 52 Balm of Gilead, African bees, Japanese beer seeds, Ale nuts, and California bees are some other 53 names for water kefir. Tibi grains and ginger beer plants are other names for water kefir 54 (Kebler 1921; Moinas et al. 1980; Pidoux 1989). Water kefir is appealing to both consumers and 55 researchers due to the variety of microbiota it contains, the fact that it is an alternative to dairy 56 products, the versatility with which it can be flavored, the fact that it is low in calories and 57 sugar, the ease with which it can be produced, and the health benefits it offers.

58 Water kefir has been used medicinally for a very long time, and recent research has 59 indicated that it may have a variety of positive effects on people's health. It has been demonstrated that water kefir contains non-pathogenic bacteria that, in conjunction with the 60 61 production of organic acids, can inhibit the growth of pathogenic microbes such as Shigella sp., 62 Salmonella sp., Staphylococcus aureus, and E. coli; as well as, filamentous fungi such as Aspergillus 63 ochraceus, A. niger, A. flavus, Penicillium sp., and Rhizopus sp. (Al-Mohammadi et al. 2021). In addition to its antibacterial properties, water kefir possesses a broad spectrum of 64 65 pharmacological effects. Some of these therapeutic effects are anti-inflammatory (Diniz et al. 2003; Aligita et al. 2020), antioxidant (Alsayadi et al. 2013; Aligita et al. 2020; Darvishzadeh et 66 67 al. 2021), hepatoprotective (Aspiras et al. 2015; Aligita et al. 2021), antihyperglycemic and 68 antihyperlipidemic (Alsayadi et al. 2014; Rocha-Gomes et al. 2018), anti-edematous (Moreira 69 et al. 2008), antitumor (Zamberi et al. 2016), antihypertensive (Gamba et al. 2019), immunomodulant (Calatayud et al. 2021), and anti-ulcerogenic (Rodrigues et al. 2016). 70 71 However, no studies have been reported on the hepatoprotective effects of water kefir against 72 carbon tetrachloride (CCl₄)-induced liver injury.

73 Studies have shown that acute liver injury is frequently accompanied by high levels of 74 oxidative stress and inflammatory responses (Dai et al. 2021a). These findings have been found 75 in several studies. The most important signaling pathways that are involved in the regulation 76 of inflammation and antioxidation are the nuclear factor (NF-KB) and nuclear factor erythroid 77 2-related factor 2 (Nrf2) pathways, respectively. It has been shown that activating Nrf2 and 78 inhibiting NF-KB can reduce the amount of damage done to the liver. For instance, curcumin 79 protects against aflatoxin B1-induced liver injury by increasing the expression of Nrf2 and 80 related downstream antioxidant molecules (such as superoxide dismutase (SOD), catalase 81 (CAT), heme oxygenase 1 (HO-1), and NAD(P)H quinone dehydrogenase 1 (NQO-1) (Wang 82 et al. 2022b). Also, it has been shown that the molecule methoxy eugenol, which comes from 83 nutmeg and Brazilian red propolis, protects the liver both in vitro and in vivo. This may be 84 attributed to the fact that it targets the NF-kB signaling pathway, which has been shown to 85 have anti-inflammatory effects (De Souza Basso et al. 2021).

86 Ethanol and lactic acid are the primary metabolites found in fermented water kefir, while 87 lower quantities of glycerol, mannitol, and acetic acid can also be found in the beverage.

Additionally, a variety of aromatic and volatile compounds are produced, including ethyl acetate, isoamyl acetate, ethyl octanoate, ethyl hexanoate, and ethyl decanoate, among others

90 (compared to their threshold levels) (Laureys and De Vuyst 2014). The chemical constituents

91 of both phytochemicals and secondary metabolites in natural products, including water kefir,

92 are certainly capable of providing various pharmacological effects for the body (A. Asnawi et

al. 2022; Nursamsiar et al. 2022). However, an *in silico* study to evaluate the metabolite content

94 in water kefir has not been reported yet. Because of its capacity to speed up the process of 95 identifying and optimizing lead compounds, the *in silico* method has become the front-runner

96 in the race to improve the speed and accuracy of the process of discovering new drugs. This is

97 because the *in silico* method can identify and optimize lead compounds more quickly.

98 Techniques such as molecular docking and molecular dynamics (MD) were able to directly

99 indicate a small number of compounds that have high affinity and selectivity by analyzing

100 how the ligand and target interact with one another (Febrina et al. 2021).

Water kefir has been used for an extensive period of time and has been recognized for its widespread benefits, especially in Indonesia. However, its level of popularity falls short in comparison to that of milk kefir. Furthermore, there is a scarcity of studies specifically focused on the hepatoprotective activity of water kefir made with Indonesian grains. Therefore, the purpose of this study was to evaluate the hepatoprotective effects of water kefir in CCl₄induced rats while also investigating the stability interactions of its metabolites within the NFkB and Nrf2 receptors using molecular docking studies.

108

109 MATERIAL AND METHODS

110 Materials and Reagents

111 Water kefir grains, sugar, dried fruit, carbon tetrachloride, rats, diagnostic kits for alanine 112 aminotransferase (ALT) (Proline, IFCC mod.), aspartate aminotransferase (AST) (Proline, 113 IFCC mod.), Elisa Kit TNF- α (Bioassay Technology Laboratory), Elisa Kit TGF- β (Bioassay 114 Technology Laboratory). Other chemicals used in this study were of analytical reagent grade.

115 Experimental sample and reference extract

The water kefir grain was obtained from Yogyakarta, Indonesia. The water kefir solution was produced using a fermentation procedure. The initial stage involved the preparation of 60 g of sugar, 2 g of raisins, 100 g of water kefir grains, and 1 L of aqua mineral distillate. The sugar and warm distilled water were mixed in a beaker, followed by the addition of water kefir grains and raisins to the resulting sugar solution. The fermentation procedure was conducted over a duration of forty-eight hours at ambient temperature. A small cloth was used to cover the beaker glass. The kefir grain was utilized in future production, while the filtratewas employed for the purpose of evaluation and analysis. (Aligita et al. 2020, 2021)

124 The rhizoma extract of Curcumae (Curcuma Xanthorrhiza Roxb) is employed as a reference 125 drug. The utilized product is a standardized herbal medicine with the brand name Tulak, 126 manufactured by the Borobudur Company Herbal Medicine Industry. The primary purpose 127 of Tulak capsules is to support and preserve optimal liver functionality.

128 Animals and Experimental Design

129 Rats (Wistar strain, male, 200-250 g) were maintained on normal pellet food and tap water 130 ad libitum. Four mice in each group were used. All procedures relating to animals and their 131 care conformed to the international guidelines Principles of Laboratory Animal Care (NIH 132 publication no. 85-23, revised 1985) with the ethical approval number 133 363/UN6.KEP/EC/2020. In order to develop an animal model with liver injury, the rats 134 received CCl₄ (20% in olive oil) at 1.25 mL/kg BW every 2 days via a gavage tube (Yeh et al. 135 2011). The rats were randomized into five groups after the development of animals with liver 136 injury, which is characterized by a significant increase in serum ALT level, as follows: (1) 137 positive control group, (2) Curcuma extract group 100 mg/kg BW, (3) water kefir 15 mL/kg 138 BW group, (4) water kefir 35 mL/kg BW group, and (5) water kefir 50 mL/kg BW group, with 139 the addition (6) negative control group. Each group received group-specific treatment for two 140 weeks, along with the administration of 1.25 mL/kg BW of CCl₄ (20% in olive oil) every three 141 days.

142 The rats, which had undergone a fasting period of 8-10 hours while being provided with 143 water, had their blood samples collected via the retro-orbital sinus using a hematocrit capillary 144 tube. The blood sample was taken into a microtube and afterwards then to centrifugation. The 145 serum was separated in order to facilitate further measurements (Parasuraman et al. 2010). 146 Serum ALT level, as the main parameter, was measured prior to induction, following 147 induction, and following treatment. Meanwhile, following therapy, serum AST, TNF-a, TGF-148 β levels, and liver histopathological were evaluated. The ALT, AST, TNF- α , TGF- β levels 149 measurements are conducted in accordance with the protocols outlined in the reagent kit.

After the euthanasia procedure, the liver specimen was promptly immersed in a 10% formalin solution at room temperature for a duration of 24 hours. Subsequently, the tissues were immersed in paraffin, subjected to sectioning, and affixed onto glass microscope slides. The slices underwent staining with hematoxylin and eosin and were afterwards analyzed using light microscopy (Konstantopoulos et al. 2017).

155 Molecular docking simulation

156 Molecular docking experiments were done with the PyRx software (Dallakyan and Olson 157 2015) to predict how metabolites, which are small-molecule ligands, bind to biological 158 macromolecules. The NCBI PubChem database (https://pubchem.nlm.nih.gov/, accessed on 159 3 May 2023) was used to derive the three-dimensional structure of water kefir metabolites 160 (Patel et al. 2022). Target proteins like NF-κB (PDB code: 1A3Q) and Keap1 (PDB code: 4L7B) were obtained from the RCSB Protein Data Bank (http://www.rcsb.org/, accessed on 03 May 161 162 2023). The PyRx program was used to reduce protein, ligand converted to PDBQT (Asnawi et 163 al. 2023), then maximize GRID parameter (Asnawi et al. 2022) and perform docking study (Febrina et al. 2022). The BIOVIA Discovery Studio 2017 R2 program was used to view the 164 165 protein and ligand complex and distance (Ischak et al. 2023). The BIOVIA Discovery Studio 166 2017 R2 tool was also utilized to find protein active sites.

167 Statistical Analysis

All of the information is displayed in the form of individual data points as well as the mean along with the standard error of the mean. The statistical analysis was carried out with the help of Minitab software (version 19.0), and to make comparisons between several different groups, a one-way analysis of variance with Dunnett's post hoc test was utilized. All statistical graphs were created with Microsoft Excel 2019 in their respective versions. The levels of significance that were considered to have been reached were *p < 0.05.

174 **RESULTS**

175 In vivo evaluation of hepatoprotective activity

176 The serum ALT levels, as the main parameter for the liver damage, were measured prior to 177 and following induction, as well as following treatment (Fig. 1). Meanwhile, after treatment 178 AST, TNF-α, and TGF-β levels were also evaluated. These findings were analyzed using a one-179 way ANOVA with a 95% confidence level and a two-sided confidence interval. A significant 180 rise in blood ALT levels demonstrated the establishment of an animal model with liver injury, 181 according to statistical analysis, following the administration of CCl₄. When compared to the 182 positive control group, ALT serum levels decreased significantly after two weeks of therapy 183 with curcuma extract or water kefir. The three doses of water kefir groups demonstrated 184 equivalent activity when curcuma extract was used as the standard treatment, and there was 185 no significant difference between the three doses of water kefir. When compared to the 186 positive control group, AST levels were also reduced dramatically following treatment with 187 curcuma extract or water kefir. TNF-q levels in the water kefir group were significantly lower 188 than in the positive control group at dosages of 35 and 50 mL/kg body weight. Even though 189 there was no statistically significant difference in TGF-β levels, the group that received the 190 treatment demonstrated a decrease in TGF-β levels.



Fig. 1. (a) Before induction, after induction, and after treatment serum ALT level; (b) After treatment serum AST level; (c) After treatment serum TNF- α level; (d) After treatment serum TGF- β level. * indicates a significantly different result when compared to the positive control group; # indicates a significantly different result when compared to the curcuma extract group; p < 0.05; n = 4 mice in each group.

192 A histological examination of a normal liver group revealed a typical central vein bordered 193 with endothelial cells and normal hepatocytes, resulting in hepatic cords of cells with distinct 194 cell borders and sinusoidal gaps (Figs 2A and 2B). The CCl4-induced group developed 195 centrilobular hepatic necrosis level 4, which was characterized by massive vacuolization and 196 necrosis of epithelial cells in the liver's centrilobular region (blue arrow) (Fig 2C & 2D). The 197 group that received either curcuma extract or water kefir treatment improved in varied 198 necrotic conditions ranging from level 1 (water kefir 50 mL/kg BW) (Fig 2K & 2L) to level 2 199 (curcuma extract (Fig 2E & 2F), water kefir 15 mL/kg BW (Figs 2G and 2H), and water kefir 200 30 mL/kg BW (Fig 2I & 2J)).



Fig. 2. Liver histology after CCl4 intoxication and treatment with HE staining. Negative control group (A & B), positive control group (C & D), curcuma extract group (E & F), water kefir 15 mL/kg BW (G & H), water kefir 30 mL/kg BW (I & J), and water kefir 50 mL/kg BW (K & L). 1: central vein; 2: normal liver epithelial cell; 3: portal track; blue arrow: necrosis and liver epithelial cell vacuolization at centrolobular region.

202 Molecular docking

203 Molecular docking studies are considered a powerful tool for predicting the potential 204 targets of bioactive molecules. In order to carry out molecular docking simulations, one of the 205 most critical steps is to identify the target active site. If the target protein is crystallized with a 206 native ligand, in many instances, the location of the active site can be established without any 207 difficulty (Li et al. 2019b). However, the NF-kB (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) 208 proteins do not have a native ligand, so the active site was determined. Active site prediction 209 in docking is a computational method for predicting the location and orientation of a receptor 210 protein's binding site for a ligand molecule. The active site prediction was based on a protein 211 structural analysis and the identification of amino acid residues that are likely to interact with 212 the ligand. The projected binding site is then utilized as a starting point for molecular docking, 213 a computer method for predicting a ligand molecule's binding affinity and orientation to a 214 receptor protein. The active site prediction for target proteins (Keap1 and NF-κB) gives the 215 grid box coordinates (x y z) of 17.500880 Å, 62.323000 Å, and 0.973748 Å; and -1.751769 Å, -216 20.743853 Å, and -29.010438 Å, respectively (Fig. 3).



Fig. 3. Binding pocket (colored in red) generated using the BIOVIA Discovery Studio 2017 tool, coupled with the sequence containing the highlighted residues that constitute the binding pocket. A) NF-κB (PDB ID 1A3Q) and B) Keap1 (PDB ID 4L7B).

217

The docking results of the 25 metabolites could interact with target proteins (Keap1 and NF-κB) (Table 1). In general, all metabolites could interact with both NF-κB receptors (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) (Table 1). The interaction of metabolites with nuclear factor-kappa B receptors (PDB ID 1A3Q) revealed that 13 metabolites possessed free energy for binding that was greater than that of curcumin. On the other hand, there was not a single metabolite that had a free energy binding that was more powerful than that of nrf2 Keap1 (PDB ID 4L7B) (Table 1).

225 For volatile compounds, 2-phenyl ethanol and benzaldehyde interact most strongly with 226 the NF-kB receptor (PDB ID 1A3Q) with almost the same binding energy of -4.19 kcal/mol. 227 As for organic acids, succinic acid, fumaric acid, and citric acid provide nearly the same strong 228 interactions. Bond energy values of fumaric acid, succinic acid, and citric acid of -6.66, -6.24, 229 and -6.25 kcal/mol, respectively. In sugars, glucose provides the strongest interaction with the 230 NF-KB receptor (PDB ID 1A3Q) where the binding energy was -3.71 kcal/mol. As for the nrf2 231 Keap1 receptor (PDB ID 4L7B), 2-phenylacetaldehyde from volatile compounds was able to 232 interact most strongly with the Keap1 receptor (PDB ID 4L7B) with a binding energy of -3.46 233 kcal/mol. As for organic acids, succinic acid provides nearly the same strong interaction with a bond energy value of -3.02 kcal/mol, respectively. In sugars, glucose provides the strongest interaction with the Keap1 receptor (PDB ID 4L7B) where the binding energy was -3.09

236 kcal/mol.

- 237
- 238

Table 1. The free energy of binding and inhibition constant of water kefir metabolite active site interaction on NF κB (PDB ID 1A3Q) and Keap1 (PDB ID 4L7B)

No.	Metabolites	PDB: 1A3Q		PDB: 4L7B	
		Free Energy of Binding, ∆G (kcal/mol)	Inhibition Constant, Ki (µM)	Free Energy of Binding, ∆G (kcal/mol)	Inhibition Constant, Ki (μΜ)
	Volatile compounds			, <u> </u>	
1	2-Methylbutanol	-3.31	3,770	-2.68	10,890
2	3-Methylbutanol	-3.22	4,360	-2.50	14,670
3	2-Phenylethanol	-4.19	851.52	-3.12	5,140
4	Ethyl acetate	-3.19	4,590	-2.43	16,670
5	2-Methylbutanal	-3.26	4,050	-2.35	19,050
6	1-Octanol	-2.65	11,490	-2.11	28,220
7	2-Phenylethyl acetate	-3.99	1,190	-3.33	3,590
8	Benzyl acetate	-4.16	887.66	-3.44	3,020
9	Benzaldehyde	-4.19	853.74	-3.19	4,550
10	Furfuryl acetate	-3.60	2,320	-3.32	3,660
11	Isobutyl acetate	-3.19	4,610	-2.94	7,020
12	Isopentyl acetate	-3.20	4,510	-3.05	5,820
13	2-Phenylacetaldehyde	-4.07	1,030	-3.46	2,930
14	3-Methylbutanal	-3.16	4,850	-2.73	10,020
15	Gluconic acid	-3.11	5,280	-1.63	64,290
	Organic acids				
16	Lactic acid	-4.92	249.28	-2.06	30,680
17	Succinic acid	-6.24	26.68	-3.02	6,140
18	Fumaric acid	-6.66	13.22	-2.45	16,100
19	Citric acid	-6.25	26.18	-1.87	42,440
20	Malic acid	-5.65	72.69	-2.53	13,860
21	Acetic acid	-4.99	219.33	-2.44	16,280
22	Ethanol	-2.39	17,800	-2.08	29,800
	Sugars				
23	Sucrose	-1.40	93,710	-1.33	105,320
24	Glucose	-3.71	1,900	-3.09	5,440
25	Fructose	-3.21	4,400	-2.65	11,440
	Curcumin	-3.44	2,990	-4.22	811.36

241

242 The theoretical binding modes of the top three metabolites with their target proteins (Keap1 243 and NF-KB) are shown in Figs. 4 and 5, respectively. The molecular docking results suggested 244 that these metabolites interacted with the Keap1 and NF-KB to form a complex through 245 hydrogen bonds with various residues. The interaction of 2-phenyl ethanol with the active site of NF-kB was formed by two hydrogen bonds and one Pi-Alkyl interaction with amino acid 246 247 residues of PRO223, LYS252, and LYS252, respectively. The interaction of benzaldehyde with the active site of NF-KB was formed by a hydrogen bond with the amino acid residue of 248 249 LYS252. The interaction of fumaric acid with the active site of NF-KB was formed by six 250 hydrogen bonds with the amino acid residues of LYS221, LYS252, and TYR285. The interaction 251 of glucose with the active site of NF-kB was formed by six hydrogen bonds with the amino 252 acid residues SER220, LYS221, SER226, and LYS252. Although glucose and fumaric acid were 253 able to be formed by six hydrogen bonds, different types of amino acid residues were involved

- 254 in the interaction, so fumaric acid interacted more strongly with the active site of NF-κB (Fig.
- 255 4).
- 256



Fig. 4. 2D and 3D representation of the kind of interaction on the formation of the NF-κB active site (PDB ID 1A3Q). A) 2-Phenylethanol, B) Benzaldehyde, C) Fumaric acid, D) Glucose, and E) Curcumin.



D Fig. 5. 2D and 3D illustration of the type of interaction on the formation of Keap1's binding pocket (PDB ID 4L7B). A) 2-Phenylacetaldehyde, B) Succinic acid, C) Glucose, and D) Curcumin.

259

260 The interaction of 2-phenylacetaldehyde with the active site of Keap1 was formed by one 261 hydrogen bond and one Pi-Alkyl interaction with amino acid residues of GLN530 and ALA556, respectively. The interaction of succinic acid with the active site of Keap1 was formed 262 by four hydrogen bonds with the amino acid residues of ARG336, HIS575, and PHE577. The 263 264 interaction of glucose with the active site of Keap1 was formed by six hydrogen bonds with the amino acid residues LYS551, ARG553, and ASP573. Although glucose can form more 265 hydrogen bonds than 2-phenylacetaldehyde, the different types of amino acid residues 266 267 involved have not been able to have a significant effect on the binding energy of its interaction 268 with the active site of Keap1 (Fig. 5).

269 Curcumin (the reference compound) created three hydrogen bonds with the amino acid 270 residues ARG52, GLU58, and LYS252 to interact with the active site of NF-KB. Meanwhile, 271 curcumin established three hydrogen bonds with the amino acid residues ARG336, TYR572, 272 and GLY600 to interact with the active site of nrf2 Keap1 (Figs 4 and 5). Despite the fact that curcumin could create three hydrogen bonds at both the active sites of NF-KB and nrf2 Keap1, 273 274 its interaction with nrf2 Keap1 had a lower free energy of binding than phenylethanol, 275 benzaldehyde, and fumaric acid (Table 1). Overall, the metabolites of water kefir prefer to 276 interact with NF-KB and nrf2 Keap1 receptors. Whereas, fumaric acid and 2-277 phenylacetaldehyde were metabolites that had the strongest interaction with NF-KB (PDB ID 278 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) receptors, respectively.

279 DISCUSSION

280 Increased liver enzyme production is one of the abnormalities indicating liver damage. This 281 increase in secretion increases the leakage of enzymes into the circulation. Transaminase is a 282 frequent enzyme used to evaluate liver disorders. The transaminases AST and ALT catalyze 283 the transfer of the α -amino group from alanine and aspartic acid to α -ketoglutaric acid. AST is found in the liver, heart, and skeletal muscle, as well as the kidney, brain, pancreas, lung, 284 285 leukocytes, and erythrocytes. While ALT is predominantly found in the liver, it is also found 286 in low concentrations in other tissues (Lee et al. 2012). Consequently, ALT was used as the 287 principal hepatotoxicity criterion in this study.

288 Carbon tetrachloride (CCl₄) is a component of the hepatotoxin, which acts after metabolic 289 activation and is extensively employed as a liver-damaging agent. In this study, the 290 administration of CCl₄ to animals caused hepatotoxicity, which was characterized by a 291 significant increase in ALT after several administrations. CCl₄ is metabolized by the enzyme 292 cytochrome p450 (CYP2E1) in the endoplasmic reticulum to a highly reactive trichloromethyl 293 radical. The trichloromethyl radical rapidly reacts with oxygen to form the trichloromethyl 294 peroxyl radical, which rapidly reacts with lipids to form lipid peroxidation products. Free 295 radicals will disrupt the endoplasmic reticulum (ER) and lead to lipid accumulation, decreased 296 protein synthesis, and increased oxidase activity. CCl₄ hepatotoxicity is characterized by 297 hepatocellular necrosis with fat deposition (Ritesh et al. 2015). At the molecular level, 298 administration of CCl₄ can activate tumor necrosis factor (TNF)-a, nitric oxide (NO), and 299 transforming growth factor (TGF)- α and - β in cells, processes that precipitate cell self-300 destruction or fibrosis. TNF-α leads to apoptosis, whereas TGF-β leads to fibrosis (Weber et al. 301 2003).

302 In terms of its pathophysiological underpinnings, liver illness is linked to a condition known 303

as dysbiosis, which refers to an imbalance in the makeup of the gut microbiota (Zavala et al.

304 2016; Laureys and De Vuyst 2017; Romero-Luna et al. 2020). Both qualitative and quantitative

changes in the gut microbiome have the potential to affect the composition of products 305

307 2020). Qualitative changes include an imbalance between harmful and helpful microbiomes, 308 whereas numeric changes involve changes to the overall microbiota. In addition to these 309 symptoms, intestinal inflammation, disruption of the intestinal barrier, and the transfer of 310 microbial products can all be caused by dysbiosis (Laureys et al. 2018). For this reason, the 311 condition of the gut microbiome is an important factor in the initiation and development of chronic liver disease (Lee et al. 2021). Based on the results of the study, treatment with water 312 313 kefir for 2 weeks after the occurrence of liver damage was able to improve the overall condition 314 of the liver which was marked by a significant decrease in the values of AST, ALT, TNF-a,

315 TGF- β , and significant improvement in liver histology.

316 Water kefir contains a number of microorganisms that have been linked to health benefits, 317 such as the probiotics *L. paracasei* and *B. cereus* (Fijan 2014). This activity is linked to an increase in antioxidants like glutathione and catalase and a decrease in pro-inflammatory transcription 318 319 factors like nuclear B-factor, lipopolysaccharide, and Toll-like receptor 4 (TLR-4). 320 Improvements in intestinal barrier function and histological integrity were also observed. 321 Increased expression of claudin-1 and occludin-1, two components of tight junctions, occurs 322 simultaneously with the restoration of the p38 MAPK pathway (Fijan 2014; Yao et al. 2019; 323 Tsai et al. 2020; Ji et al. 2022). Bacillus is a kind of endospore-forming bacterium that can 324 endure extremely cold temperatures and lengthy periods of storage without dving; its spores can even tolerate the acidic environment of the stomach and make it all the way to the small 325 326 intestine (Elshaghabee et al. 2017). B. cereus has been shown to reduce ALT levels, an indicator 327 of liver healing, in various animal models of liver injury. It protects the liver by reducing 328 inflammation, enhancing the gut flora, and strengthening the tight junctions in the intestines 329 (Kim et al. 2018; Li et al. 2019a; Xue et al. 2020). Also, when Bacillus spores were used first, 330 hepatocyte necrosis and serum levels of ALT, ZO-1, AST, and TAC went down by a lot. This 331 effect is comparable to that of the popular hepatoprotective compound silvmarin (Neag et al. 332 2020).

333 Beer, vinegar, cider, and cocoa bean masses are just some of the places where Gram-334 negative acetic acid bacteria have been found. One of the features that sets acetic acid bacteria apart from others is their alkaline-stable lipid membrane (Lynch et al. 2021). Their "oxidative" 335 336 fermentation metabolism is responsible for the principal metabolic process in these bacteria, 337 the oxidation of ethanol to acetic acid. Important in the food and beverage sector and beyond, 338 fermentation helps mediate the transition of diverse substrates into products. Although lactic 339 acid bacteria have been studied more extensively than acetic acid bacteria (Semjonovs et al. 340 2014; Hong et al. 2021), various studies have shown promising results concerning the 341 pharmacological effects of acetic acid bacteria, especially as a hepatoprotective agent. Acetic 342 acid, which is the main metabolite of acetic acid bacteria, can lower inflammation and the 343 severity of liver injury in rats with septic shock by increasing the expression of TRIM40. 344 TRIM40 has been shown to minimize liver damage, decrease the synthesis and release of 345 cytokines such as IL-6 and TNF-a, raise the expression of IL-10, improve survival in septic 346 mice, and block the activity of the TLR4 signaling pathway. Acetic acid injection decreased 347 inflammation as well as the production of inflammatory cytokines (Yang et al. 2019). Acetic 348 acid also stimulates the AMPK signaling pathway, which leads to enhanced lipid oxidation 349 and reduced hepatic lipid and body fat deposition (Kondo et al. 2009; Li et al. 2018).

Apart from microorganisms that directly provide hepatoprotective effects, the metabolites
 produced from these microorganisms also have the potential to be hepatoprotective.
 Molecular docking is a technique that is utilized in the context of NF-κB and Nrf2 to make
 predictions regarding the binding affinity and orientation of small-molecule inhibitors to their
 active sites. The transcription factor known as NF-κB is an essential component in the

355 management of both the immune system and the inflammatory response (Dai et al. 2021b). 356 The expression of important inflammatory genes can be inhibited by small-molecule inhibitors that impair the interaction between NF-κB and DNA. These inhibitors have the potential to be 357 358 used in therapeutic applications. The Nrf2 binding site is Kelch-like ECH-associated protein 1 359 in the context of nuclear factor erythroid 2-related factor 2, also known as Nrf2 (Keap1) (Zhao 360 et al. 2017a; Jiang et al. 2019). Small-molecule inhibitors that disrupt the link between Keap1 361 and Nrf2 can activate the Nrf2-ARE signaling pathway, which has been demonstrated to have 362 cytoprotective effects. Keap1 is a negative regulator of Nrf2, and this association can be 363 disrupted by small-molecule inhibitors (Zhao et al. 2017b).

364 Binding energy and Ki are important parameters used in molecular docking to evaluate the strength of the interaction between a ligand and a receptor protein. Binding energy is the 365 366 energy released when a ligand binds to a receptor protein, and it is calculated as the difference 367 between the energy of the bound complex and the energy of the unbound ligand and protein 368 (Meng et al. 2011). Ki, on the other hand, is the dissociation constant of the ligand-receptor 369 complex. It shows how much ligand is needed to bind 50% of the receptor binding sites. Both 370 binding energy and Ki are used to predict the binding affinity and selectivity of a ligand to a 371 receptor protein (Du et al. 2016). Ki, on the other hand, is the dissociation constant of the 372 ligand-receptor complex. It shows how much ligand is needed to bind 50% of the receptor 373 binding sites (Du et al. 2016).

374 Based on the results of an in silico study of water kefir metabolite compounds, it was known 375 that fumaric acid and 2-phenylacetaldehyde have the strongest interactions with NF-KB (PDB 376 ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) receptors, respectively. Fumaric acid has been 377 studied for its potential as a hepatoprotective agent. Fumaric acid protects rat livers against 378 cadmium toxicity by decreasing oxidative stress and improving hepatic serum. Fumaric acid 379 prevented cadmium-accrued hepatocellular damage and showed the potential to avert hepatic 380 injury against cadmium in rats (Kaur et al. 2020). Fumaric acid esters were found to ameliorate 381 inflammation and oxidative stress in rats, which was associated with less adiposity and ectopic 382 fat accumulation (Šilhavý et al. 2014).

383 NF- κ B and Nrf2 are two transcription factors that play important roles in regulating 384 inflammation and cell survival. While NF-KB is involved in the inflammatory response, and 385 Nrf2 is involved in the antioxidant response (Ganesh Yerra et al. 2013; Wang et al. 2022a). Both 386 transcription factors have been investigated as potential targets for the development of 387 hepatoprotective agents (Rahman et al. 2021; Gao et al. 2022; Wang et al. 2022a; Li et al. 2023). 388 There is evidence of crosstalk between the Nrf2 and NF-kB pathways (Ganesh Yerra et al. 2013; 389 Gao et al. 2022). The Nrf2 pathway inhibits the activation of the NF-KB pathway by increasing 390 antioxidant defenses and HO-1 expression, which efficiently neutralizes ROS and detoxifies 391 (Ganesh Yerra et al. 2013). The crosstalk between Nrf2 and NF-KB could be a new therapeutic 392 target against hepatotoxicity (Gao et al. 2022). Researchers have tried to identify molecule 393 activators of Nrf2 as chemoprevention for ROS-dependent carcinogenesis, while others have 394 focused on identifying Nrf2 inhibitors to increase the sensitivity of cancer cells to 395 chemotherapy (Sharifi-Rad et al. 2023). While NF-KB and Nrf2 are involved in different cellular 396 processes, they have both been investigated as potential targets for the development of 397 hepatoprotective agents. Molecular docking studies have been used to investigate the 398 interaction of potential hepatoprotective agents with these transcription factors. There is also 399 evidence of crosstalk between the Nrf2 and NF- κ B pathways, which could be a new 400 therapeutic target against hepatotoxicity.

401 In the pathogenesis of hepatocellular damage, liver fibrosis, and HCC, NF-κB plays a 402 crucial role in regulating inflammation and cell death (Luedde and Schwabe 2011). In response 403 to many stimuli that may pose a threat to the host, NF-KB is activated, setting in motion 404 processes such as inflammation, immunity, wound healing, and pathogen clearance (Luedde 405 and Schwabe 2011). Pathogen-derived chemicals that activate Toll-Like Receptors (TLRs) 406 include lipopolysaccharide (LPS), viral and bacterial DNA and RNA, and inflammatory 407 cytokines including tumor necrosis factor (TNF) and interleukins (IL)-1 (Luedde and Schwabe 408 2011). When NF-B is activated, a lot of genes with B-binding sites are transcribed. These genes 409 play important roles in controlling inflammation, the immune response, and cell survival. In 410 an NF-kB-dependent manner, lipopolysaccharide (LPS) activates TLR4 on dormant HSCs by 411 reducing BAMBI expression (an inhibitory TGF-β pseudoreceptor) and increasing Kupffer cell 412 chemotaxis. Due to low levels of BAMBI, recruited Kupffer cells secrete TGF- β , which 413 stimulates HSCs unrestrictively. When HSCs have been activated, NF-KB serves a second 414 crucial function by increasing their chances of survival. LPS, Kupffer cell-derived mediators 415 (such as IL-1 and TNF), and angiotensin II, which is produced and acts on HSCs in an autocrine way, all play roles in activating NF-KB in activated hepatic stellate cells. More activated HSCs 416 417 and extracellular matrix are deposited in the liver as a result of greater HSC activation and 418 survival (Luedde and Schwabe 2011).

419 TNF- α and TGF- β can stimulate one another's production, and TNF- α has been shown to 420 influence TGF- β expression in a variety of cells and tissues (Liu et al. 2022). TNF- α is an 421 inflammatory cytokine that contributes to liver inflammation, and chronic liver inflammation 422 results in liver fibrosis. TNF-a exerts its effects on liver fibrosis via multiple mechanisms, 423 including TGF-β signaling activation (Yang and Seki 2015). Targeting TNF-α and TGF-β 424 signaling pathways may, therefore, have therapeutic potential for treating liver diseases. In 425 regards to hepatoprotective effects, the relationship between TGF-β and TNF-α is complex and 426 not completely understood.

427 CONCLUSION

In this study, the hepatoprotective qualities of Indonesian water kefir in rats with CCl4induced liver damage has evaluated. Water kefir administration improved the condition of liver damage, characterized by a decrease in serum levels of AST, ALT, TNF, TGF, and an improvement of liver tissue profile. *In silico* evaluation showed that the metabolites in water kefir were able to interact with target proteins in the NF-κB and Nrf2 pathways. It was concluded that water kefir improves the condition of the liver by reducing the level of necrosis and fibrosis.

435

436 CONFLICT OF INTEREST

437 The authors declare no conflict of interest.

438 ACKNOWLEDGMENT

439This research was funded by the Center of Research and Community Service, Bhakti Kencana University,440Bandung, West Java, Republic of Indonesia (052/14.LPPM/PE.I/LPPM/2021)

441 **REFERENCES**

442	A. Asnawi, Aman LO, Nursamsiar, et al (2022) Molecular Docking And Molecular Dynamic
443	Studies: Screening Phytochemicals Of Acalypha Indica Against Braf Kinase Receptors

- 444 For Potential Use In Melanocytic Tumours. RJC 15:1352–1361.
 445 https://doi.org/10.31788/RJC.2022.1526769
- Aligita W, Alex V, Taaraungan S, Susilawati E (2021) Hepatoprotective Activity Of Water
 Kefir. IJBPAS 10:. https://doi.org/10.31032/IJBPAS/2021/10.6.5493
- Aligita W, Tarigan PN, Susilawati E (2020) Anti Inflammatory And Antioxidant Activity Of
 Kefir Water. IJBPAS 9:. https://doi.org/10.31032/IJBPAS/2020/9.1.4904
- Al-Mohammadi A-R, Ibrahim RA, Moustafa AH, et al (2021) Chemical Constitution and
 Antimicrobial Activity of Kefir Fermented Beverage. Molecules 26:2635.
 https://doi.org/10.3390/molecules26092635
- Alsayadi M, Aljawfi Y, Belarbi M, Sabri FZ (2013) ANTIOXIDANT POTENCY OF WATER
 KEFIR. Journal of Microbiology, Biotechnology and Food Sciences 2:2444–2447
- Alsayadi M, Jawfi YA, Belarbi M, et al (2014) Evaluation of Anti-Hyperglycemic and Anti Hyperlipidemic Activities of Water Kefir as Probiotic on Streptozotocin-Induced
 Diabetic Wistar Rats. JDM 04:85–95. https://doi.org/10.4236/jdm.2014.42015
- Asnawi A, Febrina E, Aligita W, et al (2022) Screening Of Ashitaba (Angelica Keiskei K.)
 Compounds As Potential Mycobacterium Tuberculosis Kasa Inhibitors. Int J App
 Pharm 80–85. https://doi.org/10.22159/ijap.2022.v14s5.13
- 461 Asnawi A, Nedja M, Febrina E, Purwaniati (2023) Prediction of a Stable Complex of
 462 Compounds in the Ethanol Extract of Celery Leaves (Apium graveolens L.) Function
 463 as a VKORC1 Antagonist. TJNPR 7:2362–2370.
 464 https://doi.org/10.26538/tjnpr/v7i2.10
- Aspiras BEE, Flores RFAC, Pareja MC (2015) Hepatoprotective effect of Fermented Water Kefir
 on Sprague-Dawley rats (Rattus norvegicus) induced with sublethal dose of
 Acetaminophen. INT J CURR SCI 17:18–28
- 468 Calatayud M, Börner RA, Ghyselinck J, et al (2021) Water Kefir and Derived Pasteurized
 469 Beverages Modulate Gut Microbiota, Intestinal Permeability and Cytokine Production
 470 In Vitro. Nutrients 13:3897. https://doi.org/10.3390/nu13113897
- 471 Dai C, Li H, Wang Y, et al (2021a) Inhibition of Oxidative Stress and ALOX12 and NF-κB
 472 Pathways Contribute to the Protective Effect of Baicalein on Carbon Tetrachloride473 Induced Acute Liver Injury. Antioxidants 10:976.
 474 https://doi.org/10.3390/antiox10060976
- 475 Dai C, Li H, Wang Y, et al (2021b) Inhibition of Oxidative Stress and ALOX12 and NF-κB
 476 Pathways Contribute to the Protective Effect of Baicalein on Carbon Tetrachloride477 Induced Acute Liver Injury. Antioxidants (Basel) 10:976.
 478 https://doi.org/10.3390/antiox10060976
- 479 Dallakyan S, Olson AJ (2015) Small-Molecule Library Screening by Docking with PyRx. In:
 480 Hempel JE, Williams CH, Hong CC (eds) Chemical Biology. Springer New York, New
 481 York, NY, pp 243–250

- 482 Darvishzadeh P, Orsat V, Martinez JL (2021) Process Optimization for Development of a Novel
 483 Water Kefir Drink with High Antioxidant Activity and Potential Probiotic Properties
 484 from Russian Olive Fruit (Elaeagnus angustifolia). Food Bioprocess Technol 14:248485 260. https://doi.org/10.1007/s11947-020-02563-1
- 486 De Souza Basso B, Haute GV, Ortega-Ribera M, et al (2021) Methoxyeugenol deactivates
 487 hepatic stellate cells and attenuates liver fibrosis and inflammation through a PPAR-γ
 488 and NF-kB mechanism. Journal of Ethnopharmacology 280:114433.
 489 https://doi.org/10.1016/j.jep.2021.114433
- Diniz RO, Garla LK, Schneedorf JM, Carvalho JCT (2003) Study of anti-inflammatory activity
 of Tibetan mushroom, a symbiotic culture of bacteria and fungi encapsulated into a
 polysaccharide matrix. Pharmacological Research 47:49–52.
 https://doi.org/10.1016/S1043-6618(02)00240-2
- 494 Du X, Li Y, Xia Y-L, et al (2016) Insights into Protein–Ligand Interactions: Mechanisms,
 495 Models, and Methods. IJMS 17:144. https://doi.org/10.3390/ijms17020144
- 496 Elshaghabee FMF, Rokana N, Gulhane RD, et al (2017) Bacillus As Potential Probiotics: Status,
 497 Concerns, and Future Perspectives. Front Microbiol 8:1490.
 498 https://doi.org/10.3389/fmicb.2017.01490
- Febrina E, Alamhari RK, Abdulah R, et al (2021) MOLECULAR DOCKING AND
 MOLECULAR DYNAMICS STUDIES OF ACALYPHA INDICA L.
 PHYTOCHEMICAL CONSTITUENTS WITH CASPASE-3. Int J App Pharm 210–215.
 https://doi.org/10.22159/ijap.2021.v13s4.43861
- Febrina E, Asnawi A, Abdulah R, et al (2022) Identification Of Flavonoids From Acalypha
 Indica L. (Euphorbiaceae) As Caspase-3 Activators Using Molecular Docking And
 Molecular Dynamics. Int J App Pharm 162–166.
 https://doi.org/10.22159/ijap.2022.v14s5.34
- Fijan S (2014) Microorganisms with Claimed Probiotic Properties: An Overview of Recent
 Literature. IJERPH 11:4745-4767. https://doi.org/10.3390/ijerph110504745
- 509 Gamba RR, Yamamoto S, Sasaki T, et al (2019) Microbiological and Functional
 510 Characterization of Kefir Grown in Different Sugar Solutions. FSTR 25:303–312.
 511 https://doi.org/10.3136/fstr.25.303
- 512 Ganesh Yerra V, Negi G, Sharma SS, Kumar A (2013) Potential therapeutic effects of the
 513 simultaneous targeting of the Nrf2 and NF-κB pathways in diabetic neuropathy. Redox
 514 Biology 1:394–397. https://doi.org/10.1016/j.redox.2013.07.005
- Gao W, Guo L, Yang Y, et al (2022) Dissecting the Crosstalk Between Nrf2 and NF-κB Response
 Pathways in Drug-Induced Toxicity. Front Cell Dev Biol 9:809952.
 https://doi.org/10.3389/fcell.2021.809952
- Hong Y, Sheng L, Zhong J, et al (2021) Desulfovibrio vulgaris, a potent acetic acid-producing
 bacterium, attenuates nonalcoholic fatty liver disease in mice. Gut Microbes
 13:1930874. https://doi.org/10.1080/19490976.2021.1930874
- Ischak NI, Aman LO, Hasan H, et al (2023) In silico screening of Andrographis paniculata
 secondary metabolites as anti-diabetes mellitus through PDE9 inhibition. Res Pharm
 Sci 18:100–111. https://doi.org/10.4103/1735-5362.363616
- Ji Y, Xie Q, Meng X, et al (2022) Lactobacillus paracasei improves dietary fatty liver by reducing
 insulin resistance and inflammation in obese mice model. Journal of Functional Foods
 95:105150. https://doi.org/10.1016/j.jff.2022.105150
- Jiang Z-Y, Lu M-C, You Q-D (2019) Nuclear Factor Erythroid 2-Related Factor 2 (Nrf2)
 Inhibition: An Emerging Strategy in Cancer Therapy. J Med Chem 62:3840–3856.
 https://doi.org/10.1021/acs.jmedchem.8b01121
- Kaur G, Shivanandappa TB, Kumar M, Kushwah AS (2020) Fumaric acid protect the cadmiuminduced hepatotoxicity in rats: owing to its antioxidant, anti-inflammatory action and
 aid in recast the liver function. Naunyn-Schmiedeberg's Arch Pharmacol 393:19111920. https://doi.org/10.1007/s00210-020-01900-7
- Kebler LF (1921) "California Bees."11The author stated that the work on this subject is the
 result of investigation for the Post a c e Department relative to improper use of the
 mails in connection with this product. The Journal of the American Pharmaceutical
 Association (1912) 10:939–943. https://doi.org/10.1002/jps.3080101206
- Kim B, Kwon J, Kim M-S, et al (2018) Protective effects of Bacillus probiotics against high-fat
 diet-induced metabolic disorders in mice. PLoS ONE 13:e0210120.
 https://doi.org/10.1371/journal.pone.0210120
- Kondo T, Kishi M, Fushimi T, Kaga T (2009) Acetic Acid Upregulates the Expression of Genes
 for Fatty Acid Oxidation Enzymes in Liver To Suppress Body Fat Accumulation. J
 Agric Food Chem 57:5982–5986. https://doi.org/10.1021/jf900470c
- Konstantopoulos P, Doulamis I, Tzani A, et al (2017) Metabolic effects of Crocus sativus and
 protective action against non-alcoholic fatty liver disease in diabetic rats. biom rep.
 https://doi.org/10.3892/br.2017.884
- Laureys D, Aerts M, Vandamme P, De Vuyst L (2018) Oxygen and diverse nutrients influence
 the water kefir fermentation process. Food Microbiology 73:351–361.
 https://doi.org/10.1016/j.fm.2018.02.007
- Laureys D, De Vuyst L (2017) The water kefir grain inoculum determines the characteristics of
 the resulting water kefir fermentation process. J Appl Microbiol 122:719–732.
 https://doi.org/10.1111/jam.13370
- 553Laureys D, De Vuyst L (2014) Water kefir as a promising low-sugar probiotic fermented554beverage.ArchPublicHealth72:P1,2049-3258-72-S1-P1.555https://doi.org/10.1186/2049-3258-72-S1-P1
- Lee NY, Shin MJ, Youn GS, et al (2021) *Lactobacillus* attenuates progression of nonalcoholic
 fatty liver disease by lowering cholesterol and steatosis. Clin Mol Hepatol 27:110–124.
 https://doi.org/10.3350/cmh.2020.0125
- Lee TH, Kim WR, Poterucha JJ (2012) Evaluation of Elevated Liver Enzymes. Clinics in Liver
 Disease 16:183–198. https://doi.org/10.1016/j.cld.2012.03.006

- Li H, Weng Q, Gong S, et al (2023) Kaempferol prevents acetaminophen-induced liver injury
 by suppressing hepatocyte ferroptosis *via* Nrf2 pathway activation. Food Funct
 14:1884–1896. https://doi.org/10.1039/D2FO02716J
- Li L, He M, Xiao H, et al (2018) Acetic Acid Influences BRL-3A Cell Lipid Metabolism via the
 AMPK Signalling Pathway. Cell Physiol Biochem 45:2021–2030.
 https://doi.org/10.1159/000487980
- Li Y-T, Ye J-Z, Lv L-X, et al (2019a) Pretreatment With Bacillus cereus Preserves Against DGalactosamine-Induced Liver Injury in a Rat Model. Front Microbiol 10:1751.
 https://doi.org/10.3389/fmicb.2019.01751
- Li Z, Huang Y, Wu Y, et al (2019b) Absolute Binding Free Energy Calculation and Design of a
 Subnanomolar Inhibitor of Phosphodiesterase-10. J Med Chem 62:2099–2111.
 https://doi.org/10.1021/acs.jmedchem.8b01763
- 573Liu Z, Zhang Y, Zhang L, et al (2022) Duality of Interactions Between TGF-β and TNF-α During574TumorFormation.FrontImmunol12:810286.575https://doi.org/10.3389/fimmu.2021.810286
- 576Luedde T, Schwabe RF (2011) NF-κB in the liver—linking injury, fibrosis and hepatocellular577carcinoma.NatRevGastroenterolHepatol8:108–118.578https://doi.org/10.1038/nrgastro.2010.213
- 579 Lynch KM, Wilkinson S, Daenen L, Arendt EK (2021) An update on water kefir: Microbiology,
 580 composition and production. International Journal of Food Microbiology 345:109128.
 581 https://doi.org/10.1016/j.ijfoodmicro.2021.109128
- 582Meng X-Y, Zhang H-X, Mezei M, Cui M (2011) Molecular Docking: A Powerful Approach for583Structure-BasedDrugDiscovery.CAD7:146-157.584https://doi.org/10.2174/157340911795677602
- Moinas M, Horisberger M, Bauer H (1980) The structural organization of the Tibi grain as
 revealed by light, scanning and transmission microscopy. Arch Microbiol 128:157–161.
 https://doi.org/10.1007/BF00406153
- Moreira MEC, Santos MHD, Zolini GPP, et al (2008) Anti-Inflammatory and Cicatrizing
 Activities of a Carbohydrate Fraction Isolated from Sugary Kefir. Journal of Medicinal
 Food 11:356–361. https://doi.org/10.1089/jmf.2007.329
- Neag MA, Catinean A, Muntean DM, et al (2020) Probiotic Bacillus Spores Protect Against
 Acetaminophen Induced Acute Liver Injury in Rats. Nutrients 12:632.
 https://doi.org/10.3390/nu12030632
- Nursamsiar, Nur S, Febrina E, et al (2022) Synthesis and Inhibitory Activity of Curculigoside
 A Derivatives as Potential Anti-Diabetic Agents with β-Cell Apoptosis. Journal of
 Molecular Structure 1265:133292. https://doi.org/10.1016/j.molstruc.2022.133292
- Parasuraman S, Raveendran R, Kesavan (2010) Blood sample collection in small laboratory
 animals. Journal of Pharmacology and Pharmacotherapeutics 1:87–93.
 https://doi.org/10.4103/0976-500X.72350

- Patel SH, Tan JP, Börner RA, et al (2022) A temporal view of the water kefir microbiota and
 flavour attributes. Innovative Food Science & Emerging Technologies 80:103084.
 https://doi.org/10.1016/j.ifset.2022.103084
- 603 Pidoux M (1989) Kefir Grain Flora. Mircen Journal 5:223–228
- Rahman ZU, Al Kury LT, Alattar A, et al (2021) Carveol a Naturally-Derived Potent and
 Emerging Nrf2 Activator Protects Against Acetaminophen-Induced Hepatotoxicity.
 Front Pharmacol 11:621538. https://doi.org/10.3389/fphar.2020.621538
- Ritesh KR, Suganya A, Dileepkumar HV, et al (2015) A single acute hepatotoxic dose of CCl 4
 causes oxidative stress in the rat brain. Toxicology Reports 2:891–895.
 https://doi.org/10.1016/j.toxrep.2015.05.012
- Rocha-Gomes A, Escobar A, Soares JS, et al (2018) Chemical composition and
 hypocholesterolemic effect of milk kefir and water kefir in Wistar rats. Rev Nutr
 31:137-145. https://doi.org/10.1590/1678-98652018000200001
- Rodrigues KL, Araújo TH, Schneedorf JM, et al (2016) A novel beer fermented by kefir
 enhances anti-inflammatory and anti-ulcerogenic activities found isolated in its
 constituents. Journal of Functional Foods 21:58-69.
 https://doi.org/10.1016/j.jff.2015.11.035
- Romero-Luna HE, Peredo-Lovillo A, Hernández-Mendoza A, et al (2020) Probiotic Potential
 of Lactobacillus paracasei CT12 Isolated from Water Kefir Grains (Tibicos). Curr
 Microbiol 77:2584–2592. https://doi.org/10.1007/s00284-020-02016-0
- Semjonovs P, Denina I, Linde R (2014) Evaluation of Physiological Effects of Acetic Acid
 Bacteria and Yeast Fermented Non-alchocolic Beverage Consumption in Rat Model. J
 of Medical Sciences 14:147–152. https://doi.org/10.3923/jms.2014.147.152
- Sharifi-Rad J, Seidel V, Izabela M, et al (2023) Phenolic compounds as Nrf2 inhibitors: potential
 applications in cancer therapy. Cell Commun Signal 21:89.
 https://doi.org/10.1186/s12964-023-01109-0
- Šilhavý J, Zídek V, Mlejnek P, et al (2014) Fumaric Acid Esters Can Block Pro-Inflammatory
 Actions of Human CRP and Ameliorate Metabolic Disturbances in Transgenic
 Spontaneously Hypertensive Rats. PLoS ONE 9:e101906.
 https://doi.org/10.1371/journal.pone.0101906
- 630 Tsai Y-S, Lin S-W, Chen Y-L, Chen C-C (2020) Effect of probiotics Lactobacillus paracasei GKS6, 631 L. plantarum GKM3, and L. rhamnosus GKLC1 on alleviating alcohol-induced alcoholic 632 liver Nutr 14:299. disease in а mouse model. Res Pract 633 https://doi.org/10.4162/nrp.2020.14.4.299
- Wang B, Cui S, Mao B, et al (2022a) Cyanidin Alleviated CCl4-Induced Acute Liver Injury by
 Regulating the Nrf2 and NF-κB Signaling Pathways. Antioxidants 11:2383.
 https://doi.org/10.3390/antiox11122383
- Wang Y, Liu F, Liu M, et al (2022b) Curcumin mitigates aflatoxin B1-induced liver injury via
 regulating the NLRP3 inflammasome and Nrf2 signaling pathway. Food and Chemical
 Toxicology 161:112823. https://doi.org/10.1016/j.fct.2022.112823

- 640 Ward M (1892) V. The ginger-beer plant, and the organisms composing it: a contribution to
 641 the study of fermentation-yeasts and bacteria. Phil Trans R Soc Lond B 183:125–197.
 642 https://doi.org/10.1098/rstb.1892.0006
- 643 Weber LWD, Boll M, Stampfl A (2003) Hepatotoxicity and Mechanism of Action of
 644 Haloalkanes: Carbon Tetrachloride as a Toxicological Model. Critical Reviews in
 645 Toxicology 33:105–136. https://doi.org/10.1080/713611034
- Kue J, Shen K, Hu Y, et al (2020) Effects of dietary Bacillus cereus, B. subtilis, Paracoccus marcusii, and Lactobacillus plantarum supplementation on the growth, immune response, antioxidant capacity, and intestinal health of juvenile grass carp (Ctenopharyngodon idellus). Aquaculture Reports 17:100387.
 https://doi.org/10.1016/j.aqrep.2020.100387
- Yang H, Meng L, Ai D, et al (2019) Acetic acid alleviates the inflammatory response and liver
 injury in septic mice by increasing the expression of TRIM40. Exp Ther Med.
 https://doi.org/10.3892/etm.2019.7274
- 654 Yang YM, Seki E (2015) TNFα in Liver Fibrosis. Curr Pathobiol Rep 3:253–261.
 655 https://doi.org/10.1007/s40139-015-0093-z
- Yao F, Jia R, Huang H, et al (2019) Effect of Lactobacillus paracasei N1115 and
 fructooligosaccharides in nonalcoholic fatty liver disease. aoms 15:1336–1344.
 https://doi.org/10.5114/aoms.2019.86611
- Yeh Y-H, Hsieh Y-L, Lee Y-T, Hsieh C-H (2011) Protective effects of cholestin against carbon
 tetrachloride-induced hepatotoxicity in rats. e-SPEN, the European e-Journal of
 Clinical Nutrition and Metabolism 6:e264–e271.
 https://doi.org/10.1016/j.eclnm.2011.09.002
- Zamberi NR, Abu N, Mohamed NE, et al (2016) The Antimetastatic and Antiangiogenesis
 Effects of Kefir Water on Murine Breast Cancer Cells. Integr Cancer Ther 15:NP53NP66. https://doi.org/10.1177/1534735416642862
- Zavala L, Golowczyc MA, van Hoorde K, et al (2016) Selected *Lactobacillus* strains isolated
 from sugary and milk kefir reduce *Salmonella* infection of epithelial cells *in vitro*.
 Beneficial Microbes 7:585–595. https://doi.org/10.3920/BM2015.0196
- Chao H, Eguchi S, Alam A, Ma D (2017a) The role of nuclear factor-erythroid 2 related factor
 2 (Nrf-2) in the protection against lung injury. Am J Physiol Lung Cell Mol Physiol
 312:L155–L162. https://doi.org/10.1152/ajplung.00449.2016
- 672 Zhao H, Eguchi S, Alam A, Ma D (2017b) The role of nuclear factor-erythroid 2 related factor
 673 2 (Nrf-2) in the protection against lung injury. Am J Physiol Lung Cell Mol Physiol
 674 312:L155-L162. https://doi.org/10.1152/ajplung.00449.2016

Bandung, September 19, 2023

Dear **Prof. Gabino Garrido** Editor in Chief Journal of Pharmacy & Pharmacognosy Research Garval Editorial Ltda. Antofagasta Chile

Thank you for giving us the opportunity to submit a revised draft of our manuscript (ID: JPPRes-23-1732.R1) titled *Hepatoprotective Study of Indonesian Water Kefir Against CCl4-Induced Liver Injury* in *Rats* to *Journal of Pharmacy & Pharmacognosy Research (JPPRes)*. We appreciate the time and effort that you and the reviewers have dedicated to providing your valuable feedback on our manuscript. We are grateful to the reviewers for their insightful comments on our paper. We have been able to incorporate changes to reflect most of the suggestions provided by the reviewers. We have highlighted the changes within the manuscript.

	Comment	Response
Keywords	Please check if these keywords have been accepted	1. We have
	by MeSH Browser 2022 or 2023 (https://meshb-	re-
	prev.nlm.nih.gov/). If not, type keywords accepted	checked
	by this MeSH Browser.	the
		keywords.
Introduction	1. There are numerous studies demonstrating the hepatoprotective activity of kefir. Therefore, the	1. We have added the
	authors should state here the novel effect they intend to find, which has not been reported in	research problem
	the scientific literature. Please better state the	2. We have
	research problem and how you intend to solve it.	revised the
	2. Please move this last sentence to the	last
	Conclusions, it is not valid for an Introduction.	sentence
MATERIAL AND	1. Experimental sample and reference extract	We have
METHODS	It is necessary to indicate here where these materials	revised the
	were obtained from. Indicate who identified the	research
	material. The manuscript should include references	methodology.
	to voucher specimens of the plants (deposited in a	
	major regional herbarium) or to the material	
	examined, including their registration number(s). It	
	should be mentioned which parts of the plant have	
	been used. The GPS coordinates of the collection site	
	of the species should also be indicated. It is	
	mandatory that the authors indicate how both the	
	kefir and the turmeric extract were prepared, the	
	quality of these products, etc.	
	2. Please indicate how the serum was extracted in	
	the rats? from which part of the animal and how the	
	blood was extracted. How were AST, TNF- α , TGF-a	
	levels measured? give details of this. How was the	

Here is a point-by-point response to the reviewers' comments and concerns

Results	 liver extracted from the animals? how was the liver histopathological analysis performed. 1. Please, it is mandatory to add a reference NF-kB inhibitor/activator. 2. Comment about reference compound and compare. 3. Add a reference compound 	We have revised the Results (Table 1, Figs 4 and 5)	
Bibliography/References :	1. The style of the References does not meet the requirements of JPPRes. Please write the References according to the Instructions to the Author or the examples: https://jppres.com/jppres/archive/ https://jppres.com/jppres/volume-11-issue-2/ Also, all authors and DOIs must be given. Please check that all references in this section have been cited in the text and vice versa. Please check that the journals cited are prestigious in the discipline and are not predatory journals (https://www.openacessjournal.com/blog/predatory -journals-list/#I_%E2%80%93_predatory_journals). References must be write free of codes from any bibliography processor, such as Endnotes, Mendeley, Zotero, etc2. Latest references must be used.	 We have revised the reference s referring to the sample. 	

Thank you, and we look forward to hearing more about our manuscript.

With kind regards,

Widhya Aligita

Faculty of Pharmacy, Bhakti Kencana University, Jl. Soekarno Hatta no 754, Bandung, West Java, Indonesia

E-mail: widhya.aligita@bku.ac.id

1	Hepatoprotective study of Indonesian water kefir against				
2	CCl ₄ -induced liver injury in rats				
3					
4	Ru	nning title: Hepatoprotective	e study of water kefir		
5	Widhya	n Aligita ^{1,3*} , Marlia Singgih ² , Entris	Sutrisno ³ , I Ketut Adnyana	1*	
67 89 10 11 12 13 14	¹ Department of Pharmacology & Clinical Pharmacy, School of Pharmacy, Bandung Institute of Technology, Bandung, Indonesia. ² Department of Pharmacochemistry, School of Pharmacy, Bandung Institute of Technology, Bandung, Indonesia. ³ Department of Pharmacology & Clinical Pharmacy, Faculty of Pharmacy, Bhakti Kencana University, Bandung, Indonesia. *E-mail address: widhya.aligita@bku.ac.id / ketut@itb.ac.id *Corresponding author address: Department of Pharmacology & Clinical Pharmacy, School of Pharmacy, Bandung Institute of Technology, Ganeca, 10, 40132, Bandung, Indonesia. Tel: +62-22-2504852				
15 16 17	The institutional e-mail of authors):	the ALL authors must be declared ((and add rows according to t	he number of	
	Author	Institutional e-mail (mandatory)	Other e-mail	ORCID	

autions).			
Author	Institutional e-mail (mandatory)	Other e-mail (gmail, yahoo, etc.)	ORCID (0000-1111-2222-3333)
Widhya Aligita	widhya.aligita@bku.ac.id	w.aligita@gmail.com	0000-0001-8338-4115
Marlia Singgih	marlia@fa.itb.ac.id		0000-0002-5351-1731
Entris Sutrisno	entris.sutrisno@bku.ac.id		0000-0003-3830-6411
I Ketut Adnyana	ketut@itb.ac.id		0000-0001-5217-2312

Contribution Details (to be ticked marked (X) as applicable and add columns according to the number of authors): 20

Contribution	Aligita W	Singgih M	Sutrisno E	Adnyana IK
Concepts or Ideas	x	x	х	х
Design	х	х	х	х
Definition of intellectual content	х	х	х	х
Literature search	х			х
Experimental studies	х			х
Data acquisition	х			Х
Data analysis	х			Х
Statistical analysis	х			Х
Manuscript preparation	х	х	Х	Х
Manuscript editing	х	х	Х	х
Manuscript review	х	х	х	х

23 ABSTRACT

- 24 25 26 Context: Water kefir is a fermented beverage that is typically made in the home by inoculating a sugar-rich solution with a microbial community (water kefir grains). Several studies on the metabolite content and hepatoprotective effects of water kefir have been published, but CCl4-induced acute liver injury has not been studied.
- 27 28 Objectives: To evaluate the efficacy of water kefir in vivo against hepatoprotective CCl4-induced acute liver injury and to in silico investigate metabolites that play an important role in hepatoprotective mechanisms.
- Methods: The present study aimed to investigate the hepatoprotective activity of water kefir in an animal model caused by carbon tetrachloride (CCl4). Furthermore, using molecular docking, the metabolites found in water kefir were evaluated for their role in the NF-KB and Nrf2 signaling pathways.
- 29 31 32 33 4 35 36 37 8 940 Results: Water kefir significantly and dose-dependently alleviated acute liver injury caused by carbon tetrachloride (CCl4). Water kefir administration at all doses produced results comparable to the positive control (Curcuma extract). Molecular docking simulations showed that, compared to Nrf2, the 25 metabolites were more likely to interact with the NF-B receptor. Fumaric acid is the strong metabolite that interacts with the NF-KB receptor with a free energy of binding and an inhibition constant of -6.66 kcal/mol and 13.22 µM, respectively.
- Conclusions: Water kefir administration improved the condition of liver damage, characterized by a decrease in serum levels of AST, ALT, TNF-α, TGF-β, and an improvement in the liver tissue profile. In silico evaluation showed that the metabolites in water kefir were able to interact with target proteins in the NF-κB and Nrf2 pathways. It was concluded that water kefir improves the condition of the liver by reducing the level of necrosis and fibrosis.
- 41
- 42
- Keywords: free radicals; liver diseases; kefir; molecular docking simulation; probiotics
- 43

44 **INTRODUCTION**

45 In most cases, making water kefir involves combining dried fruit, sugar, and water kefir grains in a container. Water kefir's exact origins are unknown; however, two hypotheses have 46 47 been proposed regarding its history: the first suggests that water kefir grains were brought to 48 Europe from the Caucasus by soldiers returning from the Crimean War in the late nineteenth 49 century (Ward, 1892); the second theory proposes that water kefir grains originated in Mexico 50 from the Opuntia cactus through natural processes (Moinas et al., 1980). Sugary kefir grains, 51 Balm of Gilead, African bees, Japanese beer seeds, Ale nuts, and California bees are some other 52 names for water kefir. Tibi grains and ginger beer plants are other names for water kefir 53 (Kebler, 1921; Moinas et al., 1980; Pidoux, 1989). Water kefir is appealing to both consumers 54 and researchers due to the variety of microbiota it contains, the fact that it is an alternative to 55 dairy products, the versatility with which it can be flavored, the fact that it is low in calories 56 and sugar, the ease with which it can be produced, and the health benefits it offers.

57 Water kefir has been used medicinally for a very long time, and recent research has 58 indicated that it may have a variety of positive effects on people's health. It has been 59 demonstrated that water kefir contains non-pathogenic bacteria that, in conjunction with the 60 production of organic acids, can inhibit the growth of pathogenic microbes such as *Shigella sp.*, 61 Salmonella sp., Staphylococcus aureus, and E. coli; as well as, filamentous fungi such as Aspergillus 62 ochraceus, A. niger, A. flavus, Penicillium sp., and Rhizopus sp. (Al-Mohammadi et al., 2021). In addition to its antibacterial properties, water kefir possesses a broad spectrum of 63 64 pharmacological effects. Some of these therapeutic effects are anti-inflammatory (Diniz et al., 65 2003; Aligita et al., 2020), antioxidant (Alsavadi et al., 2013; Aligita et al., 2020; Darvishzadeh 66 et al., 2021), hepatoprotective (Aspiras et al., 2015; Aligita et al., 2021), antihyperglycemic and 67 antihyperlipidemic (Alsayadi et al., 2014; Rocha-Gomes et al., 2018), anti-edematous (Moreira 68 et al., 2008), antitumor (Zamberi et al., 2016), antihypertensive (Gamba et al., 2019), immunomodulant (Calatayud et al., 2021), and anti-ulcerogenic (Rodrigues et al., 2016). 69 70 However, no studies have been reported on the hepatoprotective effects of water kefir against 71 carbon tetrachloride (CCl₄)-induced liver injury.

72 Studies have shown that acute liver injury is frequently accompanied by high levels of 73 oxidative stress and inflammatory responses (Dai et al., 2021a). These findings have been 74 found in several studies. The most important signaling pathways that are involved in the 75 regulation of inflammation and antioxidation are the nuclear factor (NF-KB) and nuclear factor related factor 2 (Nrf2) pathways, respectively. It has been shown that activating
Nrf2 and inhibiting NF-κB can reduce the amount of damage done to the liver. For instance,
curcumin protects against aflatoxin B1-induced liver injury by increasing the expression of
Nrf2 and related downstream antioxidant molecules (such as superoxide dismutase (SOD),

80 catalase (CAT), heme oxygenase 1 (HO-1), and NAD(P)H quinone dehydrogenase 1 (NQO-1)

81 (Wang et al., 2022b). Also, it has been shown that the molecule methoxy eugenol, which comes

- 82 from nutmeg and Brazilian red propolis, protects the liver both *in vitro* and *in vivo*. This may
- 83 be attributed to the fact that it targets the NF-κB signaling pathway, which has been shown to
- 84 have anti-inflammatory effects (De Souza Basso et al., 2021).

85 Ethanol and lactic acid are the primary metabolites found in fermented water kefir, while 86 lower quantities of glycerol, mannitol, and acetic acid can also be found in the beverage. 87 Additionally, a variety of aromatic and volatile compounds are produced, including ethyl 88 acetate, isoamyl acetate, ethyl octanoate, ethyl hexanoate, and ethyl decanoate, among others 89 (compared to their threshold levels) (Laureys and De Vuyst 2014). The chemical constituents 90 of both phytochemicals and secondary metabolites in natural products, including water kefir, 91 are certainly capable of providing various pharmacological effects for the body (A. Asnawi et 92 al., 2022; Nursamsiar et al., 2022). However, an in silico study to evaluate the metabolite content 93 in water kefir has not been reported yet. Because of its capacity to speed up the process of 94 identifying and optimizing lead compounds, the in silico method has become the front-runner 95 in the race to improve the speed and accuracy of the process of discovering new drugs. This is 96 because the *in silico* method can identify and optimize lead compounds more quickly. 97 Techniques such as molecular docking and molecular dynamics (MD) were able to directly 98 indicate a small number of compounds that have high affinity and selectivity by analyzing 99 how the ligand and target interact with one another (Febrina et al., 2021).

Water kefir has been used for an extensive period of time and has been recognized for its widespread benefits, especially in Indonesia. However, its level of popularity falls short in comparison to that of milk kefir. Furthermore, there is a scarcity of studies specifically focused on the hepatoprotective activity of water kefir made with Indonesian grains. Therefore, the purpose of this study was to evaluate the hepatoprotective effects of water kefir in CCl₄induced rats while also investigating the stability interactions of its metabolites within the NF-KB and Nrf2 receptors using molecular docking studies.

Please place a comma before the year in all citations throughout the document. See
 examples of already arranged citations in the Introduction.

109 MATERIAL AND METHODS

110 Materials and reagents

Water kefir grains, sugar, dried fruit, carbon tetrachloride, rats, diagnostic kits for alanine
aminotransferase (ALT) (Proline, IFCC mod.), aspartate aminotransferase (AST) (Proline,
IFCC mod.), Elisa Kit TNF-α (Bioassay Technology Laboratory), Elisa Kit TGF-β (Bioassay
Tachnalagy Laboratory) Other chemicals used in this study users of analytical response fractory

114 Technology Laboratory). Other chemicals used in this study were of analytical reagent grade.

115 Experimental sample and reference extract

The water kefir grain was obtained from Yogyakarta, Indonesia. The water kefir solution was produced using a fermentation procedure. The initial stage involved the preparation of 60 g of sugar, 2 g of raisins, 100 g of water kefir grains, and 1 L of aqua mineral distillate. The sugar and warm distilled water were mixed in a beaker, followed by the addition of water kefir grains and raisins to the resulting sugar solution. The fermentation procedure was conducted over a duration of forty-eight hours at ambient temperature. A small cloth was used to cover the beaker glass. The kefir grain was utilized in future production, while the filtratewas employed for the purpose of evaluation and analysis. (Aligita et al. 2020, 2021)

124 The rhizoma extract of Curcuma (*Curcuma xanthorrhiza* Roxb) is employed as a reference 125 drug. The utilized product is a standardized herbal medicine with the brand name Tulak, 126 manufactured by the Borobudur Company Herbal Medicine Industry. The primary purpose 127 of Tulak capsules is to support and preserve optimal liver functionality.

128 Animals and experimental design

129 Rats (Wistar strain, male, 200-250 g) were maintained on normal pellet food and tap water 130 ad libitum. Four mice in each group were used. All procedures relating to animals and their 131 care conformed to the international guidelines Principles of Laboratory Animal Care (NIH 132 publication no. 85-23, revised 1985) with the ethical approval number 133 363/UN6.KEP/EC/2020. In order to develop an animal model with liver injury, the rats 134 received CCl₄ (20% in olive oil) at 1.25 mL/kg BW every 2 days via a gavage tube (Yeh et al. 135 2011). The rats were randomized into five groups after the development of animals with liver 136 injury, which is characterized by a significant increase in serum ALT level, as follows: (1) 137 positive control group, (2) Curcuma extract group 100 mg/kg BW, (3) water kefir 15 mL/kg 138 BW group, (4) water kefir 35 mL/kg BW group, and (5) water kefir 50 mL/kg BW group, with 139 the addition (6) negative control group. Each group received group-specific treatment for two 140 weeks, along with the administration of 1.25 mL/kg BW of CCl₄ (20% in olive oil) every three 141 days.

142 The rats, which had undergone a fasting period of 8-10 hours while being provided with 143 water, had their blood samples collected via the retro-orbital sinus using a hematocrit capillary 144 tube. The blood sample was taken into a microtube and afterwards then to centrifugation. The 145 serum was separated in order to facilitate further measurements (Parasuraman et al. 2010). 146 Serum ALT level, as the main parameter, was measured prior to induction, following 147 induction, and following treatment. Meanwhile, following therapy, serum AST, TNF-a, TGF-148 β levels, and liver histopathological were evaluated. The ALT, AST, TNF- α , TGF- β levels 149 measurements are conducted in accordance with the protocols outlined in the reagent kit.

After the euthanasia procedure, the liver specimen was promptly immersed in a 10% formalin solution at room temperature for a duration of 24 hours. Subsequently, the tissues were immersed in paraffin, subjected to sectioning, and affixed onto glass microscope slides. The slices underwent staining with hematoxylin and eosin and were afterwards analyzed using light microscopy (Konstantopoulos et al. 2017).

155 Molecular docking simulation

156 Molecular docking experiments were done with the PyRx software (Dallakyan and Olson 157 2015) to predict how metabolites, which are small-molecule ligands, bind to biological 158 macromolecules. The NCBI PubChem database (https://pubchem.nlm.nih.gov/, accessed on 159 3 May 2023) was used to derive the three-dimensional structure of water kefir metabolites 160 (Patel et al. 2022). Target proteins like NF-κB (PDB code: 1A3Q) and Keap1 (PDB code: 4L7B) 161 were obtained from the RCSB Protein Data Bank (http://www.rcsb.org/, accessed on 03 May 2023). The PyRx program was used to reduce protein, ligand converted to PDBQT (Asnawi et 162 163 al. 2023), then maximize GRID parameter (Asnawi et al. 2022) and perform docking study (Febrina et al. 2022). The BIOVIA Discovery Studio 2017 R2 program was used to view the 164 165 protein and ligand complex and distance (Ischak et al. 2023). The BIOVIA Discovery Studio 166 2017 R2 tool was also utilized to find protein active sites.

167 Statistical analysis

All of the information is displayed in the form of individual data points as well as the mean along with the standard error of the mean. The statistical analysis was carried out with the help of Minitab software (version 19.0), and to make comparisons between several different groups, a one-way analysis of variance with Dunnett's post hoc test was utilized. All statistical graphs were created with Microsoft Excel 2019 in their respective versions. The levels of significance that were considered to have been reached were *p < 0.05.

174 **RESULTS**

175 In vivo evaluation of hepatoprotective activity

176 The serum ALT levels, as the main parameter for the liver damage, were measured prior to 177 and following induction, as well as following treatment (Fig. 1). Meanwhile, after treatment 178 AST, TNF-α, and TGF-β levels were also evaluated. These findings were analyzed using a one-179 way ANOVA with a 95% confidence level and a two-sided confidence interval. A significant 180 rise in blood ALT levels demonstrated the establishment of an animal model with liver injury, 181 according to statistical analysis, following the administration of CCl₄. When compared to the 182 positive control group, ALT serum levels decreased significantly after two weeks of therapy 183 with curcuma extract or water kefir. The three doses of water kefir groups demonstrated 184 equivalent activity when curcuma extract was used as the standard treatment, and there was 185 no significant difference between the three doses of water kefir. When compared to the 186 positive control group, AST levels were also reduced dramatically following treatment with 187 curcuma extract or water kefir. TNF-g levels in the water kefir group were significantly lower 188 than in the positive control group at dosages of 35 and 50 mL/kg body weight. Even though 189 there was no statistically significant difference in TGF-β levels, the group that received the 190 treatment demonstrated a decrease in TGF-β levels.



Fig. 1. (a) Before induction, after induction, and after treatment serum ALT level; (b) After treatment serum AST level; (c) After treatment serum TNF- α level; (d) After treatment serum TGF- β level. * indicates a significantly different result when compared to the positive control group; # indicates a significantly different result when compared to the curcuma extract group; p < 0.05; n = 4 mice in each group.

192 A histological examination of a normal liver group revealed a typical central vein bordered 193 with endothelial cells and normal hepatocytes, resulting in hepatic cords of cells with distinct 194 cell borders and sinusoidal gaps (Figs 2A and 2B). The CCl4-induced group developed 195 centrilobular hepatic necrosis level 4, which was characterized by massive vacuolization and 196 necrosis of epithelial cells in the liver's centrilobular region (blue arrow) (Fig 2C & 2D). The 197 group that received either curcuma extract or water kefir treatment improved in varied 198 necrotic conditions ranging from level 1 (water kefir 50 mL/kg BW) (Fig 2K & 2L) to level 2 199 (curcuma extract (Fig 2E & 2F), water kefir 15 mL/kg BW (Figs 2G and 2H), and water kefir 200 30 mL/kg BW (Fig 2I & 2J)).



Fig. 2. Liver histology after CCl4 intoxication and treatment with HE staining. Negative control group (A & B), positive control group (C & D), curcuma extract group (E & F), water kefir 15 mL/kg BW (G & H), water kefir 30 mL/kg BW (I & J), and water kefir 50 mL/kg BW (K & L). 1: central vein; 2: normal liver epithelial cell; 3: portal track; blue arrow: necrosis and liver epithelial cell vacuolization at centrolobular region.

202 Molecular docking

203 Molecular docking studies are considered a powerful tool for predicting the potential 204 targets of bioactive molecules. In order to carry out molecular docking simulations, one of the 205 most critical steps is to identify the target active site. If the target protein is crystallized with a 206 native ligand, in many instances, the location of the active site can be established without any 207 difficulty (Li et al. 2019b). However, the NF-kB (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) 208 proteins do not have a native ligand, so the active site was determined. Active site prediction 209 in docking is a computational method for predicting the location and orientation of a receptor 210 protein's binding site for a ligand molecule. The active site prediction was based on a protein 211 structural analysis and the identification of amino acid residues that are likely to interact with 212 the ligand. The projected binding site is then utilized as a starting point for molecular docking, 213 a computer method for predicting a ligand molecule's binding affinity and orientation to a 214 receptor protein. The active site prediction for target proteins (Keap1 and NF-κB) gives the 215 grid box coordinates (x y z) of 17.500880 Å, 62.323000 Å, and 0.973748 Å; and -1.751769 Å, -20.743853 Å, and -29.010438 Å, respectively (Fig. 3). 216



Fig. 3. Binding pocket (colored in red) generated using the BIOVIA Discovery Studio 2017 tool, coupled with the sequence containing the highlighted residues that constitute the binding pocket. A) NF-κB (PDB ID 1A3Q) and B) Keap1 (PDB ID 4L7B).

217

The docking results of the 25 metabolites could interact with target proteins (Keap1 and NF- κ B) (Table 1). In general, all metabolites could interact with both NF- κ B receptors (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) (Table 1). The interaction of metabolites with nuclear factor-kappa B receptors (PDB ID 1A3Q) revealed that 13 metabolites possessed free energy for binding that was greater than that of curcumin. On the other hand, there was not a single metabolite that had a free energy binding that was more powerful than that of nrf2 Keap1 (PDB ID 4L7B) (Table 1).

225 For volatile compounds, 2-phenyl ethanol and benzaldehyde interact most strongly with 226 the NF-kB receptor (PDB ID 1A3Q) with almost the same binding energy of -4.19 kcal/mol. 227 As for organic acids, succinic acid, fumaric acid, and citric acid provide nearly the same strong 228 interactions. Bond energy values of fumaric acid, succinic acid, and citric acid of -6.66, -6.24, 229 and -6.25 kcal/mol, respectively. In sugars, glucose provides the strongest interaction with the 230 NF-KB receptor (PDB ID 1A3Q) where the binding energy was -3.71 kcal/mol. As for the nrf2 231 Keap1 receptor (PDB ID 4L7B), 2-phenylacetaldehyde from volatile compounds was able to 232 interact most strongly with the Keap1 receptor (PDB ID 4L7B) with a binding energy of -3.46 233 kcal/mol. As for organic acids, succinic acid provides nearly the same strong interaction with a bond energy value of -3.02 kcal/mol, respectively. In sugars, glucose provides the strongest interaction with the Keap1 receptor (PDB ID 4L7B) where the binding energy was -3.09

236 kcal/mol.

- 237
- 238

Table 1. The free energy of binding and inhibition constant of water kefir metabolite active site interaction on NF κB (PDB ID 1A3Q) and Keap1 (PDB ID 4L7B)

No.	Metabolites	PDB: 1A3O		PDB: 4L7B	
		Free Energy of Binding, ∆G (kcal/mol)	Inhibition Constant, Ki (µM)	Free Energy of Binding, ∆G (kcal/mol)	Inhibition Constant, Ki (μΜ)
	Volatile compounds				
1	2-Methylbutanol	-3.31	3,770	-2.68	10,890
2	3-Methylbutanol	-3.22	4,360	-2.50	14,670
3	2-Phenylethanol	-4.19	851.52	-3.12	5,140
4	Ethyl acetate	-3.19	4,590	-2.43	16,670
5	2-Methylbutanal	-3.26	4,050	-2.35	19,050
6	1-Octanol	-2.65	11,490	-2.11	28,220
7	2-Phenylethyl acetate	-3.99	1,190	-3.33	3,590
8	Benzyl acetate	-4.16	887.66	-3.44	3,020
9	Benzaldehyde	-4.19	853.74	-3.19	4,550
10	Furfuryl acetate	-3.60	2,320	-3.32	3,660
11	Isobutyl acetate	-3.19	4,610	-2.94	7,020
12	Isopentyl acetate	-3.20	4,510	-3.05	5,820
13	2-Phenylacetaldehyde	-4.07	1,030	-3.46	2,930
14	3-Methylbutanal	-3.16	4,850	-2.73	10,020
15	Gluconic acid	-3.11	5,280	-1.63	64,290
	Organic acids				
16	Lactic acid	-4.92	249.28	-2.06	30,680
17	Succinic acid	-6.24	26.68	-3.02	6,140
18	Fumaric acid	-6.66	13.22	-2.45	16,100
19	Citric acid	-6.25	26.18	-1.87	42,440
20	Malic acid	-5.65	72.69	-2.53	13,860
21	Acetic acid	-4.99	219.33	-2.44	16,280
22	Ethanol	-2.39	17,800	-2.08	29,800
	Sugars				
23	Sucrose	-1.40	93,710	-1.33	105,320
24	Glucose	-3.71	1,900	-3.09	5,440
25	Fructose	-3.21	4,400	-2.65	11,440
	Reference compound				
	Curcumin	-3.44	2,990	-4.22	811.36

²⁴¹

242 The theoretical binding modes of the top three metabolites with their target proteins (Keap1 243 and NF-KB) are shown in Figs. 4 and 5, respectively. The molecular docking results suggested 244 that these metabolites interacted with the Keap1 and NF-KB to form a complex through 245 hydrogen bonds with various residues. The interaction of 2-phenyl ethanol with the active site 246 of NF-kB was formed by two hydrogen bonds and one Pi-Alkyl interaction with amino acid 247 residues of PRO223, LYS252, and LYS252, respectively. The interaction of benzaldehyde with 248 the active site of NF-KB was formed by a hydrogen bond with the amino acid residue of 249 LYS252. The interaction of fumaric acid with the active site of NF-KB was formed by six 250 hydrogen bonds with the amino acid residues of LYS221, LYS252, and TYR285. The interaction 251 of glucose with the active site of NF-kB was formed by six hydrogen bonds with the amino acid residues SER220, LYS221, SER226, and LYS252. Although glucose and fumaric acid were 252 253 able to be formed by six hydrogen bonds, different types of amino acid residues were involved

- 254 255 in the interaction, so fumaric acid interacted more strongly with the active site of NF- κ B (Fig.
- 4).
- 256



Insert here the figures corresponding to curcumin. Fig. 4. 2D and 3D representation of the kind of interaction on the formation of the NF-κB active site (PDB ID 1A3Q). A) 2-Phenylethanol, B) Benzaldehyde, C) Fumaric acid, D) Glucose, and E) Curcumin.







Fig. 5. 2D and 3D illustration of the type of interaction on the formation of Keap1's binding pocket (PDB ID 4L7B). A) 2-Phenylacetaldehyde, B) Succinic acid, C) Glucose, and D) Curcumin.

259

The interaction of 2-phenylacetaldehyde with the active site of Keap1 was formed by one 260 hydrogen bond and one Pi-Alkyl interaction with amino acid residues of GLN530 and 261 262 ALA556, respectively. The interaction of succinic acid with the active site of Keap1 was formed 263 by four hydrogen bonds with the amino acid residues of ARG336, HIS575, and PHE577. The 264 interaction of glucose with the active site of Keap1 was formed by six hydrogen bonds with 265 the amino acid residues LYS551, ARG553, and ASP573. Although glucose can form more hydrogen bonds than 2-phenylacetaldehyde, the different types of amino acid residues 266 267 involved have not been able to have a significant effect on the binding energy of its interaction 268 with the active site of Keap1 (Fig. 5).

269 Curcumin (the reference compound) created three hydrogen bonds with the amino acid 270 residues ARG52, GLU58, and LYS252 to interact with the active site of NF-KB. Meanwhile, 271 curcumin established three hydrogen bonds with the amino acid residues ARG336, TYR572, 272 and GLY600 to interact with the active site of nrf2 Keap1 (Figs. 4 and 5). Despite the fact that 273 curcumin could create three hydrogen bonds at both the active sites of NF-KB and nrf2 Keap1, 274 its interaction with nrf2 Keap1 had a lower free energy of binding than phenylethanol, 275 benzaldehyde, and fumaric acid (Table 1). Overall, the metabolites of water kefir prefer to 276 interact with NF-KB and nrf2 Keap1 receptors. Whereas fumaric acid and 2-277 phenylacetaldehyde were metabolites that had the strongest interaction with NF-KB (PDB ID 278 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) receptors, respectively.

279

280 DISCUSSION

281 Increased liver enzyme production is one of the abnormalities indicating liver damage. This 282 increase in secretion increases the leakage of enzymes into the circulation. Transaminase is a 283 frequent enzyme used to evaluate liver disorders. The transaminases AST and ALT catalyze 284 the transfer of the α -amino group from alanine and aspartic acid to α -ketoglutaric acid. AST 285 is found in the liver, heart, and skeletal muscle, as well as the kidney, brain, pancreas, lung, leukocytes, and erythrocytes. While ALT is predominantly found in the liver, it is also found 286 287 in low concentrations in other tissues (Lee et al. 2012). Consequently, ALT was used as the 288 principal hepatotoxicity criterion in this study.

289 Carbon tetrachloride (CCl₄) is a component of the hepatotoxin, which acts after metabolic 290 activation and is extensively employed as a liver-damaging agent. In this study, the 291 administration of CCl₄ to animals caused hepatotoxicity, which was characterized by a 292 significant increase in ALT after several administrations. CCl₄ is metabolized by the enzyme 293 cytochrome p450 (CYP2E1) in the endoplasmic reticulum to a highly reactive trichloromethyl 294 radical. The trichloromethyl radical rapidly reacts with oxygen to form the trichloromethyl 295 peroxyl radical, which rapidly reacts with lipids to form lipid peroxidation products. Free 296 radicals will disrupt the endoplasmic reticulum (ER) and lead to lipid accumulation, decreased 297 protein synthesis, and increased oxidase activity. CCl₄ hepatotoxicity is characterized by 298 hepatocellular necrosis with fat deposition (Ritesh et al. 2015). At the molecular level, 299 administration of CCl₄ can activate tumor necrosis factor (TNF)-a, nitric oxide (NO), and 300 transforming growth factor (TGF)- α and - β in cells, processes that precipitate cell self-301 destruction or fibrosis. TNF-a leads to apoptosis, whereas TGF-B leads to fibrosis (Weber et 302 al., 2003).

In terms of its pathophysiological underpinnings, liver illness is linked to a condition known 303 304 as dysbiosis, which refers to an imbalance in the makeup of the gut microbiota (Zavala et al., 305 2016; Laureys and De Vuyst, 2017; Romero-Luna et al., 2020). Both qualitative and quantitative 306 changes in the gut microbiome have the potential to affect the composition of products 307 produced by the microbiota, such as short-chain fatty acids and bile acids (Romero-Luna et 308 al., 2020). Qualitative changes include an imbalance between harmful and helpful 309 microbiomes, whereas numeric changes involve changes to the overall microbiota. In addition 310 to these symptoms, intestinal inflammation, disruption of the intestinal barrier, and the 311 transfer of microbial products can all be caused by dysbiosis (Laureys et al., 2018). For this 312 reason, the condition of the gut microbiome is an important factor in the initiation and 313 development of chronic liver disease (Lee et al., 2021). Based on the results of the study, 314 treatment with water kefir for 2 weeks after the occurrence of liver damage was able to 315 improve the overall condition of the liver, which was marked by a significant decrease in the 316 values of AST, ALT, TNF-α, TGF-β, and significant improvement in liver histology.

317 Water kefir contains a number of microorganisms that have been linked to health benefits, 318 such as the probiotics L. paracasei and B. cereus (Fijan 2014). This activity is linked to an increase 319 in antioxidants like glutathione and catalase and a decrease in pro-inflammatory transcription 320 factors like nuclear B-factor, lipopolysaccharide, and Toll-like receptor 4 (TLR-4). 321 Improvements in intestinal barrier function and histological integrity were also observed. 322 Increased expression of claudin-1 and occludin-1, two components of tight junctions, occurs 323 simultaneously with the restoration of the p38 MAPK pathway (Fijan 2014; Yao et al. 2019; 324 Tsai et al. 2020; Ji et al. 2022). Bacillus is a kind of endospore-forming bacterium that can 325 endure extremely cold temperatures and lengthy periods of storage without dying; its spores can even tolerate the acidic environment of the stomach and make it all the way to the small 326 intestine (Elshaghabee et al. 2017). B. cereus has been shown to reduce ALT levels, an indicator 327 328 of liver healing, in various animal models of liver injury. It protects the liver by reducing 329 inflammation, enhancing the gut flora, and strengthening the tight junctions in the intestines 330 (Kim et al. 2018; Li et al. 2019a; Xue et al. 2020). Also, when Bacillus spores were used first, 331 hepatocyte necrosis and serum levels of ALT, ZO-1, AST, and TAC went down by a lot. This 332 effect is comparable to that of the popular hepatoprotective compound silvmarin (Neag et al. 333 2020).

334 Beer, vinegar, cider, and cocoa bean masses are just some of the places where Gram-335 negative acetic acid bacteria have been found. One of the features that sets acetic acid bacteria 336 apart from others is their alkaline-stable lipid membrane (Lynch et al. 2021). Their "oxidative" 337 fermentation metabolism is responsible for the principal metabolic process in these bacteria, 338 the oxidation of ethanol to acetic acid. Important in the food and beverage sector and beyond, 339 fermentation helps mediate the transition of diverse substrates into products. Although lactic 340 acid bacteria have been studied more extensively than acetic acid bacteria (Semjonovs et al. 341 2014; Hong et al. 2021), various studies have shown promising results concerning the 342 pharmacological effects of acetic acid bacteria, especially as a hepatoprotective agent. Acetic 343 acid, which is the main metabolite of acetic acid bacteria, can lower inflammation and the 344 severity of liver injury in rats with septic shock by increasing the expression of TRIM40. 345 TRIM40 has been shown to minimize liver damage, decrease the synthesis and release of 346 cytokines such as IL-6 and TNF-a, raise the expression of IL-10, improve survival in septic 347 mice, and block the activity of the TLR4 signaling pathway. Acetic acid injection decreased 348 inflammation as well as the production of inflammatory cytokines (Yang et al. 2019). Acetic 349 acid also stimulates the AMPK signaling pathway, which leads to enhanced lipid oxidation 350 and reduced hepatic lipid and body fat deposition (Kondo et al. 2009; Li et al. 2018).

351 Apart from microorganisms that directly provide hepatoprotective effects, the metabolites 352 produced from these microorganisms also have the potential to be hepatoprotective. 353 Molecular docking is a technique that is utilized in the context of NF-kB and Nrf2 to make 354 predictions regarding the binding affinity and orientation of small-molecule inhibitors to their 355 active sites. The transcription factor known as NF-KB is an essential component in the 356 management of both the immune system and the inflammatory response (Dai et al., 2021b). 357 The expression of important inflammatory genes can be inhibited by small-molecule inhibitors 358 that impair the interaction between NF- κ B and DNA. These inhibitors have the potential to be 359 used in therapeutic applications. The Nrf2 binding site is Kelch-like ECH-associated protein 1 360 in the context of nuclear factor erythroid 2-related factor 2, also known as Nrf2 (Keap1) (Zhao 361 et al., 2017^a; Jiang et al., 2019). Small-molecule inhibitors that disrupt the link between Keap1 362 and Nrf2 can activate the Nrf2-ARE signaling pathway, which has been demonstrated to have 363 cytoprotective effects. Keap1 is a negative regulator of Nrf2, and this association can be 364 disrupted by small-molecule inhibitors (Zhao et al., 2017b).

365 Binding energy and Ki are important parameters used in molecular docking to evaluate the 366 strength of the interaction between a ligand and a receptor protein. Binding energy is the 367 energy released when a ligand binds to a receptor protein, and it is calculated as the difference 368 between the energy of the bound complex and the energy of the unbound ligand and protein 369 (Meng et al., 2011). Ki, on the other hand, is the dissociation constant of the ligand-receptor 370 complex. It shows how much ligand is needed to bind 50% of the receptor binding sites. Both 371 binding energy and Ki are used to predict the binding affinity and selectivity of a ligand to a 372 receptor protein (Du et al., 2016). Ki, on the other hand, is the dissociation constant of the 373 ligand-receptor complex. It shows how much ligand is needed to bind 50% of the receptor 374 binding sites (Du et al., 2016).

375 Based on the results of an in silico study of water kefir metabolite compounds, it was known 376 that fumaric acid and 2-phenylacetaldehyde have the strongest interactions with NF-KB (PDB 377 ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) receptors, respectively. Fumaric acid has been 378 studied for its potential as a hepatoprotective agent. Fumaric acid protects rat livers against 379 cadmium toxicity by decreasing oxidative stress and improving hepatic serum. Fumaric acid 380 prevented cadmium-accrued hepatocellular damage and showed the potential to avert hepatic 381 injury against cadmium in rats (Kaur et al., 2020). Fumaric acid esters were found to ameliorate 382 inflammation and oxidative stress in rats, which was associated with less adiposity and ectopic 383 fat accumulation (Šilhavý et al. 2014).

384 NF-κB and Nrf2 are two transcription factors that play important roles in regulating 385 inflammation and cell survival. While NF-kB is involved in the inflammatory response, and 386 Nrf2 is involved in the antioxidant response (Ganesh Yerra et al. 2013; Wang et al. 2022a). Both 387 transcription factors have been investigated as potential targets for the development of 388 hepatoprotective agents (Rahman et al. 2021; Gao et al. 2022; Wang et al. 2022a; Li et al. 2023). 389 There is evidence of crosstalk between the Nrf2 and NF-kB pathways (Ganesh Yerra et al. 2013; 390 Gao et al. 2022). The Nrf2 pathway inhibits the activation of the NF-κB pathway by increasing 391 antioxidant defenses and HO-1 expression, which efficiently neutralizes ROS and detoxifies 392 (Ganesh Yerra et al. 2013). The crosstalk between Nrf2 and NF-κB could be a new therapeutic 393 target against hepatotoxicity (Gao et al. 2022). Researchers have tried to identify molecule 394 activators of Nrf2 as chemoprevention for ROS-dependent carcinogenesis, while others have 395 focused on identifying Nrf2 inhibitors to increase the sensitivity of cancer cells to 396 chemotherapy (Sharifi-Rad et al., 2023). While NF-KB and Nrf2 are involved in different 397 cellular processes, they have both been investigated as potential targets for the development 398 of hepatoprotective agents. Molecular docking studies have been used to investigate the interaction of potential hepatoprotective agents with these transcription factors. There is also
 evidence of crosstalk between the Nrf2 and NF-κB pathways, which could be a new
 therapeutic target against hepatotoxicity.

402 In the pathogenesis of hepatocellular damage, liver fibrosis, and HCC, NF-κB plays a 403 crucial role in regulating inflammation and cell death (Luedde and Schwabe 2011). In response 404 to many stimuli that may pose a threat to the host, NF-KB is activated, setting in motion 405 processes such as inflammation, immunity, wound healing, and pathogen clearance (Luedde 406 and Schwabe 2011). Pathogen-derived chemicals that activate Toll-Like Receptors (TLRs) 407 include lipopolysaccharide (LPS), viral and bacterial DNA and RNA, and inflammatory 408 cytokines, including tumor necrosis factor (TNF) and interleukins (IL)-1 (Luedde and Schwabe 409 2011). When NF-B is activated, a lot of genes with B-binding sites are transcribed. These genes 410 play important roles in controlling inflammation, the immune response, and cell survival. In 411 an NF-kB-dependent manner, lipopolysaccharide (LPS) activates TLR4 on dormant HSCs by 412 reducing BAMBI expression (an inhibitory TGF-β pseudoreceptor) and increasing Kupffer cell 413 chemotaxis. Due to low levels of BAMBI, recruited Kupffer cells secrete TGF-B, which 414 stimulates HSCs unrestrictively. When HSCs have been activated, NF-KB serves a second 415 crucial function by increasing their chances of survival. LPS, Kupffer cell-derived mediators 416 (such as IL-1 and TNF), and angiotensin II, which is produced and acts on HSCs in an autocrine 417 way, all play roles in activating NF-KB in activated hepatic stellate cells. More activated HSCs 418 and extracellular matrix are deposited in the liver as a result of greater HSC activation and 419 survival (Luedde and Schwabe 2011).

420 TNF- α and TGF- β can stimulate one another's production, and TNF- α has been shown to 421 influence TGF-β expression in a variety of cells and tissues (Liu et al. 2022). TNF-α is an 422 inflammatory cytokine that contributes to liver inflammation, and chronic liver inflammation 423 results in liver fibrosis. TNF-a exerts its effects on liver fibrosis via multiple mechanisms, 424 including TGF-β signaling activation (Yang and Seki 2015). Targeting TNF-α and TGF-β 425 signaling pathways may, therefore, have therapeutic potential for treating liver diseases. In 426 regards to hepatoprotective effects, the relationship between TGF- β and TNF- α is complex and 427 not completely understood.

428 CONCLUSION

429 This study evaluated the hepatoprotective qualities of Indonesian water kefir in rats with 430 CCl4-induced liver damage. Water kefir administration improved the condition of liver 431 damage, characterized by a decrease in serum levels of AST, ALT, TNF, TGF, and an 432 improvement of liver tissue profile. *In silico* evaluation showed that the metabolites in water 433 kefir were able to interact with target proteins in the NF-κB and Nrf2 pathways. It was 434 concluded that water kefir improves the condition of the liver by reducing the level of necrosis 435 and fibrosis.

436

437 CONFLICT OF INTEREST

438 The authors declare no conflict of interest.

439 ACKNOWLEDGMENT

440This research was funded by the Center of Research and Community Service, Bhakti Kencana University,441Bandung, West Java, Republic of Indonesia (052/14.LPPM/PE.I/LPPM/2021)

442	REFERENCES
443	This section is one of the most important sections of the document.
444	Please pay close attention to it.
445	The style of the References does not meet the requirements of JPPRes.
446	Please write the References according to the Instructions to the Author or the examples:
447	https://jppres.com/jppres/archive/
448	https://jppres.com/jppres/volume-11-issue-2/
449	See examples in 1-3 references.
450	Also, all authors (not "et al. ") and DOIs must be given.
451	Please check that all references in this section have been cited in the text and vice versa.
452	
453	A Asnawi, Aman I.O. Nursamsiar, et al (2022) Molecular docking and molecular dynamic
454	studies: Screening phytochemicals of <i>Acalupha indica</i> against braf kinase receptors for
455	potential use in melanocytic tumours. RIC Rasayan J Chem 15: 1352–1361.
456	https://doi.org/10.31788/RJC.2022.1526769
157	Aligita W/ Alox V/ Taaraungan C Sucilawati E (2021) Hanatanratastiwa astiwity of materia
4J1 158	Aligna vv, Alex v, Taarauligan 5, Sushawati E (2021) riepatoprotective activity of water Keffr. $\frac{IIRDAC}{IIRDAC}$ Int I Biol Pharm Allied Sci 10, 1784 1704
+30 459	$\frac{1}{100}$ III J DIOI FILITII AIIEU SCI IO. 1704-1794. https://doi.org/10.31032/IIBPAS/2021/10.6.5493
т <i>Ј)</i>	mps.//doi.org/10.51052/1Jbi h5/2021/10.0.5495
160	Aligita W, Tarigan PN, Susilawati E (2020) Anti Inflammatory <mark>a</mark> nd antioxidant activity of <mark>k</mark> efir
61	water. <mark>IJBPAS</mark> Int J Biol Pharm Allied Sci 9: 2454-2464.
52	https://doi.org/10.31032/IJBPAS/2020/9.1.4904
53	Al-Mohammadi A-R Ibrahim RA Moustafa AH et al (2021) Chemical Constitution and
4	Antimicrobial Activity of Kefir Fermented Beverage Molecules 26:2635
55	https://doi.org/10.3390/molecules26092635
56	Alsayadi M, Aljawfi Y, Belarbi M, Sabri FZ (2013) ANTIOXIDANT POTENCY OF WATER
67	KEFIR. Journal of Microbiology, Biotechnology and Food Sciences 2:2444–2447
58	Alsayadi M, Jawfi YA, Belarbi M, et al (2014) Evaluation of Anti-Hyperglycemic and Anti-
9	Hyperlipidemic Activities of Water Kefir as Probiotic on Streptozotocin-Induced
0	Diabetic Wistar Rats. JDM 04:85-95. https://doi.org/10.4236/jdm.2014.42015
7 1	
/1	Asnawı A, Febrina E, Aligita W, et al (2022) Screening Of Ashitaba (Angelica Keiskei K.)
2	Compounds As Potential Mycobacterium Tuberculosis Kasa Inhibitors. Int J App
3	Pharm 80–85. https://doi.org/10.22159/ijap.2022.v14s5.13
4	Asnawi A, Nedja M, Febrina E, Purwaniati (2023) Prediction of a Stable Complex of
5	Compounds in the Ethanol Extract of Celery Leaves (Apium graveolens L.) Function
6	as a VKORC1 Antagonist. TJNPR 7:2362-2370.
7	https://doi.org/10.26538/tjnpr/v7i2.10
o	A prime REF Elemen REAC Densie MC (2015) Line (Lemeter effect of Ferry entry 114, $t \in K$
0 0	Aspiras DEE, FIORES KFAC, Fareja NIC (2015) Hepatoprotective effect of Fermented Water Kefir
7]	A cetaminophen INT I CURP SCI 17.18_28
50	Accumulophen, hvi j Connoci 17.10-20

- 481 Calatayud M, Börner RA, Ghyselinck J, et al (2021) Water Kefir and Derived Pasteurized
 482 Beverages Modulate Gut Microbiota, Intestinal Permeability and Cytokine Production
 483 In Vitro. Nutrients 13:3897. https://doi.org/10.3390/nu13113897
- 484 Dai C, Li H, Wang Y, et al (2021a) Inhibition of Oxidative Stress and ALOX12 and NF-κB
 485 Pathways Contribute to the Protective Effect of Baicalein on Carbon Tetrachloride486 Induced Acute Liver Injury. Antioxidants 10:976.
 487 https://doi.org/10.3390/antiox10060976
- 488 Dai C, Li H, Wang Y, et al (2021b) Inhibition of Oxidative Stress and ALOX12 and NF-κB
 489 Pathways Contribute to the Protective Effect of Baicalein on Carbon Tetrachloride490 Induced Acute Liver Injury. Antioxidants (Basel) 10:976.
 491 https://doi.org/10.3390/antiox10060976
- 492 Dallakyan S, Olson AJ (2015) Small-Molecule Library Screening by Docking with PyRx. In:
 493 Hempel JE, Williams CH, Hong CC (eds) Chemical Biology. Springer New York, New
 494 York, NY, pp 243-250
- 495 Darvishzadeh P, Orsat V, Martinez JL (2021) Process Optimization for Development of a Novel
 496 Water Kefir Drink with High Antioxidant Activity and Potential Probiotic Properties
 497 from Russian Olive Fruit (Elaeagnus angustifolia). Food Bioprocess Technol 14:248498 260. https://doi.org/10.1007/s11947-020-02563-1
- 499 De Souza Basso B, Haute GV, Ortega-Ribera M, et al (2021) Methoxyeugenol deactivates
 500 hepatic stellate cells and attenuates liver fibrosis and inflammation through a PPAR-γ
 501 and NF-kB mechanism. Journal of Ethnopharmacology 280:114433.
 502 https://doi.org/10.1016/j.jep.2021.114433
- 503Diniz RO, Garla LK, Schneedorf JM, Carvalho JCT (2003) Study of anti-inflammatory activity504of Tibetan mushroom, a symbiotic culture of bacteria and fungi encapsulated into a505polysaccharide matrix. Pharmacological Research 47:49–52.506https://doi.org/10.1016/S1043-6618(02)00240-2
- 507Du X, Li Y, Xia Y-L, et al (2016) Insights into Protein-Ligand Interactions: Mechanisms,508Models, and Methods. IJMS 17:144. https://doi.org/10.3390/ijms17020144
- 509 Elshaghabee FMF, Rokana N, Gulhane RD, et al (2017) Bacillus As Potential Probiotics: Status,
 510 Concerns, and Future Perspectives. Front Microbiol 8:1490.
 511 https://doi.org/10.3389/fmicb.2017.01490
- 512 Febrina E, Alamhari RK, Abdulah R, et al (2021) MOLECULAR DOCKING AND
 513 MOLECULAR DYNAMICS STUDIES OF ACALYPHA INDICA L.
 514 PHYTOCHEMICAL CONSTITUENTS WITH CASPASE-3. Int J App Pharm 210–215.
 515 https://doi.org/10.22159/ijap.2021.v13s4.43861
- Febrina E, Asnawi A, Abdulah R, et al (2022) Identification Of Flavonoids From Acalypha
 Indica L. (Euphorbiaceae) As Caspase-3 Activators Using Molecular Docking And
 Molecular Dynamics. Int J App Pharm 162–166.
 https://doi.org/10.22159/ijap.2022.v14s5.34
- Fijan S (2014) Microorganisms with Claimed Probiotic Properties: An Overview of Recent
 Literature. IJERPH 11:4745–4767. https://doi.org/10.3390/ijerph110504745

- 522 Gamba RR, Yamamoto S, Sasaki T, et al (2019) Microbiological and Functional
 523 Characterization of Kefir Grown in Different Sugar Solutions. FSTR 25:303–312.
 524 https://doi.org/10.3136/fstr.25.303
- 525 Ganesh Yerra V, Negi G, Sharma SS, Kumar A (2013) Potential therapeutic effects of the
 526 simultaneous targeting of the Nrf2 and NF-κB pathways in diabetic neuropathy. Redox
 527 Biology 1:394–397. https://doi.org/10.1016/j.redox.2013.07.005
- Gao W, Guo L, Yang Y, et al (2022) Dissecting the Crosstalk Between Nrf2 and NF-κB Response
 Pathways in Drug-Induced Toxicity. Front Cell Dev Biol 9:809952.
 https://doi.org/10.3389/fcell.2021.809952
- Hong Y, Sheng L, Zhong J, et al (2021) Desulfovibrio vulgaris, a potent acetic acid-producing
 bacterium, attenuates nonalcoholic fatty liver disease in mice. Gut Microbes
 13:1930874. https://doi.org/10.1080/19490976.2021.1930874
- Ischak NI, Aman LO, Hasan H, et al (2023) In silico screening of Andrographis paniculata
 secondary metabolites as anti-diabetes mellitus through PDE9 inhibition. Res Pharm
 Sci 18:100–111. https://doi.org/10.4103/1735-5362.363616
- Ji Y, Xie Q, Meng X, et al (2022) Lactobacillus paracasei improves dietary fatty liver by reducing
 insulin resistance and inflammation in obese mice model. Journal of Functional Foods
 95:105150. https://doi.org/10.1016/j.jff.2022.105150
- Jiang Z-Y, Lu M-C, You Q-D (2019) Nuclear Factor Erythroid 2-Related Factor 2 (Nrf2)
 Inhibition: An Emerging Strategy in Cancer Therapy. J Med Chem 62:3840–3856.
 https://doi.org/10.1021/acs.jmedchem.8b01121
- Kaur G, Shivanandappa TB, Kumar M, Kushwah AS (2020) Fumaric acid protect the cadmiuminduced hepatotoxicity in rats: owing to its antioxidant, anti-inflammatory action and
 aid in recast the liver function. Naunyn-Schmiedeberg's Arch Pharmacol 393:19111920. https://doi.org/10.1007/s00210-020-01900-7
- 547 Kebler LF (1921) "California Bees." 11The author stated that the work on this subject is the
 548 result of investigation for the Post a c e Department relative to improper use of the
 549 mails in connection with this product. The Journal of the American Pharmaceutical
 550 Association (1912) 10:939–943. https://doi.org/10.1002/jps.3080101206
- Kim B, Kwon J, Kim M-S, et al (2018) Protective effects of Bacillus probiotics against high-fat
 diet-induced metabolic disorders in mice. PLoS ONE 13:e0210120.
 https://doi.org/10.1371/journal.pone.0210120
- Kondo T, Kishi M, Fushimi T, Kaga T (2009) Acetic Acid Upregulates the Expression of Genes
 for Fatty Acid Oxidation Enzymes in Liver To Suppress Body Fat Accumulation. J
 Agric Food Chem 57:5982–5986. https://doi.org/10.1021/jf900470c
- Konstantopoulos P, Doulamis I, Tzani A, et al (2017) Metabolic effects of Crocus sativus and
 protective action against non-alcoholic fatty liver disease in diabetic rats. biom rep.
 https://doi.org/10.3892/br.2017.884

- Laureys D, Aerts M, Vandamme P, De Vuyst L (2018) Oxygen and diverse nutrients influence
 the water kefir fermentation process. Food Microbiology 73:351–361.
 https://doi.org/10.1016/j.fm.2018.02.007
- Laureys D, De Vuyst L (2017) The water kefir grain inoculum determines the characteristics of
 the resulting water kefir fermentation process. J Appl Microbiol 122:719–732.
 https://doi.org/10.1111/jam.13370
- Laureys D, De Vuyst L (2014) Water kefir as a promising low-sugar probiotic fermented
 beverage. Arch Public Health 72:P1, 2049-3258-72-S1-P1.
 https://doi.org/10.1186/2049-3258-72-S1-P1
- Lee NY, Shin MJ, Youn GS, et al (2021) *Lactobacillus* attenuates progression of nonalcoholic
 fatty liver disease by lowering cholesterol and steatosis. Clin Mol Hepatol 27:110–124.
 https://doi.org/10.3350/cmh.2020.0125
- Lee TH, Kim WR, Poterucha JJ (2012) Evaluation of Elevated Liver Enzymes. Clinics in Liver
 Disease 16:183–198. https://doi.org/10.1016/j.cld.2012.03.006
- 574 Li H, Weng Q, Gong S, et al (2023) Kaempferol prevents acetaminophen-induced liver injury
 575 by suppressing hepatocyte ferroptosis *via* Nrf2 pathway activation. Food Funct
 576 14:1884–1896. https://doi.org/10.1039/D2FO02716J
- 577 Li L, He M, Xiao H, et al (2018) Acetic Acid Influences BRL-3A Cell Lipid Metabolism via the
 578 AMPK Signalling Pathway. Cell Physiol Biochem 45:2021–2030.
 579 https://doi.org/10.1159/000487980
- Li Y-T, Ye J-Z, Lv L-X, et al (2019a) Pretreatment With Bacillus cereus Preserves Against DGalactosamine-Induced Liver Injury in a Rat Model. Front Microbiol 10:1751.
 https://doi.org/10.3389/fmicb.2019.01751
- Li Z, Huang Y, Wu Y, et al (2019b) Absolute Binding Free Energy Calculation and Design of a
 Subnanomolar Inhibitor of Phosphodiesterase-10. J Med Chem 62:2099–2111.
 https://doi.org/10.1021/acs.jmedchem.8b01763
- 586Liu Z, Zhang Y, Zhang L, et al (2022) Duality of Interactions Between TGF-β and TNF-α During587TumorFormation.FrontImmunol12:810286.588https://doi.org/10.3389/fimmu.2021.810286
- Luedde T, Schwabe RF (2011) NF-κB in the liver—linking injury, fibrosis and hepatocellular
 carcinoma. Nat Rev Gastroenterol Hepatol 8:108–118.
 https://doi.org/10.1038/nrgastro.2010.213
- Lynch KM, Wilkinson S, Daenen L, Arendt EK (2021) An update on water kefir: Microbiology,
 composition and production. International Journal of Food Microbiology 345:109128.
 https://doi.org/10.1016/j.ijfoodmicro.2021.109128
- 595Meng X-Y, Zhang H-X, Mezei M, Cui M (2011) Molecular Docking: A Powerful Approach for596Structure-BasedDrugDiscovery.CAD7:146-157.597https://doi.org/10.2174/157340911795677602

- Moinas M, Horisberger M, Bauer H (1980) The structural organization of the Tibi grain as
 revealed by light, scanning and transmission microscopy. Arch Microbiol 128:157–161.
 https://doi.org/10.1007/BF00406153
- Moreira MEC, Santos MHD, Zolini GPP, et al (2008) Anti-Inflammatory and Cicatrizing
 Activities of a Carbohydrate Fraction Isolated from Sugary Kefir. Journal of Medicinal
 Food 11:356–361. https://doi.org/10.1089/jmf.2007.329
- Neag MA, Catinean A, Muntean DM, et al (2020) Probiotic Bacillus Spores Protect Against
 Acetaminophen Induced Acute Liver Injury in Rats. Nutrients 12:632.
 https://doi.org/10.3390/nu12030632
- Nursamsiar, Nur S, Febrina E, et al (2022) Synthesis and Inhibitory Activity of Curculigoside
 A Derivatives as Potential Anti-Diabetic Agents with β-Cell Apoptosis. Journal of
 Molecular Structure 1265:133292. https://doi.org/10.1016/j.molstruc.2022.133292
- Parasuraman S, Raveendran R, Kesavan (2010) Blood sample collection in small laboratory
 animals. Journal of Pharmacology and Pharmacotherapeutics 1:87–93.
 https://doi.org/10.4103/0976-500X.72350
- Patel SH, Tan JP, Börner RA, et al (2022) A temporal view of the water kefir microbiota and
 flavour attributes. Innovative Food Science & Emerging Technologies 80:103084.
 https://doi.org/10.1016/j.ifset.2022.103084
- 616 Pidoux M (1989) Kefir Grain Flora. Mircen Journal 5:223–228
- Rahman ZU, Al Kury LT, Alattar A, et al (2021) Carveol a Naturally-Derived Potent and
 Emerging Nrf2 Activator Protects Against Acetaminophen-Induced Hepatotoxicity.
 Front Pharmacol 11:621538. https://doi.org/10.3389/fphar.2020.621538
- Ritesh KR, Suganya A, Dileepkumar HV, et al (2015) A single acute hepatotoxic dose of CCl 4
 causes oxidative stress in the rat brain. Toxicology Reports 2:891–895.
 https://doi.org/10.1016/j.toxrep.2015.05.012
- Rocha-Gomes A, Escobar A, Soares JS, et al (2018) Chemical composition and
 hypocholesterolemic effect of milk kefir and water kefir in Wistar rats. Rev Nutr
 31:137–145. https://doi.org/10.1590/1678-98652018000200001
- Rodrigues KL, Araújo TH, Schneedorf JM, et al (2016) A novel beer fermented by kefir
 enhances anti-inflammatory and anti-ulcerogenic activities found isolated in its
 constituents. Journal of Functional Foods 21:58–69.
 https://doi.org/10.1016/j.jff.2015.11.035
- Romero-Luna HE, Peredo-Lovillo A, Hernández-Mendoza A, et al (2020) Probiotic Potential
 of Lactobacillus paracasei CT12 Isolated from Water Kefir Grains (Tibicos). Curr
 Microbiol 77:2584–2592. https://doi.org/10.1007/s00284-020-02016-0
- 633 Semjonovs P, Denina I, Linde R (2014) Evaluation of Physiological Effects of Acetic Acid
 634 Bacteria and Yeast Fermented Non-alchocolic Beverage Consumption in Rat Model. J
 635 of Medical Sciences 14:147–152. https://doi.org/10.3923/jms.2014.147.152

- 636 Sharifi-Rad J, Seidel V, Izabela M, et al (2023) Phenolic compounds as Nrf2 inhibitors: potential
 637 applications in cancer therapy. Cell Commun Signal 21:89.
 638 https://doi.org/10.1186/s12964-023-01109-0
- Šilhavý J, Zídek V, Mlejnek P, et al (2014) Fumaric Acid Esters Can Block Pro-Inflammatory
 Actions of Human CRP and Ameliorate Metabolic Disturbances in Transgenic
 Spontaneously Hypertensive Rats. PLoS ONE 9:e101906.
 https://doi.org/10.1371/journal.pone.0101906
- 643 Tsai Y-S, Lin S-W, Chen Y-L, Chen C-C (2020) Effect of probiotics Lactobacillus paracasei GKS6, 644 L. plantarum GKM3, and L. rhamnosus GKLC1 on alleviating alcohol-induced alcoholic 645 liver disease in а mouse model. Nutr Res Pract 14:299. 646 https://doi.org/10.4162/nrp.2020.14.4.299
- Wang B, Cui S, Mao B, et al (2022a) Cyanidin Alleviated CCl4-Induced Acute Liver Injury by
 Regulating the Nrf2 and NF-κB Signaling Pathways. Antioxidants 11:2383.
 https://doi.org/10.3390/antiox11122383
- Wang Y, Liu F, Liu M, et al (2022b) Curcumin mitigates aflatoxin B1-induced liver injury via
 regulating the NLRP3 inflammasome and Nrf2 signaling pathway. Food and Chemical
 Toxicology 161:112823. https://doi.org/10.1016/j.fct.2022.112823
- Ward M (1892) V. The ginger-beer plant, and the organisms composing it: a contribution to
 the study of fermentation-yeasts and bacteria. Phil Trans R Soc Lond B 183:125–197.
 https://doi.org/10.1098/rstb.1892.0006
- Weber LWD, Boll M, Stampfl A (2003) Hepatotoxicity and Mechanism of Action of
 Haloalkanes: Carbon Tetrachloride as a Toxicological Model. Critical Reviews in
 Toxicology 33:105–136. https://doi.org/10.1080/713611034
- Kue J, Shen K, Hu Y, et al (2020) Effects of dietary Bacillus cereus, B. subtilis, Paracoccus marcusii, and Lactobacillus plantarum supplementation on the growth, immune response, antioxidant capacity, and intestinal health of juvenile grass carp (Ctenopharyngodon idellus). Aquaculture Reports 17:100387.
 https://doi.org/10.1016/j.aqrep.2020.100387
- Yang H, Meng L, Ai D, et al (2019) Acetic acid alleviates the inflammatory response and liver
 injury in septic mice by increasing the expression of TRIM40. Exp Ther Med.
 https://doi.org/10.3892/etm.2019.7274
- 667 Yang YM, Seki E (2015) TNFα in Liver Fibrosis. Curr Pathobiol Rep 3:253–261.
 668 https://doi.org/10.1007/s40139-015-0093-z
- Yao F, Jia R, Huang H, et al (2019) Effect of Lactobacillus paracasei N1115 and
 fructooligosaccharides in nonalcoholic fatty liver disease. aoms 15:1336–1344.
 https://doi.org/10.5114/aoms.2019.86611
- Yeh Y-H, Hsieh Y-L, Lee Y-T, Hsieh C-H (2011) Protective effects of cholestin against carbon
 tetrachloride-induced hepatotoxicity in rats. e-SPEN, the European e-Journal of
 Clinical Nutrition and Metabolism 6:e264–e271.
 https://doi.org/10.1016/j.eclnm.2011.09.002

- 676 Zamberi NR, Abu N, Mohamed NE, et al (2016) The Antimetastatic and Antiangiogenesis
 677 Effects of Kefir Water on Murine Breast Cancer Cells. Integr Cancer Ther 15:NP53–
 678 NP66. https://doi.org/10.1177/1534735416642862
- Zavala L, Golowczyc MA, van Hoorde K, et al (2016) Selected *Lactobacillus* strains isolated
 from sugary and milk kefir reduce *Salmonella* infection of epithelial cells *in vitro*.
 Beneficial Microbes 7:585–595. https://doi.org/10.3920/BM2015.0196
- Kato H, Eguchi S, Alam A, Ma D (2017) The role of nuclear factor-erythroid 2 related factor
 2 (Nrf-2) in the protection against lung injury. Am J Physiol Lung Cell Mol Physiol
 312:L155–L162. https://doi.org/10.1152/ajplung.00449.2016
- 685 Zhao H, Eguchi S, Alam A, Ma D (2017b) The role of nuclear factor-crythroid 2 related factor
 686 2 (Nrf-2) in the protection against lung injury. Am J Physiol Lung Cell Mol Physiol
 687 312:L155–L162. https://doi.org/10.1152/ajplung.00449.2016
- 688
- 689

1	Hepatoprotective study of Indonesian water kefir against				
2	CCl ₄ -induced liver injury in rats				
3					
4	Ru	nning title: Hepatoprotective	e study of water kefir		
5	Widhya	n Aligita ^{1,3*} , Marlia Singgih ² , Entris	Sutrisno ³ , I Ketut Adnyana	1*	
67 89 10 11 12 13 14	¹ Department of Pharmacology & Clinical Pharmacy, School of Pharmacy, Bandung Institute of Technology, Bandung, Indonesia. ² Department of Pharmacochemistry, School of Pharmacy, Bandung Institute of Technology, Bandung, Indonesia. ³ Department of Pharmacology & Clinical Pharmacy, Faculty of Pharmacy, Bhakti Kencana University, Bandung, Indonesia. *E-mail address: widhya.aligita@bku.ac.id / ketut@itb.ac.id *Corresponding author address: Department of Pharmacology & Clinical Pharmacy, School of Pharmacy, Bandung Institute of Technology, Ganeca, 10, 40132, Bandung, Indonesia. Tel: +62-22-2504852				
15 16 17	The institutional e-mail of authors):	the ALL authors must be declared ((and add rows according to t	he number of	
	Author	Institutional e-mail (mandatory)	Other e-mail	ORCID	

autions).			
Author	Institutional e-mail (mandatory)	Other e-mail (gmail, yahoo, etc.)	ORCID (0000-1111-2222-3333)
Widhya Aligita	widhya.aligita@bku.ac.id	w.aligita@gmail.com	0000-0001-8338-4115
Marlia Singgih	marlia@fa.itb.ac.id		0000-0002-5351-1731
Entris Sutrisno	entris.sutrisno@bku.ac.id		0000-0003-3830-6411
I Ketut Adnyana	ketut@itb.ac.id		0000-0001-5217-2312

Contribution Details (to be ticked marked (X) as applicable and add columns according to the number of authors): 20

Contribution	Aligita W	Singgih M	Sutrisno E	Adnyana IK
Concepts or Ideas	x	x	х	х
Design	х	х	х	х
Definition of intellectual content	х	х	х	х
Literature search	х			х
Experimental studies	х			х
Data acquisition	х			Х
Data analysis	х			Х
Statistical analysis	х			Х
Manuscript preparation	х	х	Х	Х
Manuscript editing	х	х	Х	х
Manuscript review	х	х	х	х

23 ABSTRACT

- 24 25 26 Context: Water kefir is a fermented beverage that is typically made in the home by inoculating a sugar-rich solution with a microbial community (water kefir grains). Several studies on the metabolite content and hepatoprotective effects of water kefir have been published, but CCl4-induced acute liver injury has not been studied.
- 27 28 Objectives: To evaluate the efficacy of water kefir in vivo against hepatoprotective CCl4-induced acute liver injury and to in silico investigate metabolites that play an important role in hepatoprotective mechanisms.
- Methods: The present study aimed to investigate the hepatoprotective activity of water kefir in an animal model caused by carbon tetrachloride (CCl4). Furthermore, using molecular docking, the metabolites found in water kefir were evaluated for their role in the NF-KB and Nrf2 signaling pathways.
- 29 31 32 33 4 35 36 37 8 940 Results: Water kefir significantly and dose-dependently alleviated acute liver injury caused by carbon tetrachloride (CCl4). Water kefir administration at all doses produced results comparable to the positive control (Curcuma extract). Molecular docking simulations showed that, compared to Nrf2, the 25 metabolites were more likely to interact with the NF-B receptor. Fumaric acid is the strong metabolite that interacts with the NF-KB receptor with a free energy of binding and an inhibition constant of -6.66 kcal/mol and 13.22 µM, respectively.
- Conclusions: Water kefir administration improved the condition of liver damage, characterized by a decrease in serum levels of AST, ALT, TNF-a, TGF-B, and an improvement in the liver tissue profile. In silico evaluation showed that the metabolites in water kefir were able to interact with target proteins in the NF-κB and Nrf2 pathways. It was concluded that water kefir improves the condition of the liver by reducing the level of necrosis and fibrosis.
- 41
- 42 Keywords: free radicals; liver diseases; kefir; molecular docking simulation; probiotics
- 43

44 **INTRODUCTION**

45 In most cases, making water kefir involves combining dried fruit, sugar, and water kefir grains in a container. Water kefir's exact origins are unknown; however, two hypotheses have 46 47 been proposed regarding its history: the first suggests that water kefir grains were brought to 48 Europe from the Caucasus by soldiers returning from the Crimean War in the late nineteenth 49 century (Ward, 1892); the second theory proposes that water kefir grains originated in Mexico 50 from the Opuntia cactus through natural processes (Moinas et al., 1980). Sugary kefir grains, 51 Balm of Gilead, African bees, Japanese beer seeds, Ale nuts, and California bees are some other 52 names for water kefir. Tibi grains and ginger beer plants are other names for water kefir 53 (Kebler, 1921; Moinas et al., 1980). Water kefir is appealing to both consumers and researchers 54 due to the variety of microbiota it contains, the fact that it is an alternative to dairy products, 55 the versatility with which it can be flavored, the fact that it is low in calories and sugar, the

56 ease with which it can be produced, and the health benefits it offers.

57 Water kefir has been used medicinally for a very long time, and recent research has 58 indicated that it may have a variety of positive effects on people's health. It has been 59 demonstrated that water kefir contains non-pathogenic bacteria that, in conjunction with the 60 production of organic acids, can inhibit the growth of pathogenic microbes such as *Shigella sp.*, 61 Salmonella sp., Staphylococcus aureus, and E. coli; as well as, filamentous fungi such as Aspergillus 62 ochraceus, A. niger, A. flavus, Penicillium sp., and Rhizopus sp. (Al-Mohammadi et al., 2021). In 63 addition to its antibacterial properties, water kefir possesses a broad spectrum of pharmacological effects. Some of these therapeutic effects are anti-inflammatory (Diniz et al., 64 65 2003; Aligita et al., 2020), antioxidant (Aligita et al., 2020; Darvishzadeh et al., 2021), 66 hepatoprotective (Aspiras et al., 2015; Aligita et al., 2021), antihyperglycemic and antihyperlipidemic (Alsayadi et al., 2014; Rocha-Gomes et al., 2018), anti-edematous (Moreira 67 68 et al., 2008), antitumor (Zamberi et al., 2016), antihypertensive (Gamba et al., 2019), immunomodulant (Calatayud et al., 2021), and anti-ulcerogenic (Rodrigues et al., 2016). 69 70 However, no studies have been reported on the hepatoprotective effects of water kefir against 71 carbon tetrachloride (CCl₄)-induced liver injury.

72 Studies have shown that acute liver injury is frequently accompanied by high levels of 73 oxidative stress and inflammatory responses (Dai et al., 2021). These findings have been found 74 in several studies. The most important signaling pathways that are involved in the regulation 75 of inflammation and antioxidation are the nuclear factor (NF-KB) and nuclear factor erythroid 76 2-related factor 2 (Nrf2) pathways, respectively. It has been shown that activating Nrf2 and 77 inhibiting NF-KB can reduce the amount of damage done to the liver. For instance, curcumin protects against aflatoxin B1-induced liver injury by increasing the expression of Nrf2 and 78 79 related downstream antioxidant molecules (such as superoxide dismutase (SOD), catalase 80 (CAT), heme oxygenase 1 (HO-1), and NAD(P)H quinone dehydrogenase 1 (NQO-1) (Wang et al., 2022b). Also, it has been shown that the molecule methoxy eugenol, which comes from 81 82 nutmeg and Brazilian red propolis, protects the liver both in vitro and in vivo. This may be 83 attributed to the fact that it targets the NF-kB signaling pathway, which has been shown to 84 have anti-inflammatory effects (De Souza Basso et al., 2021).

85 Ethanol and lactic acid are the primary metabolites found in fermented water kefir, while 86 lower quantities of glycerol, mannitol, and acetic acid can also be found in the beverage. 87 Additionally, a variety of aromatic and volatile compounds are produced, including ethyl 88 acetate, isoamyl acetate, ethyl octanoate, ethyl hexanoate, and ethyl decanoate, among others 89 (compared to their threshold levels) (Laureys and De Vuyst 2014). The chemical constituents 90 of both phytochemicals and secondary metabolites in natural products, including water kefir, 91 are certainly capable of providing various pharmacological effects for the body (Asnawi et al., 92 2022; Nursamsiar et al., 2022). However, an in silico study to evaluate the metabolite content 93 in water kefir has not been reported yet. Because of its capacity to speed up the process of 94 identifying and optimizing lead compounds, the in silico method has become the front-runner 95 in the race to improve the speed and accuracy of the process of discovering new drugs. This is 96 because the *in silico* method can identify and optimize lead compounds more quickly. 97 Techniques such as molecular docking and molecular dynamics (MD) were able to directly 98 indicate a small number of compounds that have high affinity and selectivity by analyzing 99 how the ligand and target interact with one another (Febrina et al., 2021).

Water kefir has been used for an extensive period of time and has been recognized for its widespread benefits, especially in Indonesia. However, its level of popularity falls short in comparison to that of milk kefir. Furthermore, there is a scarcity of studies specifically focused on the hepatoprotective activity of water kefir made with Indonesian grains. Therefore, the purpose of this study was to evaluate the hepatoprotective effects of water kefir in CCl₄induced rats while also investigating the stability interactions of its metabolites within the NF-KB and Nrf2 receptors using molecular docking studies.

107

108 MATERIAL AND METHODS

109 Materials and reagents

Water kefir grains, sugar, dried fruit, carbon tetrachloride, rats, diagnostic kits for alanine
aminotransferase (ALT) (Proline, IFCC mod.), aspartate aminotransferase (AST) (Proline,
IFCC mod.), Elisa Kit TNF-α (Bioassay Technology Laboratory), Elisa Kit TGF-β (Bioassay
Technology Laboratory). Other chemicals used in this study were of analytical reagent grade.

114 **Experimental sample and reference extract**

The water kefir grain was obtained from Yogyakarta, Indonesia. The water kefir solution was produced using a fermentation procedure. The initial stage involved the preparation of 60 g of sugar, 2 g of raisins, 100 g of water kefir grains, and 1 L of aqua mineral distillate. The sugar and warm distilled water were mixed in a beaker, followed by the addition of water kefir grains and raisins to the resulting sugar solution. The fermentation procedure was conducted over a duration of forty-eight hours at ambient temperature. A small cloth was used 121 to cover the beaker glass. The kefir grain was utilized in future production, while the filtrate 122 was employed for the purpose of evaluation and analysis. (Aligita et al., 2020, 2021)

123 The rhizoma extract of Curcuma (*Curcuma xanthorrhiza* Roxb) is employed as a reference 124 drug. The utilized product is a standardized herbal medicine with the brand name Tulak, 125 manufactured by the Borobudur Company Herbal Medicine Industry. The primary purpose 126 of Tulak capsules is to support and preserve optimal liver functionality.

127 Animals and experimental design

128 Rats (Wistar strain, male, 200-250 g) were maintained on normal pellet food and tap water 129 ad libitum. Four mice in each group were used. All procedures relating to animals and their 130 care conformed to the international guidelines Principles of Laboratory Animal Care (NIH publication 131 no. 85-23, revised 1985) with the ethical approval number 132 363/UN6.KEP/EC/2020. In order to develop an animal model with liver injury, the rats 133 received CCl₄ (20% in olive oil) at 1.25 mL/kg BW every 2 days via a gavage tube (Yeh et al. 134 2011). The rats were randomized into five groups after the development of animals with liver 135 injury, which is characterized by a significant increase in serum ALT level, as follows: (1) 136 positive control group, (2) Curcuma extract group 100 mg/kg BW, (3) water kefir 15 mL/kg 137 BW group, (4) water kefir 35 mL/kg BW group, and (5) water kefir 50 mL/kg BW group, with 138 the addition (6) negative control group. Each group received group-specific treatment for two 139 weeks, along with the administration of 1.25 mL/kg BW of CCl₄ (20% in olive oil) every three 140 days.

141 The rats, which had undergone a fasting period of 8-10 hours while being provided with 142 water, had their blood samples collected via the retro-orbital sinus using a hematocrit capillary 143 tube. The blood sample was taken into a microtube and afterwards then to centrifugation. The 144 serum was separated in order to facilitate further measurements (Parasuraman et al., 2010). 145 Serum ALT level, as the main parameter, was measured prior to induction, following induction, and following treatment. Meanwhile, following therapy, serum AST, TNF-a, TGF-146 147 β levels, and liver histopathological were evaluated. The ALT, AST, TNF- α , TGF- β levels 148 measurements are conducted in accordance with the protocols outlined in the reagent kit.

After the euthanasia procedure, the liver specimen was promptly immersed in a 10% formalin solution at room temperature for a duration of 24 hours. Subsequently, the tissues were immersed in paraffin, subjected to sectioning, and affixed onto glass microscope slides. The slices underwent staining with hematoxylin and eosin and were afterwards analyzed using light microscopy (Konstantopoulos et al., 2017).

154 Molecular docking simulation

155 Molecular docking experiments were done with the PyRx software (Dallakyan and Olson, 156 2015) to predict how metabolites, which are small-molecule ligands, bind to biological 157 macromolecules. The NCBI PubChem database (https://pubchem.nlm.nih.gov/, accessed on 158 3 May 2023) was used to derive the three-dimensional structure of water kefir metabolites 159 (Patel et al., 2022). Target proteins like NF-κB (PDB code: 1A3Q) and Keap1 (PDB code: 4L7B) 160 were obtained from the RCSB Protein Data Bank (http://www.rcsb.org/, accessed on 03 May 2023). The PyRx program was used to reduce protein, ligand converted to PDBQT (Asnawi et 161 162 al., 2023), then maximize GRID parameter (Asnawi et al. 2022) and perform docking study (Febrina et al., 2022). The BIOVIA Discovery Studio 2017 R2 program was used to view the 163 164 protein and ligand complex and distance (Ischak et al., 2023). The BIOVIA Discovery Studio 165 2017 R2 tool was also utilized to find protein active sites.

166 Statistical analysis

All of the information is displayed in the form of individual data points as well as the mean along with the standard error of the mean. The statistical analysis was carried out with the help of Minitab software (version 19.0), and to make comparisons between several different groups, a one-way analysis of variance with Dunnett's post hoc test was utilized. All statistical graphs were created with Microsoft Excel 2019 in their respective versions. The levels of significance that were considered to have been reached were *p < 0.05.

173 **RESULTS**

174 In vivo evaluation of hepatoprotective activity

175 The serum ALT levels, as the main parameter for the liver damage, were measured prior to 176 and following induction, as well as following treatment (Fig. 1). Meanwhile, after treatment 177 AST, TNF-α, and TGF-β levels were also evaluated. These findings were analyzed using a one-178 way ANOVA with a 95% confidence level and a two-sided confidence interval. A significant 179 rise in blood ALT levels demonstrated the establishment of an animal model with liver injury, 180 according to statistical analysis, following the administration of CCl₄. When compared to the 181 positive control group, ALT serum levels decreased significantly after two weeks of therapy 182 with curcuma extract or water kefir. The three doses of water kefir groups demonstrated 183 equivalent activity when curcuma extract was used as the standard treatment, and there was 184 no significant difference between the three doses of water kefir. When compared to the 185 positive control group, AST levels were also reduced dramatically following treatment with 186 curcuma extract or water kefir. TNF-a levels in the water kefir group were significantly lower 187 than in the positive control group at dosages of 35 and 50 mL/kg body weight. Even though 188 there was no statistically significant difference in TGF-β levels, the group that received the 189 treatment demonstrated a decrease in TGF-β levels.



Fig. 1. (a) Before induction, after induction, and after treatment serum ALT level; (b) After treatment serum AST level; (c) After treatment serum TNF- α level; (d) After treatment serum TGF- β level. * indicates a significantly different result when compared to the positive control group; # indicates a significantly different result when compared to the curcuma extract group; p < 0.05; n = 4 mice in each group.

191 A histological examination of a normal liver group revealed a typical central vein bordered 192 with endothelial cells and normal hepatocytes, resulting in hepatic cords of cells with distinct 193 cell borders and sinusoidal gaps (Figs 2A and 2B). The CCl4-induced group developed 194 centrilobular hepatic necrosis level 4, which was characterized by massive vacuolization and 195 necrosis of epithelial cells in the liver's centrilobular region (blue arrow) (Fig 2C & 2D). The 196 group that received either curcuma extract or water kefir treatment improved in varied 197 necrotic conditions ranging from level 1 (water kefir 50 mL/kg BW) (Fig 2K & 2L) to level 2 198 (curcuma extract (Fig 2E & 2F), water kefir 15 mL/kg BW (Figs 2G and 2H), and water kefir 199 30 mL/kg BW (Fig 2I & 2J)).



Fig. 2. Liver histology after CCl4 intoxication and treatment with HE staining. Negative control group (A & B), positive control group (C & D), curcuma extract group (E & F), water kefir 15 mL/kg BW (G & H), water kefir 30 mL/kg BW (I & J), and water kefir 50 mL/kg BW (K & L). 1: central vein; 2: normal liver epithelial cell; 3: portal track; blue arrow: necrosis and liver epithelial cell vacuolization at centrolobular region.

- 200
- 201
- 202

203 Molecular docking

204 Molecular docking studies are considered a powerful tool for predicting the potential 205 targets of bioactive molecules. In order to carry out molecular docking simulations, one of the 206 most critical steps is to identify the target active site. If the target protein is crystallized with a native ligand, in many instances, the location of the active site can be established without any 207 208 difficulty (Li et al., 2019b). However, the NF-KB (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) 209 proteins do not have a native ligand, so the active site was determined. Active site prediction 210 in docking is a computational method for predicting the location and orientation of a receptor 211 protein's binding site for a ligand molecule. The active site prediction was based on a protein structural analysis and the identification of amino acid residues that are likely to interact with 212 213 the ligand. The projected binding site is then utilized as a starting point for molecular docking, 214 a computer method for predicting a ligand molecule's binding affinity and orientation to a receptor protein. The active site prediction for target proteins (Keap1 and NF-KB) gives the 215 216 grid box coordinates (x y z) of 17.500880 Å, 62.323000 Å, and 0.973748 Å; and -1.751769 Å, -217 20.743853 Å, and -29.010438 Å, respectively (Fig. 3).



Fig. 3. Binding pocket (colored in red) generated using the BIOVIA Discovery Studio 2017 tool, coupled with the sequence containing the highlighted residues that constitute the binding pocket. A) NF-κB (PDB ID 1A3Q) and B) Keap1 (PDB ID 4L7B).

218

The docking results of the 25 metabolites could interact with target proteins (Keap1 and NF- κ B) (Table 1). In general, all metabolites could interact with both NF- κ B receptors (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) (Table 1). The interaction of metabolites with nuclear factor-kappa B receptors (PDB ID 1A3Q) revealed that 13 metabolites possessed free energy for binding that was greater than that of curcumin. On the other hand, there was not a single metabolite that had a free energy binding that was more powerful than that of nrf2 Keap1 (PDB ID 4L7B) (Table 1).

For volatile compounds, 2-phenyl ethanol and benzaldehyde interact most strongly with
the NF-κB receptor (PDB ID 1A3Q) with almost the same binding energy of -4.19 kcal/mol.
As for organic acids, succinic acid, fumaric acid, and citric acid provide nearly the same strong
interactions. Bond energy values of fumaric acid, succinic acid, and citric acid of -6.66, -6.24,
and -6.25 kcal/mol, respectively. In sugars, glucose provides the strongest interaction with the
NF-κB receptor (PDB ID 1A3Q) where the binding energy was -3.71 kcal/mol. As for the nrf2
Keap1 receptor (PDB ID 4L7B), 2-phenylacetaldehyde from volatile compounds was able to interact most strongly with the Keap1 receptor (PDB ID 4L7B) with a binding energy of -3.46 kcal/mol. As for organic acids, succinic acid provides nearly the same strong interaction with a bond energy value of -3.02 kcal/mol, respectively. In sugars, glucose provides the strongest interaction with the Keap1 receptor (PDB ID 4L7B) where the binding energy was -3.09 kcal/mol.

	No. Matabalitas	DDD, 1 4 2 O	DIDD, 41 7D	
239	кВ (PDB ID 1A3Q) and Keap1 (PD	OB ID 4L7B)		
238	Table 1. The free energy of bindin	ng and inhibition constant o	of water kefir metabolite active site int	eraction on NF-

110.	Metabolites	FDB: 1A3Q Free Energy of Binding, ΔG (kcal/mol)	Inhibition Constant, Ki (µM)	FDB: 4L/B Free Energy of Binding, ΔG (kcal/mol)	Inhibition Constant, Ki (μΜ)
	Volatile compounds				
1	2-Methylbutanol	-3.31	3,770	-2.68	10,890
2	3-Methylbutanol	-3.22	4,360	-2.50	14,670
3	2-Phenylethanol	-4.19	851.52	-3.12	5,140
4	Ethyl acetate	-3.19	4,590	-2.43	16,670
5	2-Methylbutanal	-3.26	4,050	-2.35	19,050
6	1-Octanol	-2.65	11,490	-2.11	28,220
7	2-Phenylethyl acetate	-3.99	1,190	-3.33	3,590
8	Benzyl acetate	-4.16	887.66	-3.44	3,020
9	Benzaldehyde	-4.19	853.74	-3.19	4,550
10	Furfuryl acetate	-3.60	2,320	-3.32	3,660
11	Isobutyl acetate	-3.19	4,610	-2.94	7,020
12	Isopentyl acetate	-3.20	4,510	-3.05	5,820
13	2-Phenylacetaldehyde	-4.07	1,030	-3.46	2,930
14	3-Methylbutanal	-3.16	4,850	-2.73	10,020
15	Gluconic acid	-3.11	5,280	-1.63	64,290
	Organic acids				
16	Lactic acid	-4.92	249.28	-2.06	30,680
17	Succinic acid	-6.24	26.68	-3.02	6,140
18	Fumaric acid	-6.66	13.22	-2.45	16,100
19	Citric acid	-6.25	26.18	-1.87	42,440
20	Malic acid	-5.65	72.69	-2.53	13,860
21	Acetic acid	-4.99	219.33	-2.44	16,280
22	Ethanol	-2.39	17,800	-2.08	29,800
	Sugars				
23	Sucrose	-1.40	93,710	-1.33	105,320
24	Glucose	-3.71	1,900	-3.09	5,440
25	Fructose	-3.21	4,400	-2.65	11,440
	Reference compound				
	Curcumin	-3.44	2,990	-4.22	811.36

240

241 The theoretical binding modes of the top three metabolites with their target proteins (Keap1 242 and NF-KB) are shown in Figs. 4 and 5, respectively. The molecular docking results suggested 243 that these metabolites interacted with the Keap1 and NF-KB to form a complex through hydrogen bonds with various residues. The interaction of 2-phenyl ethanol with the active site 244 245 of NF-kB was formed by two hydrogen bonds and one Pi-Alkyl interaction with amino acid 246 residues of PRO223, LYS252, and LYS252, respectively. The interaction of benzaldehyde with 247 the active site of NF-KB was formed by a hydrogen bond with the amino acid residue of 248 LYS252. The interaction of fumaric acid with the active site of NF-KB was formed by six 249 hydrogen bonds with the amino acid residues of LYS221, LYS252, and TYR285. The interaction 250 of glucose with the active site of NF-kB was formed by six hydrogen bonds with the amino 251 acid residues SER220, LYS221, SER226, and LYS252. Although glucose and fumaric acid were 252 able to be formed by six hydrogen bonds, different types of amino acid residues were involved

- 253 254 in the interaction, so fumaric acid interacted more strongly with the active site of NF- κ B (Fig.
- 4).
- 255



Fig. 4. 2D and 3D representation of the kind of interaction on the formation of the NF-κB active site (PDB ID 1A3Q). A) 2-Phenylethanol, B) Benzaldehyde, C) Fumaric acid, D) Glucose, and E) Curcumin.

256 257



Fig. 5. 2D and 3D illustration of the type of interaction on the formation of Keap1's binding pocket (PDB ID 4L7B). A) 2-Phenylacetaldehyde, B) Succinic acid, C) Glucose, and D) Curcumin.

258

259 The interaction of 2-phenylacetaldehyde with the active site of Keap1 was formed by one 260 hydrogen bond and one Pi-Alkyl interaction with amino acid residues of GLN530 and 261 ALA556, respectively. The interaction of succinic acid with the active site of Keap1 was formed 262 by four hydrogen bonds with the amino acid residues of ARG336, HIS575, and PHE577. The 263 interaction of glucose with the active site of Keap1 was formed by six hydrogen bonds with 264 the amino acid residues LYS551, ARG553, and ASP573. Although glucose can form more hydrogen bonds than 2-phenylacetaldehyde, the different types of amino acid residues 265 involved have not been able to have a significant effect on the binding energy of its interaction 266 267 with the active site of Keap1 (Fig. 5).

268 Curcumin (the reference compound) created three hydrogen bonds with the amino acid 269 residues ARG52, GLU58, and LYS252 to interact with the active site of NF-KB. Meanwhile, 270 curcumin established three hydrogen bonds with the amino acid residues ARG336, TYR572, 271 and GLY600 to interact with the active site of nrf2 Keap1 (Figs. 4 and 5). Despite the fact that 272 curcumin could create three hydrogen bonds at both the active sites of NF- κ B and nrf2 Keap1, 273 its interaction with nrf2 Keap1 had a lower free energy of binding than phenylethanol, 274 benzaldehyde, and fumaric acid (Table 1). Overall, the metabolites of water kefir prefer to 275 interact with NF-KB and nrf2 Keap1 receptors. Whereas fumaric acid and 2-276 phenylacetaldehyde were metabolites that had the strongest interaction with NF-KB (PDB ID 277 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) receptors, respectively.

278

279 **DISCUSSION**

280 Increased liver enzyme production is one of the abnormalities indicating liver damage. This 281 increase in secretion increases the leakage of enzymes into the circulation. Transaminase is a 282 frequent enzyme used to evaluate liver disorders. The transaminases AST and ALT catalyze 283 the transfer of the α -amino group from alanine and aspartic acid to α -ketoglutaric acid. AST 284 is found in the liver, heart, and skeletal muscle, as well as the kidney, brain, pancreas, lung, 285 leukocytes, and erythrocytes. While ALT is predominantly found in the liver, it is also found 286 in low concentrations in other tissues (Lee et al., 2012). Consequently, ALT was used as the 287 principal hepatotoxicity criterion in this study.

288 Carbon tetrachloride (CCl_4) is a component of the hepatotoxin, which acts after metabolic activation and is extensively employed as a liver-damaging agent. In this study, the 289 290 administration of CCl₄ to animals caused hepatotoxicity, which was characterized by a 291 significant increase in ALT after several administrations. CCl₄ is metabolized by the enzyme 292 cytochrome p450 (CYP2E1) in the endoplasmic reticulum to a highly reactive trichloromethyl 293 radical. The trichloromethyl radical rapidly reacts with oxygen to form the trichloromethyl 294 peroxyl radical, which rapidly reacts with lipids to form lipid peroxidation products. Free 295 radicals will disrupt the endoplasmic reticulum (ER) and lead to lipid accumulation, decreased 296 protein synthesis, and increased oxidase activity. CCl₄ hepatotoxicity is characterized by 297 hepatocellular necrosis with fat deposition (Ritesh et al., 2015). At the molecular level, 298 administration of CCl₄ can activate tumor necrosis factor (TNF)-a, nitric oxide (NO), and 299 transforming growth factor (TGF)- α and - β in cells, processes that precipitate cell self-300 destruction or fibrosis. TNF-a leads to apoptosis, whereas TGF-B leads to fibrosis (Weber et 301 al., 2003).

In terms of its pathophysiological underpinnings, liver illness is linked to a condition knownas dysbiosis, which refers to an imbalance in the makeup of the gut microbiota (Zavala et al.,

304 2016; Laureys and De Vuyst, 2017; Romero-Luna et al., 2020). Both qualitative and quantitative 305 changes in the gut microbiome have the potential to affect the composition of products 306 produced by the microbiota, such as short-chain fatty acids and bile acids (Romero-Luna et 307 al., 2020). Qualitative changes include an imbalance between harmful and helpful 308 microbiomes, whereas numeric changes involve changes to the overall microbiota. In addition 309 to these symptoms, intestinal inflammation, disruption of the intestinal barrier, and the transfer of microbial products can all be caused by dysbiosis (Laurevs et al., 2018). For this 310 311 reason, the condition of the gut microbiome is an important factor in the initiation and 312 development of chronic liver disease (Lee et al., 2021). Based on the results of the study, 313 treatment with water kefir for 2 weeks after the occurrence of liver damage was able to 314 improve the overall condition of the liver, which was marked by a significant decrease in the 315 values of AST, ALT, TNF-α, TGF-β, and significant improvement in liver histology.

316 Water kefir contains a number of microorganisms that have been linked to health benefits, 317 such as the probiotics *L. paracasei* and *B. cereus* (Fijan, 2014). This activity is linked to an increase in antioxidants like glutathione and catalase and a decrease in pro-inflammatory transcription 318 319 factors like nuclear B-factor, lipopolysaccharide, and Toll-like receptor 4 (TLR-4). 320 Improvements in intestinal barrier function and histological integrity were also observed. 321 Increased expression of claudin-1 and occludin-1, two components of tight junctions, occurs 322 simultaneously with the restoration of the p38 MAPK pathway (Fijan, 2014; Yao et al., 2019; Tsai et al., 2020; Ji et al., 2022). Bacillus is a kind of endospore-forming bacterium that can 323 324 endure extremely cold temperatures and lengthy periods of storage without dying; its spores 325 can even tolerate the acidic environment of the stomach and make it all the way to the small 326 intestine (Elshaghabee et al., 2017). Bacillus cereus has been shown to reduce ALT levels, an indicator of liver healing, in various animal models of liver injury. It protects the liver by 327 328 reducing inflammation, enhancing the gut flora, and strengthening the tight junctions in the 329 intestines (Kim et al., 2018; Li et al., 2019a; Xue et al., 2020). Also, when Bacillus spores were 330 used first, hepatocyte necrosis and serum levels of ALT, ZO-1, AST, and TAC went down by 331 a lot. This effect is comparable to that of the popular hepatoprotective compound silymarin 332 (Neag et al., 2020).

333 Beer, vinegar, cider, and cocoa bean masses are just some of the places where Gram-334 negative acetic acid bacteria have been found. One of the features that sets acetic acid bacteria 335 apart from others is their alkaline-stable lipid membrane (Lynch et al., 2021). Their "oxidative" 336 fermentation metabolism is responsible for the principal metabolic process in these bacteria, 337 the oxidation of ethanol to acetic acid. Important in the food and beverage sector and beyond, 338 fermentation helps mediate the transition of diverse substrates into products. Although lactic 339 acid bacteria have been studied more extensively than acetic acid bacteria (Semjonovs et al., 340 2014; Hong et al., 2021), various studies have shown promising results concerning the 341 pharmacological effects of acetic acid bacteria, especially as a hepatoprotective agent. Acetic 342 acid, which is the main metabolite of acetic acid bacteria, can lower inflammation and the 343 severity of liver injury in rats with septic shock by increasing the expression of TRIM40. 344 TRIM40 has been shown to minimize liver damage, decrease the synthesis and release of 345 cytokines such as IL-6 and TNF-a, raise the expression of IL-10, improve survival in septic mice, and block the activity of the TLR4 signaling pathway. Acetic acid injection decreased 346 347 inflammation as well as the production of inflammatory cytokines (Yang et al., 2019). Acetic 348 acid also stimulates the AMPK signaling pathway, which leads to enhanced lipid oxidation 349 and reduced hepatic lipid and body fat deposition (Kondo et al., 2009; Li et al., 2018).

Apart from microorganisms that directly provide hepatoprotective effects, the metabolites produced from these microorganisms also have the potential to be hepatoprotective. 352 Molecular docking is a technique that is utilized in the context of NF- κ B and Nrf2 to make 353 predictions regarding the binding affinity and orientation of small-molecule inhibitors to their 354 active sites. The transcription factor known as NF-kB is an essential component in the 355 management of both the immune system and the inflammatory response (Dai et al., 2021). The 356 expression of important inflammatory genes can be inhibited by small-molecule inhibitors that 357 impair the interaction between NF-KB and DNA. These inhibitors have the potential to be used 358 in therapeutic applications. The Nrf2 binding site is Kelch-like ECH-associated protein 1 in the 359 context of nuclear factor erythroid 2-related factor 2, also known as Nrf2 (Keap1) (Zhao et al., 360 2017; Jiang et al., 2019). Small-molecule inhibitors that disrupt the link between Keap1 and 361 Nrf2 can activate the Nrf2-ARE signaling pathway, which has been demonstrated to have 362 cytoprotective effects. Keap1 is a negative regulator of Nrf2, and this association can be 363 disrupted by small-molecule inhibitors (Zhao et al., 2017).

364 Binding energy and Ki are important parameters used in molecular docking to evaluate the 365 strength of the interaction between a ligand and a receptor protein. Binding energy is the 366 energy released when a ligand binds to a receptor protein, and it is calculated as the difference 367 between the energy of the bound complex and the energy of the unbound ligand and protein 368 (Meng et al., 2011). Ki, on the other hand, is the dissociation constant of the ligand-receptor 369 complex. It shows how much ligand is needed to bind 50% of the receptor binding sites. Both 370 binding energy and Ki are used to predict the binding affinity and selectivity of a ligand to a 371 receptor protein (Du et al., 2016). Ki, on the other hand, is the dissociation constant of the 372 ligand-receptor complex. It shows how much ligand is needed to bind 50% of the receptor 373 binding sites (Du et al., 2016).

374 Based on the results of an in silico study of water kefir metabolite compounds, it was known 375 that fumaric acid and 2-phenylacetaldehyde have the strongest interactions with NF-KB (PDB 376 ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) receptors, respectively. Fumaric acid has been 377 studied for its potential as a hepatoprotective agent. Fumaric acid protects rat livers against 378 cadmium toxicity by decreasing oxidative stress and improving hepatic serum. Fumaric acid 379 prevented cadmium-accrued hepatocellular damage and showed the potential to avert hepatic 380 injury against cadmium in rats (Kaur et al., 2020). Fumaric acid esters were found to ameliorate inflammation and oxidative stress in rats, which was associated with less adiposity and ectopic 381 382 fat accumulation (Šilhavý et al., 2014).

383 NF-KB and Nrf2 are two transcription factors that play important roles in regulating 384 inflammation and cell survival. While NF-kB is involved in the inflammatory response, and 385 Nrf2 is involved in the antioxidant response (Ganesh Yerra et al., 2013; Wang et al., 2022a). 386 Both transcription factors have been investigated as potential targets for the development of 387 hepatoprotective agents (Rahman et al., 2021; Gao et al., 2022; Wang et al., 2022a; Li et al., 2023). There is evidence of crosstalk between the Nrf2 and NF-kB pathways (Ganesh Yerra et 388 389 al., 2013; Gao et al., 2022). The Nrf2 pathway inhibits the activation of the NF-κB pathway by 390 increasing antioxidant defenses and HO-1 expression, which efficiently neutralizes ROS and 391 detoxifies (Ganesh Yerra et al., 2013). The crosstalk between Nrf2 and NF-kB could be a new 392 therapeutic target against hepatotoxicity (Gao et al., 2022). Researchers have tried to identify 393 molecule activators of Nrf2 as chemoprevention for ROS-dependent carcinogenesis, while 394 others have focused on identifying Nrf2 inhibitors to increase the sensitivity of cancer cells to 395 chemotherapy (Sharifi-Rad et al., 2023). While NF-KB and Nrf2 are involved in different 396 cellular processes, they have both been investigated as potential targets for the development 397 of hepatoprotective agents. Molecular docking studies have been used to investigate the 398 interaction of potential hepatoprotective agents with these transcription factors. There is also evidence of crosstalk between the Nrf2 and NF-κB pathways, which could be a newtherapeutic target against hepatotoxicity.

401 In the pathogenesis of hepatocellular damage, liver fibrosis, and HCC, NF-κB plays a 402 crucial role in regulating inflammation and cell death (Luedde and Schwabe, 2011). In 403 response to many stimuli that may pose a threat to the host, NF-KB is activated, setting in 404 motion processes such as inflammation, immunity, wound healing, and pathogen clearance 405 (Luedde and Schwabe, 2011). Pathogen-derived chemicals that activate Toll-Like Receptors 406 (TLRs) include lipopolysaccharide (LPS), viral and bacterial DNA and RNA, and 407 inflammatory cytokines, including tumor necrosis factor (TNF) and interleukins (IL)-1 408 (Luedde and Schwabe, 2011). When NF-B is activated, a lot of genes with B-binding sites are 409 transcribed. These genes play important roles in controlling inflammation, the immune 410 response, and cell survival. In an NF-KB-dependent manner, lipopolysaccharide (LPS) 411 activates TLR4 on dormant HSCs by reducing BAMBI expression (an inhibitory TGF-β 412 pseudoreceptor) and increasing Kupffer cell chemotaxis. Due to low levels of BAMBI, 413 recruited Kupffer cells secrete TGF-β, which stimulates HSCs unrestrictively. When HSCs 414 have been activated, NF-KB serves a second crucial function by increasing their chances of 415 survival. LPS, Kupffer cell-derived mediators (such as IL-1 and TNF), and angiotensin II, which is produced and acts on HSCs in an autocrine way, all play roles in activating NF-KB in 416 417 activated hepatic stellate cells. More activated HSCs and extracellular matrix are deposited in 418 the liver as a result of greater HSC activation and survival (Luedde and Schwabe, 2011).

419 TNF- α and TGF- β can stimulate one another's production, and TNF- α has been shown to 420 influence TGF-β expression in a variety of cells and tissues (Liu et al., 2022). TNF-α is an 421 inflammatory cytokine that contributes to liver inflammation, and chronic liver inflammation 422 results in liver fibrosis. TNF-a exerts its effects on liver fibrosis via multiple mechanisms, 423 including TGF- β signaling activation (Yang and Seki, 2015). Targeting TNF- α and TGF- β 424 signaling pathways may, therefore, have therapeutic potential for treating liver diseases. In 425 regards to hepatoprotective effects, the relationship between TGF-β and TNF-α is complex and 426 not completely understood.

427 CONCLUSION

428 This study evaluated the hepatoprotective qualities of Indonesian water kefir in rats with 429 CCl4-induced liver damage. Water kefir administration improved the condition of liver 430 damage, characterized by a decrease in serum levels of AST, ALT, TNF, TGF, and an 431 improvement of liver tissue profile. *In silico* evaluation showed that the metabolites in water 432 kefir were able to interact with target proteins in the NF-κB and Nrf2 pathways. It was 433 concluded that water kefir improves the condition of the liver by reducing the level of necrosis 434 and fibrosis.

435

436 CONFLICT OF INTEREST

437 The authors declare no conflict of interest.

438 ACKNOWLEDGMENT

439 This research was funded by the Center of Research and Community Service, Bhakti Kencana University, 440 Bandung, West Java, Republic of Indonesia (052/14.LPPM/PE.I/LPPM/2021)

441	REFERENCES
442	Aligita W, Alex V, Taaraungan S, Susilawati E (2021) Hepatoprotective activity of water kefir.
443	Int J Biol Pharm Allied Sci 10: 1784-1794.
444	https://doi.org/10.31032/IJBPAS/2021/10.6.5493
445	Aligita W, Tarigan PN, Susilawati E (2020) Anti Inflammatory and antioxidant activity of kefir
446	water. Int J Biol Pharm Allied Sci 9: 2454-2464.
447	https://doi.org/10.31032/IJBPAS/2020/9.1.4904
448	Al-Mohammadi A-R, Ibrahim RA, Moustafa AH, Ismaiel AA, Zeid AA, Enan G (2021)
449	Chemical constitution and antimicrobial activity of kefir fermented beverage.
450	Molecules 26:2635. https://doi.org/10.3390/molecules26092635
451 452 453 454	Alsayadi M, Jawfi YA, Belarbi M, Soualem-Mami Z, Merzouk H, Sari DC, Sabri F, Ghalim M (2014) Evaluation of anti-hyperglycemic and anti-hyperlipidemic activities of water kefir as probiotic on streptozotocin-induced diabetic wistar rats. Journal of Diabetes Mellitus 04:85–95. https://doi.org/10.4236/jdm.2014.42015
455	Asnawi A, Aman LO, Nursamsiar, Yuliantini A, Febrina E (2022) Molecular docking and
456	molecular dynamic studies: Screening phytochemicals of <i>Acalypha indica</i> against BRAF
457	kinase receptors for potential use in melanocytic tumours. Rasayan J Chem 15: 1352–
458	1361. https://doi.org/10.31788/RJC.2022.1526769
459 460 461	Asnawi A, Febrina E, Aligita W, Kurnia D, Aman LO, Yuliantini A (2022) Screening Of ashitaba (<i>Angelica keiskei</i> K.) compounds as potential <i>Mycobacterium tuberculosis</i> kasa inhibitors. Int J App Pharm 80–85. https://doi.org/10.22159/ijap.2022.v14s5.13
462	Asnawi A, Nedja M, Febrina E, Purwaniati (2023) Prediction of a stable complex of compounds
463	in the ethanol extract of celery leaves (<i>Apium graveolens</i> L.) function as a VKORC1
464	antagonist. Trop J Nat Prod Res 7:2362–2370. https://doi.org/10.26538/tjnpr/v7i2.10
465	Aspiras BEE, Flores RFAC, Pareja MC (2015) Hepatoprotective effect of fermented water kefir
466	on sprague-dawley rats (<i>Rattus norvegicus</i>) induced with sublethal dose of
467	acetaminophen. Int J Curr Sci 17:18–28
468	Calatayud M, Börner RA, Ghyselinck J, Verstrepen L, De Medts J, Van den Abbeele P,
469	Boulangé CL, Priour S, Marzorati M, Damak S (2021) Water kefir and derived
470	pasteurized beverages modulate gut microbiota, intestinal permeability and cytokine
471	production in vitro. Nutrients 13:3897. https://doi.org/10.3390/nu13113897
472	Dai C, Li H, Wang Y, Tang S, Velkov T, Shen J (2021) Inhibition of oxidative stress and ALOX12
473	and NF-KB pathways contribute to the protective effect of baicalein on carbon
474	tetrachloride-induced acute liver injury. Antioxidants 10:976.
475	https://doi.org/10.3390/antiox10060976
476	Dallakyan S, Olson AJ (2015) Small-Molecule Library Screening by Docking with PyRx. In:
477	Hempel JE, Williams CH, Hong CC (eds) Chemical Biology. Springer New York, New
478	York, pp 243–250
479	Darvishzadeh P, Orsat V, Martinez JL (2021) Process optimization for development of a novel
480	water kefir drink with high antioxidant activity and potential probiotic properties from

- 481 russian olive fruit (*Elaeagnus angustifolia*). Food Bioprocess Technol 14:248–260.
 482 https://doi.org/10.1007/s11947-020-02563-1
- 483 De Souza Basso B, Haute GV, Ortega-Ribera M, Luft C, Antunes GL, Bastos MS, Carlessi LP,
 484 Levorse VG, Cassel E, Donadio MVF, Santarém ER, Gracia-Sancho J, Rodrigues De
 485 Oliveira J (2021) Methoxyeugenol deactivates hepatic stellate cells and attenuates liver
 486 fibrosis and inflammation through a PPAR-γ and NF-kB mechanism. Journal of
 487 Ethnopharmacology 280:114433. https://doi.org/10.1016/j.jep.2021.114433
- 488 Diniz RO, Garla LK, Schneedorf JM, Carvalho JCT (2003) Study of anti-inflammatory activity
 489 of Tibetan mushroom, a symbiotic culture of bacteria and fungi encapsulated into a
 490 polysaccharide matrix. Pharmacological Research 47:49–52.
 491 https://doi.org/10.1016/S1043-6618(02)00240-2
- 492 Du X, Li Y, Xia YL, Ai SM, Liang J, Sang P, Ji XL, Liu SQ (2016) Insights into protein-ligand
 493 interactions: mechanisms, models, and methods. International Journal of Molecular
 494 Sciences 17:144. https://doi.org/10.3390/ijms17020144
- Elshaghabee FMF, Rokana N, Gulhane RD, Sharma C, Panwar H (2017) Bacillus as potential
 probiotics: status, concerns, and future perspectives. Front Microbiol 8:1490.
 https://doi.org/10.3389/fmicb.2017.01490
- Febrina E, Alamhari RK, Asnawi A, Abdulah R, Lestari K, Levita J, Supratman U (2021)
 Molecular docking and molecular dynamics studies of *Acalypha indica* L.
 phytochemical constituents with caspase-3. Int J App Pharm 13:210–215. https://doi.org/10.22159/ijap.2021.v13s4.43861
- Febrina E, Asnawi A, Abdulah R, Lestari K, Supratman U (2022) Identification of flavonoids
 from Acalypha indica L. (Euphorbiaceae) as caspase-3 activators using molecular docking
 and molecular dynamics. Int J App Pharm 14:162–166.
 https://doi.org/10.22159/ijap.2022.v14s5.34
- Fijan S (2014) Microorganisms with claimed probiotic properties: an overview of recent
 literature. International Journal of Environmental Research and Public Health 11:4745 4767. https://doi.org/10.3390/ijerph110504745
- Gamba RR, Yamamoto S, Sasaki T, Michihata T, Mahmoud AH, Koyanagi T, Enomoto T (2019)
 Microbiological and functional characterization of kefir grown in different sugar
 solutions. Food Science and Technology Research 25:303–312.
 https://doi.org/10.3136/fstr.25.303
- 513 Ganesh Yerra V, Negi G, Sharma SS, Kumar A (2013) Potential therapeutic effects of the
 514 simultaneous targeting of the Nrf2 and NF-κB pathways in diabetic neuropathy. Redox
 515 Biology 1:394–397. https://doi.org/10.1016/j.redox.2013.07.005
- 516 Gao W, Guo L, Yang Y, Wang Y, Xia S, Gong H, Zhang BK, Yan M (2022) Dissecting the
 517 crosstalk between Nrf2 and NF-κB response pathways in drug-induced toxicity. Front
 518 Cell Dev Biol 9:809952. https://doi.org/10.3389/fcell.2021.809952
- Hong Y, Sheng L, Zhong J, Tao X, Zhu W, Ma J, Yan J, Zhao A, Zheng X, Wu G, Li B, Han B,
 Ding K, Zheng N, Jia W, Li H (2021) Desulfovibrio vulgaris, a potent acetic acid-

- producing bacterium, attenuates nonalcoholic fatty liver disease in mice. Gut Microbes
 13:1930874. https://doi.org/10.1080/19490976.2021.1930874
- Ischak NI, Aman LO, Hasan H, Kilo AL, Asnawi A (2023) In silico screening of *Andrographis paniculata* secondary metabolites as anti-diabetes mellitus through PDE9 inhibition.
 Res Pharm Sci 18:100–111. https://doi.org/10.4103/1735-5362.363616
- Ji Y, Xie Q, Meng X, Wang W, Li S, Lang X, Zhao C, Yuan Y, Ye H (2022) Lactobacillus paracasei
 improves dietary fatty liver by reducing insulin resistance and inflammation in obese
 mice model. Journal of Functional Foods 95:105150.
 https://doi.org/10.1016/j.jff.2022.105150
- Jiang ZY, Lu MC, You QD (2019) Nuclear factor erythroid 2-related factor 2 (Nrf2) inhibition:
 an emerging strategy in cancer therapy. J Med Chem 62:3840–3856.
 https://doi.org/10.1021/acs.jmedchem.8b01121
- Kaur G, Shivanandappa TB, Kumar M, Kushwah AS (2020) Fumaric acid protect the cadmiuminduced hepatotoxicity in rats: owing to its antioxidant, anti-inflammatory action and
 aid in recast the liver function. Naunyn-Schmiedeberg's Arch Pharmacol 393:19111920. https://doi.org/10.1007/s00210-020-01900-7
- Kebler LF (1921) California Bees. The Journal of the American Pharmaceutical Association
 10:939–943. https://doi.org/10.1002/jps.3080101206
- Kim B, Kwon J, Kim MS, Park H, Ji Y, Holzapfel W, Hyun CK, Aguila MB (2018) Protective
 effects of *Bacillus* probiotics against high-fat diet-induced metabolic disorders in mice.
 PLoS ONE 13:e0210120. https://doi.org/10.1371/journal.pone.0210120
- Kondo T, Kishi M, Fushimi T, Kaga T (2009) Acetic acid upregulates the expression of genes
 for fatty acid oxidation enzymes in liver to suppress body fat accumulation. J Agric
 Food Chem 57:5982–5986. https://doi.org/10.1021/jf900470c
- Konstantopoulos P, Doulamis I, Tzani A, Korou ML, Agapitos E, Vlachos I, Pergialiotis V,
 Verikokos C, Mastorakos G, Katsilambros N, Perrea D (2017) Metabolic effects of *Crocus sativus* and protective action against non-alcoholic fatty liver disease in diabetic
 rats. Biom Rep 6: 513-518. https://doi.org/10.3892/br.2017.884
- Laureys D, Aerts M, Vandamme P, De Vuyst L (2018) Oxygen and diverse nutrients influence
 the water kefir fermentation process. Food Microbiology 73:351–361.
 https://doi.org/10.1016/j.fm.2018.02.007
- Laureys D, De Vuyst L (2017) The water kefir grain inoculum determines the characteristics of
 the resulting water kefir fermentation process. J Appl Microbiol 122:719–732.
 https://doi.org/10.1111/jam.13370
- Laureys D, De Vuyst L (2014) Water kefir as a promising low-sugar probiotic fermented
 beverage. Arch Public Health 72:P1. https://doi.org/10.1186/2049-3258-72-S1-P1
- Lee NY, Shin MJ, Youn GS, Yoon SJ, Choi YR, Kim HS, Gupta H, Han SH, Kim BK, Lee DY,
 Park TS, Sung H, Kim BY, Suk KT (2021) *Lactobacillus* attenuates progression of
 nonalcoholic fatty liver disease by lowering cholesterol and steatosis. Clin Mol Hepatol
 27:110–124. https://doi.org/10.3350/cmh.2020.0125

- Lee TH, Kim WR, Poterucha JJ (2012) Evaluation of elevated liver enzymes. Clinics in Liver
 Disease 16:183–198. https://doi.org/10.1016/j.cld.2012.03.006
- Li H, Weng Q, Gong S, Wang J, Huang Y, Li Y, Guo J, Lan T (2023) Kaempferol prevents
 acetaminophen-induced liver injury by suppressing hepatocyte ferroptosis *via* Nrf2
 pathway activation. Food Funct 14:1884–1896. https://doi.org/10.1039/D2FO02716J
- Li L, He M, Xiao H, Liu X, Wang K, Zhang Y (2018) Acetic acid influences BRL-3A cell lipid
 metabolism via the AMPK signalling pathway. Cell Physiol Biochem 45:2021–2030.
 https://doi.org/10.1159/000487980
- Li YT, Ye JZ, Lv LX, Xu H, Yang LY, Jiang XW, Wu WR, Shi D, Fang DQ, Bian XY, Wang KC,
 Wang QQ, Xie JJ, Lu YM, Li LJ (2019a) Pretreatment with *Bacillus cereus* preserves
 against D-galactosamine-induced liver injury in a rat model. Front Microbiol 10:1751.
 https://doi.org/10.3389/fmicb.2019.01751
- Li Z, Huang Y, Wu Y, Chen J, Wu D, Zhan CG, Luo HB (2019b) Absolute binding free energy
 calculation and design of a subnanomolar inhibitor of phosphodiesterase-10. J Med
 Chem 62:2099–2111. https://doi.org/10.1021/acs.jmedchem.8b01763
- Liu Z, Zhang Y, Zhang L, Zhou T, Li YY, Zhou GC, Miao ZM, Shang M, He JP, Ding N, Liu
 YQ (2022) Duality of interactions between TGF-β and TNF-α during tumor formation.
 Front Immunol 12:810286. https://doi.org/10.3389/fimmu.2021.810286
- 579Luedde T, Schwabe RF (2011) NF-κB in the liver—linking injury, fibrosis and hepatocellular580carcinoma.NatRevGastroenterolHepatol8:108-118.581https://doi.org/10.1038/nrgastro.2010.213
- Lynch KM, Wilkinson S, Daenen L, Arendt EK (2021) An update on water kefir: microbiology,
 composition and production. International Journal of Food Microbiology 345:109128.
 https://doi.org/10.1016/j.ijfoodmicro.2021.109128
- Meng XY, Zhang HX, Mezei M, Cui M (2011) Molecular docking: a powerful approach for
 structure-based drug discovery. Current Computer Aided-Drug Design 7:146–157.
 https://doi.org/10.2174/157340911795677602
- Moinas M, Horisberger M, Bauer H (1980) The structural organization of the tibi grain as
 revealed by light, scanning and transmission microscopy. Arch Microbiol 128:157–161.
 https://doi.org/10.1007/BF00406153
- Moreira MEC, Santos MHD, Zolini GPP, Wouters ATB, Carvalho JCT, Schneedorf JM (2008)
 Anti-inflammatory and cicatrizing activities of a carbohydrate fraction isolated from
 sugary kefir. Journal of Medicinal Food 11:356–361.
 https://doi.org/10.1089/jmf.2007.329
- Neag MA, Catinean A, Muntean DM, Pop MR, Bocsan CI, Botan EC, Buzoianu AD (2020)
 Probiotic *Bacillus* spores protect against acetaminophen induced acute liver injury in rats. Nutrients 12:632. https://doi.org/10.3390/nu12030632
- Nursamsiar, Nur S, Febrina E, Asnawi A, Syafiie S (2022) Synthesis and inhibitory activity of
 curculigoside a derivatives as potential anti-diabetic agents with β-cell apoptosis.

- 600
 Journal
 of
 Molecular
 Structure
 1265:133292.

 601
 https://doi.org/10.1016/j.molstruc.2022.133292

 1265:133292.
- Parasuraman S, Raveendran R, Kesavan (2010) Blood sample collection in small laboratory
 animals. Journal of Pharmacology and Pharmacotherapeutics 1:87–93.
 https://doi.org/10.4103/0976-500X.72350
- Patel SH, Tan JP, Börner RA, Zhang SJ, Priour S, Lima A, Ngom-Bru C, Cotter PD, Duboux S
 (2022) A temporal view of the water kefir microbiota and flavour attributes. Innovative
 Food Science & Emerging Technologies 80:103084.
 https://doi.org/10.1016/j.ifset.2022.103084
- Rahman ZU, Al Kury LT, Alattar A, Tan Z, Alshaman R, Malik I, Badshah H, Uddin Z, Khan
 Khalil AA, Muhammad N, Khan S, Ali A, Shah FA, Li JB, Li S (2021) Carveol a
 naturally-derived potent and emerging Nrf2 activator protects against acetaminopheninduced hepatotoxicity. Front Pharmacol 11:621538.
 https://doi.org/10.3389/fphar.2020.621538
- Ritesh KR, Suganya A, Dileepkumar HV, Rajashekar Y, Shivanandappa T (2015) A single acute
 hepatotoxic dose of CCl 4 causes oxidative stress in the rat brain. Toxicology Reports
 2:891–895. https://doi.org/10.1016/j.toxrep.2015.05.012
- Rocha-Gomes A, Escobar A, Soares JS, Da Silva AA, Dessimoni-Pinto NA, Riul TR (2018)
 Chemical composition and hypocholesterolemic effect of milk kefir and water kefir in
 Wistar rats. Rev Nutr 31:137–145. https://doi.org/10.1590/1678-98652018000200001
- Rodrigues KL, Araújo TH, Schneedorf JM, Ferreira CS, Moraes GO, Coimbra RS, Rodrigues
 MR (2016) A novel beer fermented by kefir enhances anti-inflammatory and antiulcerogenic activities found isolated in its constituents. Journal of Functional Foods 21:58–69. https://doi.org/10.1016/j.jff.2015.11.035
- Romero-Luna HE, Peredo-Lovillo A, Hernández-Mendoza A, Hernández-Sánchez H, Cauich Sánchez PI, Ribas-Aparicio RM, Dávila-Ortiz G (2020) Probiotic potential of
 Lactobacillus paracasei CT12 isolated from water kefir grains (tibicos). Curr Microbiol
 77:2584-2592. https://doi.org/10.1007/s00284-020-02016-0
- Semjonovs P, Denina I, Linde R (2014) Evaluation of physiological effects of acetic acid bacteria
 and yeast fermented non-alchocolic beverage consumption in rat model. J of Medical
 Sciences 14:147–152. https://doi.org/10.3923/jms.2014.147.152
- Sharifi-Rad J, Seidel V, Izabela M, Monserrat-Mequida M, Sureda A, Ormazabal V, Zuniga FA,
 Mangalpady SS, Pezzani R, Ydyrys A, Tussupbekova G, Martorell M, Calina D, Cho
 WC (2023) Phenolic compounds as Nrf2 inhibitors: potential applications in cancer
 therapy. Cell Commun Signal 21:89. https://doi.org/10.1186/s12964-023-01109-0
- 635 Šilhavý J, Zídek V, Mlejnek P, Landa V, Šimáková M, Strnad H, Olivarnyk O, Škop V, Kazdová L, Kurtz T, Pravenec M, Sookoian SC (2014) Fumaric acid esters can block pro-636 inflammatory actions of human CRP and ameliorate metabolic disturbances in 637 638 *s*pontaneously hypertensive PLoS 9:e101906. transgenic rats. ONE 639 https://doi.org/10.1371/journal.pone.0101906

- 640 Tsai YS, Lin SW, Chen YL, Chen CC (2020) Effect of probiotics Lactobacillus paracasei GKS6, L. 641 plantarum GKM3, and L. rhamnosus GKLC1 on alleviating alcohol-induced alcoholic 642 Nutr liver disease in а mouse model. Res Pract 14:299. 643 https://doi.org/10.4162/nrp.2020.14.4.299
- Wang B, Cui S, Mao B, Zhang Q, Tian F, Zhao J, Tang X, Chen W (2022a) Cyanidin alleviated
 CCl₄-induced acute liver injury by regulating the Nrf2 and NF-κB signaling pathways.
 Antioxidants 11:2383. https://doi.org/10.3390/antiox11122383
- Wang Y, Liu F, Liu M, Zhou X, Wang M, Cao K, Jin S, Shan A, Feng X (2022b) Curcumin
 mitigates aflatoxin B1-induced liver injury via regulating the NLRP3 inflammasome
 and Nrf2 signaling pathway. Food and Chemical Toxicology 161:112823.
 https://doi.org/10.1016/j.fct.2022.112823
- Ward M (1892) The ginger-beer plant, and the organisms composing it: a contribution to the
 study of fermentation-yeasts and bacteria. Phil Trans R Soc Lond B 183:125–197.
 https://doi.org/10.1098/rstb.1892.0006
- Weber LWD, Boll M, Stampfl A (2003) Hepatotoxicity and mechanism of action of haloalkanes:
 carbon tetrachloride as a toxicological model. Critical Reviews in Toxicology 33:105–
 136. https://doi.org/10.1080/713611034
- Kue J, Shen K, Hu Y, Hu Y, Kumar V, Yang G, Wen C (2020) Effects of dietary *Bacillus cereus*, *B. subtilis, Paracoccus marcusii*, and *Lactobacillus plantarum* supplementation on the
 growth, immune response, antioxidant capacity, and intestinal health of juvenile grass
 carp (*Ctenopharyngodon idellus*). Aquaculture Reports 17:100387.
 https://doi.org/10.1016/j.aqrep.2020.100387
- Yang H, Meng L, Ai D, Hou N, Li H, Shuai X, Peng X (2019) Acetic acid alleviates the
 inflammatory response and liver injury in septic mice by increasing the expression of
 TRIM40. Exp Ther Med 17: 2789-2798. https://doi.org/10.3892/etm.2019.7274
- 665 Yang YM, Seki E (2015) TNFα in liver fibrosis. Curr Pathobiol Rep 3:253–261.
 666 https://doi.org/10.1007/s40139-015-0093-z
- Yao F, Jia R, Huang H, Yu Y, Mei L, Bai L, Ding Y, Zheng P (2019) Effect of *Lactobacillus paracasei* N1115 and fructooligosaccharides in nonalcoholic fatty liver disease. Archives of
 Medical Science 15:1336–1344. https://doi.org/10.5114/aoms.2019.86611
- Yeh Y-H, Hsieh Y-L, Lee Y-T, Hsieh C-H (2011) Protective effects of cholestin against carbon
 tetrachloride-induced hepatotoxicity in rats. e-SPEN, the European e-Journal of
 Clinical Nutrition and Metabolism 6:e264-e271.
 https://doi.org/10.1016/j.eclnm.2011.09.002
- Zamberi NR, Abu N, Mohamed NE, Nordin N, Keong YS, Beh BK, Zakaria ZAB, Nik Abdul
 Rahman NM, Alitheen NB (2016) The antimetastatic and antiangiogenesis effects of
 kefir water on murine breast cancer cells. Integr Cancer Ther 15:NP53–NP66.
 https://doi.org/10.1177/1534735416642862
- Zavala L, Golowczyc MA, van Hoorde K, Medrano M, Huys G, Vandamme P, Abraham AG
 (2016) Selected *Lactobacillus* strains isolated from sugary and milk kefir reduce

- 680 Salmonella infection of epithelial cells in vitro. Beneficial Microbes 7:585–595.
 681 https://doi.org/10.3920/BM2015.0196
- Kato H, Eguchi S, Alam A, Ma D (2017) The role of nuclear factor-erythroid 2 related factor 2
 (Nrf-2) in the protection against lung injury. Am J Physiol Lung Cell Mol Physiol
 312:L155–L162. https://doi.org/10.1152/ajplung.00449.2016

Bandung, September 21, 2023

Dear **Prof. Gabino Garrido** Editor in Chief Journal of Pharmacy & Pharmacognosy Research Garval Editorial Ltda. Antofagasta Chile

Thank you for giving us the opportunity to submit a revised draft of our manuscript (ID: JPPRes-23-1732.R2) titled *Hepatoprotective Study of Indonesian Water Kefir Against CCl4-Induced Liver Injury* in *Rats* to *Journal of Pharmacy & Pharmacognosy Research (JPPRes)*. We appreciate the time and effort that you and the reviewers have dedicated to providing your valuable feedback on our manuscript. We are grateful to the reviewers for their insightful comments on our paper. We have been able to incorporate changes to reflect most of the suggestions provided by the reviewers. We have highlighted the changes within the manuscript.

	Comment	Response
	Please place a comma before the year in all citations	We have revised
	throughout the document. See examples of already	the citations
	arranged citations in the Introduction.	
Fig 4	Insert here the figures corresponding to curcumin.	The image of curcumin has been listed above the letter E
Fig 5	Insert here the figures corresponding to curcumin.	The image of curcumin has been listed above the letter D
References	This section is one of the most important sections of the document. Please pay close attention to it. The style of the References does not meet the requirements of JPPRes. Please write the References according to the Instructions to the Author or the examples: https://jppres.com/jppres/archive/ https://jppres.com/jppres/volume-11-issue-2/ See examples in 1-3 references. Also, all authors (not "et al.") and DOIs must be given. Please check that all references in this section have been cited in the text and vice versa.	We have revised the references referring to the sample.

Here is a point-by-point response to the reviewers' comments and concerns

Thank you, and we look forward to hearing more about our manuscript.

With kind regards,

Widhya Aligita

Faculty of Pharmacy, Bhakti Kencana University, Jl. Soekarno Hatta no 754, Bandung, West Java, Indonesia

E-mail: widhya.aligita@bku.ac.id



DOI: https://doi.org/10.56499/jppres

Original Article

Hepatoprotective study of Indonesian water kefir against CCl₄-induced liver injury in rats

[Estudio hepatoprotector del kéfir de agua indonesio contra la lesión hepática inducida por CCl₄ en ratas]

Widhya Aligita^{1,3*}, Marlia Singgih², Entris Sutrisno³, I Ketut Adnyana^{1*}

¹Department of Pharmacology & Clinical Pharmacy, School of Pharmacy, Bandung Institute of Technology, Bandung, Indonesia.
 ²Department of Pharmacochemistry, School of Pharmacy, Bandung Institute of Technology, Bandung, Indonesia.
 ³Department of Pharmacology & Clinical Pharmacy, Faculty of Pharmacy, Bhakti Kencana University, Bandung, Indonesia.
 *E-mail: widhya.aligita@bku.ac.id; ketut@itb.ac.id

Abstract

Context: Water kefir is a fermented beverage that is typically made in the home by inoculating a sugar-rich solution with a microbial community (water kefir grains). Several studies on the metabolite content and hepatoprotective effects of water kefir have been published, but carbon tetrachloride (CCl₄)-induced acute liver injury has not been studied.

Aims: To evaluate the efficacy of water kefir in vivo against hepatoprotective CCl₄-induced acute liver injury and to in silico investigate metabolites that play an important role in hepatoprotective mechanisms.

Methods: The present study aimed to investigate the hepatoprotective activity of water kefir in an animal model caused by CCl₄. Furthermore, using molecular docking, the metabolites found in water kefir were evaluated for their role in the NF-κB and Nrf2 signaling pathways.

Results: Water kefir significantly and dose-dependently alleviated acute liver injury caused by CCl₄. Water kefir administration at all doses produced results comparable to the positive control (*Curcuma* extract). Molecular docking simulations showed that, compared to Nrf2, the 25 metabolites were more likely to interact with the NF- κ B receptor. Fumaric acid is the strong metabolite that interacts with the NF- κ B receptor with a free energy of binding and an inhibition constant of -6.66 kcal/mol and 13.22 μ M, respectively.

Conclusions: Water kefir administration improved the condition of liver damage, characterized by a decrease in serum levels of AST, ALT, TNF- α , TGF- β , and an improvement in the liver tissue profile. *In silico* evaluation showed that the metabolites in water kefir were able to interact with target proteins in the NF- κ B and Nrf2 pathways. It was concluded that water kefir improves the condition of the liver by reducing the level of necrosis and fibrosis.

Keywords: free radicals; liver diseases; kefir; molecular docking simulation; probiotics.

Resumen

Contexto: El kéfir de agua es una bebida fermentada que se suele elaborar en casa inoculando una solución rica en azúcar con una comunidad microbiana (granos de kéfir de agua). Se han publicado varios estudios sobre el contenido de metabolitos y los efectos hepatoprotectores del kéfir de agua, pero no se ha estudiado la lesión hepática aguda inducida por tetracloruro de carbono (CCl₄).

Objetivos: Evaluar la eficacia del kéfir de agua *in vivo* contra la lesión hepática aguda inducida por CCl₄ hepatoprotectora e investigar in silico los metabolitos que desempeñan un papel importante en los mecanismos hepatoprotectores.

Métodos: El presente estudio tuvo como objetivo investigar la actividad hepatoprotectora del kéfir de agua en un modelo animal causado por CCl₄. Además, mediante docking molecular, se evaluó el papel de los metabolitos presentes en el kéfir de agua en las vías de señalización NF-κB y Nrf2.

Resultados: El kéfir de agua alivió de forma significativa y dependiente de la dosis la lesión hepática aguda causada por CCl₄. La administración de kéfir de agua a todas las dosis produjo resultados comparables a los del control positivo (extracto de cúrcuma). Las simulaciones de acoplamiento molecular mostraron que, en comparación con el Nrf2, los 25 metabolitos eran más propensos a interactuar con el receptor NF-κB. El ácido fumárico es el metabolito fuerte que interacciona con el receptor NF-κB con una energía libre de unión y una constante de inhibición de -6,66 kcal/mol y 13,22 μM, respectivamente.

Conclusiones: La administración de kéfir de agua mejoró el estado de daño hepático, caracterizado por una disminución de los niveles séricos de AST, ALT, TNF- α , TGF- β , y una mejora del perfil tisular hepático. La evaluación *in silico* mostró que los metabolitos del kéfir de agua eran capaces de interactuar con proteínas diana en las vías NF- κ B y Nrf2. Se concluyó que el kéfir de agua mejora el estado del hígado al reducir el nivel de necrosis y fibrosis.

Palabras Clave: enfermedades hepáticas; kéfir; probióticos; radicales libres; simulación de acoplamiento molecular.

ARTICLE INFO Received: June 28, 2023. Accepted: September 21, 2023. Available Online: September xx, 2023. AUTHOR INFO ORCID:

<u>0000-0001-8338-4115</u> (WA) <u>0000-0002-5351-1731</u> (MS) 0000-0003-3830-6411 (ES) 0000-0001-5217-2312 (IKA)

INTRODUCTION

In most cases, making water kefir involves combining dried fruit, sugar, and water kefir grains in a container. Water kefir's exact origins are unknown; however, two hypotheses have been proposed regarding its history: the first suggests that water kefir grains were brought to Europe from the Caucasus by soldiers returning from the Crimean War in the late nineteenth century (Ward, 1892); the second theory proposes that water kefir grains originated in Mexico from the Opuntia cactus through natural processes (Moinas et al., 1980). Sugary kefir grains, Balm of Gilead, African bees, Japanese beer seeds, Ale nuts, and California bees are some other names for water kefir. Tibi grains and ginger beer plants are other names for water kefir (Kebler, 1921; Moinas et al., 1980). Water kefir is appealing to both consumers and researchers due to the variety of microbiota it contains, the fact that it is an alternative to dairy products, the versatility with which it can be flavored, the fact that it is low in calories and sugar, the ease with which it can be produced, and the health benefits it offers.

Water kefir has been used medicinally for a very long time, and recent research has indicated that it may have a variety of positive effects on people's health. It has been demonstrated that water kefir contains non-pathogenic bacteria that, in conjunction with the production of organic acids, can inhibit the growth of pathogenic microbes such as Shigella sp., Salmonella sp., Staphylococcus aureus, and E. coli; as well as, filamentous fungi such as Aspergillus ochraceus, A. niger, A. flavus, Penicillium sp., and Rhizopus sp. (Al-Mohammadi et al., 2021). In addition to its antibacterial properties, water kefir possesses a broad spectrum of pharmacological effects. Some of these therapeutic effects are anti-inflammatory (Aligita et al., 2020; Diniz et al., 2003), antioxidant (Aligita et al., 2020; Darvishzadeh et al., 2021), hepatoprotective (Aligita et al., 2021; Aspiras et al., 2015), antihyperglycemic and antihyperlipidemic (Alsayadi et al., 2014; Rocha-Gomes et al., 2018), anti-edematous (Moreira et al., 2008), antitumor (Zamberi et al., 2016), antihypertensive (Gamba et al., 2019), immunomodulant (Calatayud et al., 2021), and anti-ulcerogenic (Rodrigues et al., 2016). However, no studies have been reported on the hepatoprotective effects of water kefir against carbon tetrachloride (CCl₄)-induced liver injury.

Studies have shown that acute liver injury is frequently accompanied by high levels of oxidative stress and inflammatory responses (Dai et al., 2021). These findings have been found in several studies. The most important signaling pathways that are involved in the regulation of inflammation and antioxidation are the nuclear factor (NF-KB) and nuclear factor erythroid 2-related factor 2 (Nrf2) pathways, respectively. It has been shown that activating Nrf2 and inhibiting NF-KB can reduce the amount of damage done to the liver. For instance, curcumin protects against aflatoxin B1-induced liver injury by increasing the expression of Nrf2 and related downstream antioxidant molecules (such as superoxide dismutase (SOD), catalase (CAT), heme oxygenase 1 (HO-1), and NAD(P)H quinone dehydrogenase 1 (NQO-1) (Wang et al., 2022b). Also, it has been shown that the molecule methoxy eugenol, which comes from nutmeg and Brazilian red propolis, protects the liver both in vitro and in vivo. This may be attributed to the fact that it targets the NF-KB signaling pathway, which has been shown to have anti-inflammatory effects (De Souza Basso et al., 2021).

Ethanol and lactic acid are the primary metabolites found in fermented water kefir, while lower quantities of glycerol, mannitol, and acetic acid can also be found in the beverage. Additionally, a variety of aromatic and volatile compounds are produced, including ethyl acetate, isoamyl acetate, ethyl octanoate, ethyl hexanoate, and ethyl decanoate, among others (compared to their threshold levels) (Laureys and De Vuyst 2014). The chemical constituents of both phytochemicals and secondary metabolites in natural products, including water kefir, are certainly capable of providing various pharmacological effects for the body (Asnawi et al., 2022a; Nursamsiar et al., 2022). However, an *in silico* study to evaluate the metabolite content in water kefir has not been reported yet. Because of its capacity to speed up the process of identifying and optimizing lead compounds, the in silico method has become the front-runner in the race to improve the speed and accuracy of the process of discovering new drugs. This is because the in silico method can identify and optimize lead compounds more quickly. Techniques such as molecular docking and molecular dynamics (MD) were able to directly indicate a small number of compounds that have high affinity and selectivity by analyzing how the ligand and target interact with one another (Febrina et al., 2021).

Water kefir has been used for an extensive period of time and has been recognized for its widespread benefits, especially in Indonesia. However, its level of popularity falls short in comparison to that of milk kefir. Furthermore, there is a scarcity of studies specifically focused on the hepatoprotective activity of water kefir made with Indonesian grains. Therefore, the purpose of this study was to evaluate the hepatoprotective effects of water kefir in CCl₄-induced rats while also investigating the stability interactions of its metabolites within the NF- κ B and Nrf2 receptors using molecular docking studies.

MATERIAL AND METHODS

Materials and reagents

Water kefir grains, sugar, dried fruit, carbon tetrachloride, rats, diagnostic kits for alanine aminotransferase (ALT) (Proline, IFCC mod.), aspartate aminotransferase (AST) (Proline, IFCC mod.), Elisa Kit TNF- α (Bioassay Technology Laboratory), Elisa Kit TGF- β (Bioassay Technology Laboratory). Other chemicals used in this study were of analytical reagent grade.

Experimental sample and reference extract

The water kefir grain was obtained from Yogyakarta, Indonesia. The water kefir solution was produced using a fermentation procedure. The initial stage involved the preparation of 60 g of sugar, 2 g of raisins, 100 g of water kefir grains, and 1 L of mineral distillated water. The sugar and warm distilled water were mixed in a beaker, followed by the addition of water kefir grains and raisins to the resulting sugar solution. The fermentation procedure was conducted over a duration of forty-eight hours at ambient temperature. A small cloth was used to cover the beaker glass. The kefir grain was utilized in future production, while the filtrate was employed for the purpose of evaluation and analysis. (Aligita et al., 2020; 2021)

The rhizome extract of *Curcuma (Curcuma zanthorrhiza* Roxb, (family *Zingiberaceae*) was employed as a reference drug. The utilized product was a standardized herbal medicine with the brand name Tulak, manufactured by the Borobudur Company Herbal Medicine Industry. The primary purpose of Tulak capsules was to support and preserve optimal liver functionality.

Animals and experimental design

Rats (Wistar strain, male, 200-250 g) were maintained on normal pellet food and tap water ad libitum. Four mice in each group were used. All procedures relating to animals and their care conformed to the international guidelines Principles of Laboratory Animal Care (NIH publication no. 85-23, revised 1985) approval with the ethical number 363/UN6.KEP/EC/2020. In order to develop an animal model with liver injury, the rats received CCl₄ (20% in olive oil) at 1.25 mL/kg BW every 2 days via a gavage tube (Yeh et al. 2011). The rats were randomized into five groups after the development of animals with liver injury, which is characterized by a significant increase in serum ALT level, as follows: (1) positive control group, (2) *Curcuma* extract group 100 mg/kg BW, (3) water kefir 15 mL/kg BW group, (4) water kefir 35 mL/kg BW group, and (5) water kefir 50 mL/kg BW group, with the addition (6) negative control group. Each group received group-specific treatment for two weeks, along with the administration of 1.25 mL/kg BW of CCl₄ (20% in olive oil) every three days.

The rats, which had undergone a fasting period of 8–10 hours while being provided with water, had their blood samples collected via the retro-orbital sinus using a hematocrit capillary tube. The blood sample was taken into a microtube and afterwards then to centrifugation. The serum was separated in order to facilitate further measurements (Parasuraman et al., 2010). Serum ALT level, as the main parameter, was measured prior to induction, following induction, and following treatment. Meanwhile, following therapy, serum AST, TNF- α , TGF- β levels, and liver histopathological were evaluated. The ALT, AST, TNF- α , TGF- β levels measurements are conducted in accordance with the protocols outlined in the reagent kit.

After the euthanasia procedure, the liver specimen was promptly immersed in a 10% formalin solution at room temperature for a duration of 24 hours. Subsequently, the tissues were immersed in paraffin, subjected to sectioning, and affixed onto glass microscope slides. The slices underwent staining with hematoxylin and eosin and were afterwards analyzed using light microscopy (Konstantopoulos et al., 2017).

Molecular docking simulation

Molecular docking experiments were done with the PyRx software (Dallakyan and Olson, 2015) to predict how metabolites, which are small-molecule ligands, bind to biological macromolecules. The NCBI PubChem database (https://pubchem.nlm.nih.gov/, accessed on 3 May 2023) was used to derive the threedimensional structure of water kefir metabolites (Patel et al., 2022). Target proteins like NF-KB (PDB code: 1A3Q) and Keap1 (PDB code: 4L7B) were obtained from the RCSB Protein Data Bank (http://www.rcsb.org/, accessed on 03 May 2023). The PyRx program was used to reduce protein, ligand converted to PDBQT (Asnawi et al., 2023), then maximize GRID parameter (Asnawi et al. 2022b) and perform docking study (Febrina et al., 2022). The BIOVIA Discovery Studio 2017 R2 program was used to view the protein and ligand complex and distance (Ischak et al., 2023). The BIOVIA Discovery Studio 2017 R2 tool was also utilized to find protein active sites.



Data represent mean \pm SEM (n = 4). *p<0.05 significantly different result when compared to the positive control group.

Statistical analysis

All of the information is displayed in the form of individual data points as well as the mean along with the standard error of the mean (SEM). The statistical analysis was carried out with the help of Minitab software (version 19.0), and to make comparisons between several different groups, a one-way analysis of variance with Dunnett's post hoc test was utilized. All statistical graphs were created with Microsoft Excel 2019 in their respective versions. The level of significance that were considered to have been reached was p<0.05.

RESULTS

In vivo evaluation of hepatoprotective activity

The serum ALT levels, as the main parameter for the liver damage, were measured prior to and following induction, as well as following treatment (Fig. 1). Meanwhile, after treatment AST, TNF- α , and TGF- β levels were also evaluated. These findings were analyzed using a one-way ANOVA with a 95% confidence level and a two-sided confidence interval. A significant rise in blood ALT levels demonstrated the establishment of an animal model with liver injury, according to statistical analysis, following the administration of CCl₄. When compared to the positive control group, ALT serum levels decreased significantly after two weeks of therapy with curcuma extract or water kefir (p < 0.05). The three doses of water kefir groups demonstrated equivalent activity when curcuma extract was used as the standard treatment, and there was no significant difference between the three doses of water kefir (p>0.05). When compared to the positive control group, AST levels were also reduced dramatically following treatment with curcuma extract or water kefir. TNF-a levels in the water kefir group were significantly lower than in the positive control group at dosages of 35 and 50 mL/kg body weight (p<0.05). Even though there was no statistically significant difference in TGF- β levels, the group that received the treatment demonstrated a decrease in TGF- β levels.

A histological examination of a normal liver group revealed a typical central vein bordered with endothelial cells and normal hepatocytes, resulting in hepatic cords of cells with distinct cell borders and sinusoidal gaps (Fig. 2A-B). The CCl₄-induced group developed centrilobular hepatic necrosis level 4, which was characterized by massive vacuolization and necrosis of epithelial cells in the liver's centrilobular region (blue arrow) (Fig. 2C-D). The groups that received either *Curcuma* extract or water kefir treatment improved in varied necrotic conditions ranging



(A-B) Negative control group; (C-D) positive control group; (E-F) *Curcuma* extract group; (G-H) water kefir 15 mL/kg BW; (I-J) water kefir 30 mL/kg BW; and (K-L water kefir 50 mL/kg BW. 1: central vein; 2: normal liver epithelial cell; 3: portal track; blue arrow: necrosis and liver epithelial cell vacuolization at centrolobular region.

from level 1 (water kefir 50 mL/kg BW) (Fig. 2K-L) to level 2 [*Curcuma* extract (Fig. 2E-F), water kefir 15 mL/kg BW (Fig. 2G-H), and water kefir 30 mL/kg BW (Fig. 2I-J)].

Molecular docking

Molecular docking studies are considered a powerful tool for predicting the potential targets of bioactive molecules. In order to carry out molecular docking simulations, one of the most critical steps is to identify the target active site. If the target protein is crystallized with a native ligand, in many instances, the location of the active site can be established without any difficulty (Li et al., 2019b). However, the NF- κ B (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) proteins do not have a native ligand, so the active site



was determined. Active site prediction in docking is a computational method for predicting the location and orientation of a receptor protein's binding site for a ligand molecule. The active site prediction was based on a protein structural analysis and the identification of amino acid residues that are likely to interact with the ligand. The projected binding site is then utilized as a starting point for molecular docking, a computer method for predicting a ligand molecule's binding affinity and orientation to a receptor protein. The active site prediction for target proteins (Keap1 and NF- κ B) gives the grid box coordinates (x y z) of 17.500880 Å, 62.323000 Å, and 0.973748 Å; and -1.751769 Å, -20.743853 Å, and -29.010438 Å, respectively (Fig. 3).

The docking results indicated that the 25 metabolites could interact with the target proteins (Keap1 and NF- κ B) (Table 1). In general, all metabolites could interact with both NF- κ B receptors (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) (Table 1). The interaction of metabolites with nuclear factor-kappa B receptors (PDB ID 1A3Q) revealed that 13 metabolites possessed free energy for binding that was greater than that of curcumin. On the other hand, there was not a single metabolite that had a free energy binding that was more powerful than that of nrf2 Keap1 (PDB ID 4L7B) (Table 1).

For volatile compounds, 2-phenyl ethanol and benzaldehyde interacted most strongly with the NF- κ B receptor (PDB ID 1A3Q) with almost the same binding energy of -4.19 kcal/mol. As for organic acids, succinic acid, fumaric acid, and citric acid provided nearly the same strong interactions. Bond energy values of fumaric acid, succinic acid, and citric acid were -6.66, -6.24, and -6.25 kcal/mol, respectively. In sugars, glucose provided the strongest interaction

with the NF-κB receptor (PDB ID 1A3Q) where the binding energy was -3.71 kcal/mol. As for the nrf2 Keap1 receptor (PDB ID 4L7B), 2-phenylacetaldehyde from volatile compounds was able to interact most strongly with the Keap1 receptor (PDB ID 4L7B) with a binding energy of -3.46 kcal/mol. As for organic acids, succinic acid provided nearly the same strong interaction with a bond energy value of -3.02 kcal/mol, respectively. In sugars, glucose showed the strongest interaction with the Keap1 receptor (PDB ID 4L7B) where the binding energy was -3.09 kcal/mol.

The theoretical binding modes of the top three metabolites with their target proteins (Keap1 and NF-κB) are shown in Figs. 4 and 5, respectively. The molecular docking results suggested that these metabolites interacted with the Keap1 and NF-KB to form a complex through hydrogen bonds with various residues. The interaction of 2-phenyl ethanol with the active site of NF-KB was formed by two hydrogen bonds and one Pi-Alkyl interaction with amino acid residues of PRO223, LYS252, and LYS252, respectively. The interaction of benzaldehyde with the active site of NFκB was formed by a hydrogen bond with the amino acid residue of LYS252. The interaction of fumaric acid with the active site of NF-KB was formed by six hydrogen bonds with the amino acid residues of LYS221, LYS252, and TYR285. The interaction of glucose with the active site of NF-KB was formed by six hydrogen bonds with the amino acid residues SER220, LYS221, SER226, and LYS252. Although glucose and fumaric acid were able to be formed by six hydrogen bonds, different types of amino acid residues were involved in the interaction, so fumaric acid interacted more strongly with the active site of NF-KB (Fig. 4).

Table 1. The free energy of binding and inhibition constant of water kefir metabolite active site interaction on NF-κB (PDB ID 1A3Q) and Keap1 (PDB ID 4L7B).

		PDB: 1A3Q		PDB: 4L7B	
No.	Metabolites	Free energy of binding, ∆G (kcal/mol)	Inhibition constant, Ki (µM)	Free energy of binding, ∆G (kcal/mol)	Inhibition constant, Ki (uM)
	Volatile compounds	((F7)	((1)
1	2-Methylbutanol	-3.31	3,770	-2.68	10,890
2	3-Methylbutanol	-3.22	4,360	-2.50	14,670
3	2-Phenylethanol	-4.19	851.52	-3.12	5,140
4	Ethyl acetate	-3.19	4,590	-2.43	16,670
5	2-Methylbutanal	-3.26	4,050	-2.35	19,050
6	1-Octanol	-2.65	11,490	-2.11	28,220
7	2-Phenylethyl acetate	-3.99	1,190	-3.33	3,590
8	Benzyl acetate	-4.16	887.66	-3.44	3,020
9	Benzaldehyde	-4.19	853.74	-3.19	4,550
10	Furfuryl acetate	-3.60	2,320	-3.32	3,660
11	Isobutyl acetate	-3.19	4,610	-2.94	7,020
12	Isopentyl acetate	-3.20	4,510	-3.05	5,820
13	2-Phenylacetaldehyde	-4.07	1,030	-3.46	2,930
14	3-Methylbutanal	-3.16	4,850	-2.73	10,020
15	Gluconic acid	-3.11	5,280	-1.63	64,290
	Organic acids				
16	Lactic acid	-4.92	249.28	-2.06	30,680
17	Succinic acid	-6.24	26.68	-3.02	6,140
18	Fumaric acid	-6.66	13.22	-2.45	16,100
19	Citric acid	-6.25	26.18	-1.87	42,440
20	Malic acid	-5.65	72.69	-2.53	13,860
21	Acetic acid	-4.99	219.33	-2.44	16,280
22	Ethanol	-2.39	17,800	-2.08	29,800
	Sugars				
23	Sucrose	-1.40	93,710	-1.33	105,320
24	Glucose	-3.71	1,900	-3.09	5,440
25	Fructose	-3.21	4,400	-2.65	11,440
	Reference compound				
	Curcumin	-3.44	2,990	-4.22	811.36

The interaction of 2-phenylacetaldehyde with the active site of Keap1 was formed by one hydrogen bond and one Pi-Alkyl interaction with amino acid residues of GLN530 and ALA556, respectively. The interaction of succinic acid with the active site of Keap1 was formed by four hydrogen bonds with the amino acid residues of ARG336, HIS575, and PHE577. The interaction of glucose with the active site of Keap1 was formed by six hydrogen bonds with the

amino acid residues LYS551, ARG553, and ASP573. Although glucose can form more hydrogen bonds than 2-phenylacetaldehyde, the different types of amino acid residues involved have not been able to have a significant effect on the binding energy of its interaction with the active site of Keap1 (Fig. 5).

Curcumin (the reference compound) created three hydrogen bonds with the amino acid residues ARG52,





GLU58, and LYS252 to interact with the active site of NF- κ B. Meanwhile, curcumin established three hydrogen bonds with the amino acid residues ARG336, TYR572, and GLY600 to interact with the active site of nrf2 Keap1 (Figs. 4 and 5). Despite the fact that curcumin could create three hydrogen bonds at both the active sites of NF- κ B and nrf2 Keap1, its interaction with nrf2 Keap1 had a lower free energy of binding than phenylethanol, benzaldehyde, and fumaric acid (Table 1). Overall, the metabolites of water kefir prefer to interact with NF- κ B and nrf2 Keap1 receptors. Whereas fumaric acid and 2-phenylacetaldehyde were metabolites that had the strongest interaction with NF- κ B (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) receptors, respectively.

DISCUSSION

Increased liver enzyme production is one of the abnormalities indicating liver damage. This increase in secretion increases the leakage of enzymes into the circulation. Transaminase is a frequent enzyme used to evaluate liver disorders. The transaminases AST and ALT catalyze the transfer of the α -amino group from alanine and aspartic acid to α -ketoglutaric acid. AST is found in the liver, heart, and skeletal muscle, as well as the kidney, brain, pancreas, lung, leukocytes, and erythrocytes. While ALT is predominantly found in the liver, it is also found in low concentrations in other tissues (Lee et al., 2012). Consequently, ALT was used as the principal hepatotoxicity criterion in this study.

Carbon tetrachloride (CCl₄) is a component of the hepatotoxin, which acts after metabolic activation and is extensively employed as a liver-damaging agent. In this study, the administration of CCl₄ to animals caused hepatotoxicity, which was characterized by a significant increase in ALT after several administrations. CCl₄ is metabolized by the enzyme cytochrome p450 (CYP2E1) in the endoplasmic reticulum to a highly reactive trichloromethyl radical. The trichloromethyl radical rapidly reacts with oxygen to form the trichloromethyl peroxyl radical, which rapidly reacts with lipids to form lipid peroxidation products. Free radicals will disrupt the endoplasmic reticulum (ER) and lead to lipid accumulation, decreased protein synthesis, and increased oxidase activity. CCl₄ hepatotoxicity is characterized by hepatocellular necrosis with fat deposition (Ritesh et al., 2015). At the molecular level, administration of CCl₄ can activate tumor necrosis factor (TNF)-a, nitric oxide (NO), and transforming growth factor (TGF)- α and -\(\beta\) in cells, processes that precipitate cell selfdestruction or fibrosis. TNF-a leads to apoptosis, whereas TGF- β leads to fibrosis (Weber et al., 2003).

In terms of its pathophysiological underpinnings, liver illness is linked to a condition known as dysbiosis, which refers to an imbalance in the makeup of the gut microbiota (Laureys and De Vuyst, 2017; Romero-Luna et al., 2020; Zavala et al., 2016). Both qualitative and quantitative changes in the gut microbiome have the potential to affect the composition of products produced by the microbiota, such as short-chain fatty acids and bile acids (Romero-Luna et al., 2020). Qualitative changes include an imbalance between harmful and helpful microbiomes, whereas numeric changes involve changes to the overall microbiota. In addition to these symptoms, intestinal inflammation, disruption of the intestinal barrier, and the transfer of microbial products can all be caused by dysbiosis (Laureys et al., 2018). For this reason, the condition of the gut microbiome is an important factor in the initiation and development of chronic liver disease (Lee et al., 2021). Based on the results of the study, treatment with water kefir for two weeks after the occurrence of liver damage was able to improve the overall condition of the liver, which was marked by a significant decrease in the values of AST, ALT, TNF-a, TGF-β, and significant improvement in liver histology.

Water kefir contains a number of microorganisms that have been linked to health benefits, such as the probiotics L. paracasei and B. cereus (Fijan, 2014). This activity is linked to an increase in antioxidants like glutathione and catalase and a decrease in proinflammatory transcription factors like nuclear Bfactor, lipopolysaccharide, and Toll-like receptor 4 (TLR-4). Improvements in intestinal barrier function and histological integrity were also observed. Increased expression of claudin-1 and occludin-1, two components of tight junctions, occurs simultaneously with the restoration of the p38 MAPK pathway (Fijan, 2014; Yao et al., 2019; Tsai et al., 2020; Ji et al., 2022). Bacillus is a kind of endospore-forming bacterium that can endure extremely cold temperatures and lengthy periods of storage without dying; its spores can even tolerate the acidic environment of the stomach and make it all the way to the small intestine (Elshaghabee et al., 2017). Bacillus cereus has been shown to reduce ALT levels, an indicator of liver healing, in various animal models of liver injury. It protects the liver by reducing inflammation, enhancing the gut flora, and strengthening the tight junctions in the intestines (Kim et al., 2018; Li et al., 2019a; Xue et al., 2020). Also, when Bacillus spores were used first, hepatocyte necrosis and serum levels of ALT, ZO-1, AST, and TAC went down by a lot. This effect is comparable to that of the popular hepatoprotective compound silymarin (Neag et al., 2020).

Beer, vinegar, cider, and cocoa bean masses are just some of the places where Gram-negative acetic acid bacteria have been found. One of the features that sets acetic acid bacteria apart from others is their alkaline-stable lipid membrane (Lynch et al., 2021). Their "oxidative" fermentation metabolism is responsible for the principal metabolic process in these bacteria, the oxidation of ethanol to acetic acid. Important in the food and beverage sector and beyond, fermentation helps mediate the transition of diverse substrates into products. Although lactic acid bacteria have been studied more extensively than acetic acid bacteria (Hong et al., 2021; Semjonovs et al., 2014), various studies have shown promising results concerning the pharmacological effects of acetic acid bacteria, especially as a hepatoprotective agent. Acetic acid, which is the main metabolite of acetic acid bacteria, can lower inflammation and the severity of liver injury in rats with septic shock by increasing the expression of TRIM40. TRIM40 has been shown to minimize liver damage, decrease the synthesis and release of cytokines such as IL-6 and TNF-a, raise the expression of IL-10, improve survival in septic mice, and block the activity of the TLR4 signaling pathway. Acetic acid injection decreased inflammation as well as the production of inflammatory cytokines (Yang et al., 2019). Acetic acid also stimulates the AMPK signaling pathway, which leads to enhanced lipid oxidation and reduced hepatic lipid and body fat deposition (Kondo et al., 2009; Li et al., 2018).

Apart from microorganisms that directly provide hepatoprotective effects, the metabolites produced from these microorganisms also have the potential to be hepatoprotective. Molecular docking is a technique that is utilized in the context of NF-KB and Nrf2 to make predictions regarding the binding affinity and orientation of small-molecule inhibitors to their active sites. The transcription factor known as NF-KB is an essential component in the management of both the immune system and the inflammatory response (Dai et al., 2021). The expression of important inflammatory genes can be inhibited by small-molecule inhibitors that impair the interaction between NF-KB and DNA. These inhibitors have the potential to be used in therapeutic applications. The Nrf2 binding site is Kelchlike ECH-associated protein 1 in the context of nuclear factor erythroid 2-related factor 2, also known as Nrf2 (Keap1) (Jiang et al., 2019; Zhao et al., 2017). Small-molecule inhibitors that disrupt the link between Keap1 and Nrf2 can activate the Nrf2-ARE signaling pathway, which has been demonstrated to have cytoprotective effects. Keap1 is a negative regulator of Nrf2, and this association can be disrupted by small-molecule inhibitors (Zhao et al., 2017).

Binding energy and Ki are important parameters used in molecular docking to evaluate the strength of the interaction between a ligand and a receptor protein. Binding energy is the energy released when a ligand binds to a receptor protein, and it is calculated as the difference between the energy of the bound complex and the energy of the unbound ligand and protein (Meng et al., 2011). Ki, on the other hand, is the dissociation constant of the ligand-receptor complex. It shows how much ligand is needed to bind 50% of the receptor binding sites. Both binding energy and Ki are used to predict the binding affinity and selectivity of a ligand to a receptor protein (Du et al., 2016). Ki, on the other hand, is the dissociation constant of the ligand-receptor complex. It shows how much ligand is needed to bind solve the binding affinity and selectivity of a ligand to a receptor protein (Du et al., 2016). Ki, on the other hand, is the dissociation constant of the ligand-receptor complex. It shows how much ligand is needed to bind 50% of the receptor binding sites (Du et al., 2016).

Based on the results of an in silico study of water kefir metabolite compounds, it was known that fumaric acid and 2-phenylacetaldehyde have the strongest interactions with NF-KB (PDB ID 1A3Q) and nrf2 Keap1 (PDB ID 4L7B) receptors, respectively. Fumaric acid has been studied for its potential as a hepatoprotective agent. Fumaric acid protects rat livers against cadmium toxicity by decreasing oxidative stress and improving hepatic serum. Fumaric acid prevented cadmium-accrued hepatocellular damage and showed the potential to avert hepatic injury against cadmium in rats (Kaur et al., 2020). Fumaric acid esters were found to ameliorate inflammation and oxidative stress in rats, which was associated with less adiposity and ectopic fat accumulation (Šilhavý et al., 2014).

NF-kB and Nrf2 are two transcription factors that play important roles in regulating inflammation and cell survival. While NF-KB is involved in the inflammatory response, and Nrf2 is involved in the antioxidant response (Ganesh Yerra et al., 2013; Wang et al., 2022a). Both transcription factors have been investigated as potential targets for the development of hepatoprotective agents (Gao et al., 2022; Li et al., 2023; Rahman et al., 2021; Wang et al., 2022a). There is evidence of crosstalk between the Nrf2 and NF-kB pathways (Ganesh Yerra et al., 2013; Gao et al., 2022). The Nrf2 pathway inhibits the activation of the NF-κB pathway by increasing antioxidant defenses and HO-1 expression, which efficiently neutralizes ROS and detoxifies (Ganesh Yerra et al., 2013). The crosstalk between Nrf2 and NF-KB could be a new therapeutic target against hepatotoxicity (Gao et al., 2022). Researchers have tried to identify molecule activators of Nrf2 as chemoprevention for ROS-dependent carcinogenesis, while others have focused on identifying Nrf2 inhibitors to increase the sensitivity of cancer cells to chemotherapy (Sharifi-Rad et al., 2023). While NF-KB and Nrf2 are involved in different cellular processes, they have both been investigated as potential targets for the development of hepatoprotective agents. Molecular docking studies have been used to investigate the interaction of potential hepatoprotective agents with these transcription factors. There is also evidence of crosstalk between the Nrf2 and NF- κ B pathways, which could be a new therapeutic target against hepatotoxicity.

In the pathogenesis of hepatocellular damage, liver fibrosis, and HCC, NF-KB plays a crucial role in regulating inflammation and cell death (Luedde and Schwabe, 2011). In response to many stimuli that may pose a threat to the host, NF-KB is activated, setting in motion processes such as inflammation, immunity, wound healing, and pathogen clearance (Luedde and Schwabe, 2011). Pathogen-derived chemicals that activate Toll-Like Receptors (TLRs) include lipopolysaccharide (LPS), viral and bacterial DNA and RNA, and inflammatory cytokines, including tumor necrosis factor (TNF) and interleukins (IL)-1 (Luedde and Schwabe, 2011). When NF-B is activated, a lot of genes with B-binding sites are transcribed. These genes play important roles in controlling inflammation, the immune response, and cell survival. In an NF-kB-dependent manner, lipopolysaccharide (LPS) activates TLR4 on dormant HSCs by reducing BAMBI expression (an inhibitory TGF- β pseudoreceptor) and increasing Kupffer cell chemotaxis. Due to low levels of BAMBI, recruited Kupffer cells secrete TGF-β, which stimulates HSCs unrestrictively. When HSCs have been activated, NF-KB serves a second crucial function by increasing their chances of survival. LPS, Kupffer cell-derived mediators (such as IL-1 and TNF), and angiotensin II, which is produced and acts on HSCs in an autocrine way, all play roles in activating NF-KB in activated hepatic stellate cells. More activated HSCs and extracellular matrix are deposited in the liver as a result of greater HSC activation and survival (Luedde and Schwabe, 2011).

TNF- α and TGF- β can stimulate one another's production, and TNF- α has been shown to influence TGF- β expression in a variety of cells and tissues (Liu et al., 2022). TNF- α is an inflammatory cytokine that contributes to liver inflammation, and chronic liver inflammation results in liver fibrosis. TNF- α exerts its effects on liver fibrosis via multiple mechanisms, including TGF- β signaling activation (Yang and Seki, 2015). Targeting TNF- α and TGF- β signaling pathways may, therefore, have therapeutic potential for treating liver diseases. In regard to hepatoprotective effects, the relationship between TGF- β and TNF- α is complex and not completely understood.

CONCLUSION

This study evaluated the hepatoprotective qualities of Indonesian water kefir in rats with CCl₄induced liver damage. Water kefir administration improved the condition of liver damage, characterized by a decrease in serum levels of AST, ALT, TNF, TGF, and an improvement of liver tissue profile. *In silico* evaluation showed that the metabolites in water kefir were able to interact with target proteins in the NF- κ B and Nrf2 pathways. It was concluded that water kefir improves the condition of the liver by reducing the level of necrosis and fibrosis.

CONFLICT OF INTEREST

The authors declare no conflicts of interest.

ACKNOWLEDGMENTS

This research was funded by the Center of Research and Community Service, Bhakti Kencana University, Bandung, West Java, Republic of Indonesia (052/14.LPPM/PE.I/ LPPM/2021).

REFERENCES

- Aligita W, Alex V, Taaraungan S, Susilawati E (2021) Hepatoprotective activity of water kefir. Int J Biol Pharm Allied Sci 10: 1784–1794. https://doi.org/10.31032/IJBPAS/2021/10.6.5493
- Aligita W, Tarigan PN, Susilawati E (2020) Anti inflammatory and antioxidant activity of kefir water. Int J Biol Pharm Allied Sci 9: 2454–2464. <u>https://doi.org/10.31032/IJBPAS/2020/9.1.4904</u>
- Al-Mohammadi A-R, Ibrahim RA, Moustafa AH, Ismaiel AA, Zeid AA, Enan G (2021) Chemical constitution and antimicrobial activity of kefir fermented beverage. Molecules 26: 2635. <u>https://doi.org/10.3390/molecules26092635</u>
- Alsayadi M, Jawfi YA, Belarbi M, Soualem-Mami Z, Merzouk H, Sari DC, Sabri F, Ghalim M (2014) Evaluation of antihyperglycemic and anti-hyperlipidemic activities of water kefir as probiotic on streptozotocin-induced diabetic wistar rats. J Diabetes Mellit 4: 85–95. <u>https://doi.org/10.4236/jdm.2014.42015</u>
- Asnawi A, Aman LO, Nursamsiar, Yuliantini A, Febrina E (2022a) Molecular docking and molecular dynamic studies: Screening phytochemicals of *Acalypha indica* against BRAF kinase receptors for potential use in melanocytic tumours. Rasayan J Chem 15: 1352-1361. <u>https://doi.org/10.31788/RJC.202</u>2.1526769
- Asnawi A, Febrina E, Aligita W, Kurnia D, Aman LO, Yuliantini A (2022b) Screening of ashitaba (*Angelica keiskei* K.) compounds as potential *Mycobacterium tuberculosis* kasa inhibitors. Int J App Pharm 14: 80–85. <u>https://doi.org/10.22159/ijap.2022.v14s5.13</u>
- Asnawi A, Nedja M, Febrina E, Purwaniati (2023) Prediction of a stable complex of compounds in the ethanol extract of celery leaves (*Apium graveolens* L.) function as a VKORC1 antagonist. Trop J Nat Prod Res 7: 2362–2370. https://doi.org/10.26538/tjnpr/v7i2.10
- Aspiras BEE, Flores RFAC, Pareja MC (2015) Hepatoprotective effect of fermented water kefir on Sprague-Dawley rats (*Rattus norvegicus*) induced with sublethal dose of acetaminophen. Int J Curr Sci 17: 18–28.
- Calatayud M, Börner RA, Ghyselinck J, Verstrepen L, De Medts J, Van den Abbeele P, Boulangé CL, Priour S, Marzorati M, Damak S (2021) Water kefir and derived pasteurized beverages modulate gut microbiota, intestinal permeability and cytokine production *in vitro*. Nutrients 13: 3897. <u>https://doi.org/10.3390/nu13113897</u>

- Dai C, Li H, Wang Y, Tang S, Velkov T, Shen J (2021) Inhibition of oxidative stress and ALOX12 and NF-κB pathways contribute to the protective effect of baicalein on carbon tetrachlorideinduced acute liver injury. Antioxidants 10: 976. <u>https://doi.org/10.3390/antiox10060976</u>
- Dallakyan S, Olson AJ (2015) Small-Molecule Library Screening by Docking with PyRx. In: Hempel JE, Williams CH, Hong CC (eds) Chemical Biology. New York: Springer New York, pp. 243–250. <u>https://doi.org/10.1007/978-1-4939-2269-7_19</u>
- Darvishzadeh P, Orsat V, Martinez JL (2021) Process optimization for development of a novel water kefir drink with high antioxidant activity and potential probiotic properties from Russian olive fruit (*Elaeagnus angustifolia*). Food Bioprocess Technol 14: 248–260. <u>https://doi.org/10.1007/s11947-020-02563-1</u>
- De Souza Basso B, Haute GV, Ortega-Ribera M, Luft C, Antunes GL, Bastos MS, Carlessi LP, Levorse VG, Cassel E, Donadio MVF, Santarém ER, Gracia-Sancho J, Rodrigues De Oliveira J (2021) Methoxyeugenol deactivates hepatic stellate cells and attenuates liver fibrosis and inflammation through a PPAR-y and NF-kB mechanism. J Ethnopharmacol 280: 114433. https://doi.org/10.1016/j.jep.2021.114433
- Diniz RO, Garla LK, Schneedorf JM, Carvalho JCT (2003) Study of anti-inflammatory activity of Tibetan mushroom, a symbiotic culture of bacteria and fungi encapsulated into a polysaccharide matrix. Pharmacol Res 47: 49–52. https://doi.org/10.1016/S1043-6618(02)00240-2
- Du X, Li Y, Xia YL, Ai SM, Liang J, Sang P, Ji XL, Liu SQ (2016) Insights into protein-ligand interactions: Mechanisms, models, and methods. Int J Mol Sci 17: 144. https://doi.org/10.3390/ijms17020144
- Elshaghabee FMF, Rokana N, Gulhane RD, Sharma C, Panwar H (2017) *Bacillus* as potential probiotics: status, concerns, and future perspectives. Front Microbiol 8: 1490. <u>https://doi.org/10.3389/fmicb.2017.01490</u>
- Febrina E, Alamhari RK, Asnawi A, Abdulah R, Lestari K, Levita J, Supratman U (2021) Molecular docking and molecular dynamics studies of *Acalypha indica* L. phytochemical constituents with caspase-3. Int J App Pharm 13: 210–215. <u>https://doi.org/10.22159/ijap.2021.v13s4.43861</u>
- Febrina E, Asnawi A, Abdulah R, Lestari K, Supratman U (2022) Identification of flavonoids from *Acalypha indica* L. (*Euphorbiaceae*) as caspase-3 activators using molecular docking and molecular dynamics. Int J App Pharm 14: 162– 166. https://doi.org/10.22159/ijap.2022.v14s5.34
- Fijan S (2014) Microorganisms with claimed probiotic properties: an overview of recent literature. Int J Environ Res Public Health 11: 4745–4767. https://doi.org/10.3390/ijerph110504745
- Gamba RR, Yamamoto S, Sasaki T, Michihata T, Mahmoud AH, Koyanagi T, Enomoto T (2019) Microbiological and functional characterization of kefir grown in different sugar solutions. Food Sci Technol Res 25: 303–312. https://doi.org/10.3136/fstr.25.303
- Ganesh Yerra V, Negi G, Sharma SS, Kumar A (2013) Potential therapeutic effects of the simultaneous targeting of the Nrf2 and NF-κB pathways in diabetic neuropathy. Redox Biol 1: 394–397. https://doi.org/10.1016/j.redox.2013.07.005
- Gao W, Guo L, Yang Y, Wang Y, Xia S, Gong H, Zhang BK, Yan M (2022) Dissecting the crosstalk between Nrf2 and NF-κB response pathways in drug-induced toxicity. Front Cell Dev Biol 9: 809952. https://doi.org/10.3389/fcell.2021.809952
- Hong Y, Sheng L, Zhong J, Tao X, Zhu W, Ma J, Yan J, Zhao A, Zheng X, Wu G, Li B, Han B, Ding K, Zheng N, Jia W, Li H (2021) Desulfovibrio vulgaris, a potent acetic acid-producing bacterium, attenuates nonalcoholic fatty liver disease in mice. Gut Microbes 13: 1930874. https://doi.org/10.1080/19490976.2021.1930874

- Ischak NI, Aman LO, Hasan H, Kilo AL, Asnawi A (2023) In silico screening of Andrographis paniculata secondary metabolites as anti-diabetes mellitus through PDE9 inhibition. Res Pharm Sci 18: 100–111. <u>https://doi.org/10.4103/1735-5362.363616</u>
- Ji Y, Xie Q, Meng X, Wang W, Li S, Lang X, Zhao C, Yuan Y, Ye H (2022) *Lactobacillus paracasei* improves dietary fatty liver by reducing insulin resistance and inflammation in obese mice model. J Funct Foods 95: 105150. https://doi.org/10.1016/j.jff.2022.105150
- Jiang ZY, Lu MC, You QD (2019) Nuclear factor erythroid 2-related factor 2 (Nrf2) inhibition: An emerging strategy in cancer therapy. J Med Chem 62: 3840–3856. https://doi.org/10.1021/acs.jmedchem.8b01121
- Kaur G, Shivanandappa TB, Kumar M, Kushwah AS (2020) Fumaric acid protect the cadmium-induced hepatotoxicity in rats: Owing to its antioxidant, anti-inflammatory action and aid in recast the liver function. Naunyn-Schmiedeberg's Arch Pharmacol 393: 1911–1920. <u>https://doi.org/10.1007/s00210-020-01900-7</u>
- Kebler LF (1921) California bees. J Am Pharm Assoc 10: 939-943. https://doi.org/10.1002/jps.3080101206
- Kim B, Kwon J, Kim MS, Park H, Ji Y, Holzapfel W, Hyun CK, Aguila MB (2018) Protective effects of *Bacillus* probiotics against high-fat diet-induced metabolic disorders in mice. PLoS One 13: e0210120. https://doi.org/10.1371/journal.pone.0210120
- Kondo T, Kishi M, Fushimi T, Kaga T (2009) Acetic acid upregulates the expression of genes for fatty acid oxidation enzymes in liver to suppress body fat accumulation. J Agric Food Chem 57: 5982–5986. <u>https://doi.org/10.1021/jf900470c</u>
- Konstantopoulos P, Doulamis I, Tzani A, Korou ML, Agapitos E, Vlachos I, Pergialiotis V, Verikokos C, Mastorakos G, Katsilambros N, Perrea D (2017) Metabolic effects of *Crocus* sativus and protective action against non-alcoholic fatty liver disease in diabetic rats. Biom Rep 6: 513–518. https://doi.org/10.3892/br.2017.884
- Laureys D, Aerts M, Vandamme P, De Vuyst L (2018) Oxygen and diverse nutrients influence the water kefir fermentation process. Food Microbiol 73: 351–361. https://doi.org/10.1016/j.fm.2018.02.007
- Laureys D, De Vuyst L (2014) Water kefir as a promising low-sugar probiotic fermented beverage. Arch Public Health 72: P1. https://doi.org/10.1186/2049-3258-72-S1-P1
- Laureys D, De Vuyst L (2017) The water kefir grain inoculum determines the characteristics of the resulting water kefir fermentation process. J Appl Microbiol 122: 719–732. https://doi.org/10.1111/jam.13370
- Lee NY, Shin MJ, Youn GS, Yoon SJ, Choi YR, Kim HS, Gupta H, Han SH, Kim BK, Lee DY, Park TS, Sung H, Kim BY, Suk KT (2021) *Lactobacillus* attenuates progression of nonalcoholic fatty liver disease by lowering cholesterol and steatosis. Clin Mol Hepatol 27: 110–124. https://doi.org/10.3350/cmh.2020.0125
- Lee TH, Kim WR, Poterucha JJ (2012) Evaluation of elevated liver enzymes. Clin Liver Dis 16: 183–198. https://doi.org/10.1016/j.cld.2012.03.006
- Li H, Weng Q, Gong S, Wang J, Huang Y, Li Y, Guo J, Lan T (2023) Kaempferol prevents acetaminophen-induced liver injury by suppressing hepatocyte ferroptosis *via* Nrf2 pathway activation. Food Funct 14: 1884–1896. https://doi.org/10.1039/D2FO02716J
- Li L, He M, Xiao H, Liu X, Wang K, Zhang Y (2018) Acetic acid influences BRL-3A cell lipid metabolism via the AMPK signalling pathway. Cell Physiol Biochem 45: 2021–2030. https://doi.org/10.1159/000487980

- Li YT, Ye JZ, Lv LX, Xu H, Yang LY, Jiang XW, Wu WR, Shi D, Fang DQ, Bian XY, Wang KC, Wang QQ, Xie JJ, Lu YM, Li LJ (2019a) Pretreatment with *Bacillus cereus* preserves against Dgalactosamine-induced liver injury in a rat model. Front Microbiol 10: 1751. <u>https://doi.org/10.3389/fmicb.2019.01751</u>
- Li Z, Huang Y, Wu Y, Chen J, Wu D, Zhan CG, Luo HB (2019b) Absolute binding free energy calculation and design of a subnanomolar inhibitor of phosphodiesterase-10. J Med Chem 62: 2099-2111. https://doi.org/10.1021/acs.jmedchem.8b01763
- Liu Z, Zhang Y, Zhang L, Zhou T, Li YY, Zhou GC, Miao ZM, Shang M, He JP, Ding N, Liu YQ (2022) Duality of interactions between TGF-β and TNF-α during tumor formation. Front Immunol 12: 810286. https://doi.org/10.3389/fimmu.2021.810286
- Luedde T, Schwabe RF (2011) NF-κB in the liver—linking injury, fibrosis and hepatocellular carcinoma. Nat Rev Gastroenterol Hepatol 8: 108-118. https://doi.org/10.1038/nrgastro.2010.213
- Lynch KM, Wilkinson S, Daenen L, Arendt EK (2021) An update on water kefir: Microbiology, composition and production. Int J Food Microbiol 345: 109128. https://doi.org/10.1016/j.ijfoodmicro.2021.109128
- Meng XY, Zhang HX, Mezei M, Cui M (2011) Molecular docking: A powerful approach for structure-based drug discovery. Curr Comput Aided Drug Des 7: 146–157. <u>https://doi.org/10.2174/157340911795677602</u>
- Moinas M, Horisberger M, Bauer H (1980) The structural organization of the Tibi grain as revealed by light, scanning and transmission microscopy. Arch Microbiol 128: 157–161. https://doi.org/10.1007/BF00406153
- Moreira MEC, Santos MHD, Zolini GPP, Wouters ATB, Carvalho JCT, Schneedorf JM (2008) Anti-inflammatory and cicatrizing activities of a carbohydrate fraction isolated from sugary kefir. J Med Food 11: 356–361. <u>https://doi.org/10.1089/jmf.2007.329</u>
- Neag MA, Catinean A, Muntean DM, Pop MR, Bocsan CI, Botan EC, Buzoianu AD (2020) Probiotic *Bacillus* spores protect against acetaminophen induced acute liver injury in rats. Nutrients 12: 632. <u>https://doi.org/10.3390/nu12030632</u>
- Nursamsiar, Nur S, Febrina E, Asnawi A, Syafiie S (2022) Synthesis and inhibitory activity of curculigoside a derivatives as potential anti-diabetic agents with β-cell apoptosis. J Mol Struct 1265: 133292. https://doi.org/10.1016/j.molstruc.2022.133292
- Parasuraman S, Raveendran R, Kesavan (2010) Blood sample collection in small laboratory animals. J Pharmacol Pharmacother 1: 87–93. <u>https://doi.org/10.4103/0976-500X.72350</u>
- Patel SH, Tan JP, Börner RA, Zhang SJ, Priour S, Lima A, Ngom-Bru C, Cotter PD, Duboux S (2022) A temporal view of the water kefir microbiota and flavour attributes. Innov Food Sci Emerg Technol 80: 103084. https://doi.org/10.1016/j.ifset.2022.103084
- Rahman ZU, Al Kury LT, Alattar A, Tan Z, Alshaman R, Malik I, Badshah H, Uddin Z, Khan Khalil AA, Muhammad N, Khan S, Ali A, Shah FA, Li JB, Li S (2021) Carveol a naturallyderived potent and emerging Nrf2 activator protects against acetaminophen-induced hepatotoxicity. Front Pharmacol 11: 621538. <u>https://doi.org/10.3389/fphar.2020.621538</u>
- Ritesh KR, Suganya A, Dileepkumar HV, Rajashekar Y, Shivanandappa T (2015) A single acute hepatotoxic dose of CCl₄ causes oxidative stress in the rat brain. Toxicol Rep 2: 891–895. https://doi.org/10.1016/j.toxrep.2015.05.012
- Rocha-Gomes A, Escobar A, Soares JS, Da Silva AA, Dessimoni-Pinto NA, Riul TR (2018) Chemical composition and hypocholesterolemic effect of milk kefir and water kefir in

Wistar rats. Rev Nutr 31: 137-145. https://doi.org/10.1590/1678-98652018000200001

- Rodrigues KL, Araújo TH, Schneedorf JM, Ferreira CS, Moraes GO, Coimbra RS, Rodrigues MR (2016) A novel beer fermented by kefir enhances anti-inflammatory and anti-ulcerogenic activities found isolated in its constituents. J Funct Foods 21: 58–69. <u>https://doi.org/10.1016/j.jff.2015.11.035</u>
- Romero-Luna HE, Peredo-Lovillo A, Hernández-Mendoza A, Hernández-Sánchez H, Cauich-Sánchez PI, Ribas-Aparicio RM, Dávila-Ortiz G (2020) Probiotic potential of *Lactobacillus paracasei* CT12 isolated from water kefir grains (tibicos). Curr Microbiol 77: 2584–2592. <u>https://doi.org/10.1007/s00284-020-02016-0</u>
- Semjonovs P, Denina I, Linde R (2014) Evaluation of physiological effects of acetic acid bacteria and yeast fermented nonalchocolic beverage consumption in rat model. J Med Sci 14: 147-152. https://doi.org/10.3923/jms.2014.147.152
- Sharifi-Rad J, Seidel V, Izabela M, Monserrat-Mequida M, Sureda A, Ormazabal V, Zuniga FA, Mangalpady SS, Pezzani R, Ydyrys A, Tussupbekova G, Martorell M, Calina D, Cho WC (2023) Phenolic compounds as Nrf2 inhibitors: Potential applications in cancer therapy. Cell Commun Signal 21: 89. <u>https://doi.org/10.1186/s12964-023-01109-0</u>
- Šilhavý J, Zídek V, Mlejnek P, Landa V, Šimáková M, Strnad H, Oliyarnyk O, Škop V, Kazdová L, Kurtz T, Pravenec M, Sookoian SC (2014) Fumaric acid esters can block proinflammatory actions of human CRP and ameliorate metabolic disturbances in transgenic spontaneously hypertensive rats. PLoS One 9: e101906. https://doi.org/10.1371/journal.pone.0101906
- Tsai YS, Lin SW, Chen YL, Chen CC (2020) Effect of probiotics Lactobacillus paracasei GKS6, L. plantarum GKM3, and L. rhamnosus GKLC1 on alleviating alcohol-induced alcoholic liver disease in a mouse model. Nutr Res Pract 14: 299. https://doi.org/10.4162/nrp.2020.14.4.299
- Wang B, Cui S, Mao B, Zhang Q, Tian F, Zhao J, Tang X, Chen W (2022a) Cyanidin alleviated CCl₄-induced acute liver injury by regulating the Nrf2 and NF-kB signaling pathways. Antioxidants 11: 2383. <u>https://doi.org/10.3390/antiox11122383</u>
- Wang Y, Liu F, Liu M, Zhou X, Wang M, Cao K, Jin S, Shan A, Feng X (2022b) Curcumin mitigates aflatoxin B1-induced liver injury via regulating the NLRP3 inflammasome and Nrf2 signaling pathway. Food Chem Toxicol 161: 112823. <u>https://doi.org/10.1016/j.fct.2022.112823</u>
- Ward M (1892) The ginger-beer plant, and the organisms composing it: A contribution to the study of fermentationyeasts and bacteria. Phil Trans R Soc Lond B 183: 125-197. <u>https://doi.org/10.1098/rstb.1892.0006</u>
- Weber LWD, Boll M, Stampfl A (2003) Hepatotoxicity and mechanism of action of haloalkanes: Carbon tetrachloride as a toxicological model. Crit Rev Toxicol 33: 105–136. <u>https://doi.org/10.1080/713611034</u>
- Xue J, Shen K, Hu Y, Hu Y, Kumar V, Yang G, Wen C (2020) Effects of dietary *Bacillus cereus*, *B. subtilis*, *Paracoccus marcusii*, and *Lactobacillus plantarum* supplementation on the growth, immune response, antioxidant capacity, and intestinal health of juvenile grass carp (*Ctenopharyngodon idellus*). Aquac Reports 17: 100387. https://doi.org/10.1016/j.aqrep.2020.100387
- Yang H, Meng L, Ai D, Hou N, Li H, Shuai X, Peng X (2019) Acetic acid alleviates the inflammatory response and liver injury in septic mice by increasing the expression of TRIM40. Exp Ther Med 17: 2789-2798. <u>https://doi.org/10.3892/etm.2019.7274</u>
- Yang YM, Seki E (2015) TNFa in liver fibrosis. Curr Pathobiol Rep 3: 253–261. <u>https://doi.org/10.1007/s40139-015-0093-z</u>

- Yao F, Jia R, Huang H, Yu Y, Mei L, Bai L, Ding Y, Zheng P (2019) Effect of *Lactobacillus paracasei* N1115 and fructooligosaccharides in nonalcoholic fatty liver disease. Arch Med Sci 15: 1336–1344. <u>https://doi.org/10.5114/aoms.2019.86611</u>
- Yeh Y-H, Hsieh Y-L, Lee Y-T, Hsieh C-H (2011) Protective effects of cholestin against carbon tetrachloride-induced hepatotoxicity in rats. E Spen Eur E J Clin Nutr Metab 6: e264–e271. <u>https://doi.org/10.1016/j.eclnm.2011.09.002</u>
- Zamberi NR, Abu N, Mohamed NE, Nordin N, Keong YS, Beh BK, Zakaria ZAB, Nik Abdul Rahman NM, Alitheen NB (2016) The antimetastatic and antiangiogenesis effects of kefir water

on murine breast cancer cells. Integr Cancer Ther 15: NP53-NP66. https://doi.org/10.1177/1534735416642862

- Zavala L, Golowczyc MA, van Hoorde K, Medrano M, Huys G, Vandamme P, Abraham AG (2016) Selected *Lactobacillus* strains isolated from sugary and milk kefir reduce *Salmonella* infection of epithelial cells *in vitro*. Benef Microbes 7: 585–595. <u>https://doi.org/10.3920/BM2015.0196</u>
- Zhao H, Eguchi S, Alam A, Ma D (2017) The role of nuclear factorerythroid 2 related factor 2 (Nrf-2) in the protection against lung injury. Am J Physiol Lung Cell Mol Physiol 312: L155– L162. https://doi.org/10.1152/ajplung.00449.2016

AUTHOR CONTRIBUTION.					
Contribution	Aligita W	Singgih M	Sutrisno E	Adnyana IK	
Concepts or ideas	х	х	х	х	
Design	х	x	х	x	
Definition of intellectual content	х	x	х	x	
Literature search	х			x	
Experimental studies	х			x	
Data acquisition	х			x	
Data analysis	х			x	
Statistical analysis	х			x	
Manuscript preparation	х	х	x	x	
Manuscript editing	х	x	x	x	
Manuscript review	х	x	x	x	

Citation Format: Aligita W, Singgih M, Sutrisno E, Adnyana IK (2023) Hepatoprotective study of Indonesian water kefir against CCl₄-induced liver injury in rats. J Pharm Pharmacogn Res x(x): xxx-xxx. https://doi.org/10.56499/jppres

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Open Access: This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/ licenses/by/4.0/), which permits use, duplication, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license and indicate if changes were made.

AUTHOR CONTRIBUTION: